

METHYL JASMONATE BRINGS ABOUT RESISTANCE AGAINST SALINITY STRESSED TOMATO PLANTS BY ALTERING BIOCHEMICAL AND PHYSIOLOGICAL PROCESSES

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This manuscript mainly defines about the effect of foliar application of methyl jasmonate (C₁₃H₂₀O₃) (MeJA) on physiological and biochemical processes in tomato under both saline and non-saline conditions. Two tomato genotypes Rio Grande (tolerant) and Savera (sensitive) were grown in pots having sand as growth medium. The salinity substantially decreased the physiological and biochemical parameters. Different doses of MeJA (0.0, 10, 20, 30, 40, 50, 60 µM) were applied on both control and salt stressed tomato plants. Methyl Jasmonate MeJA significantly ameliorated the deleterious effects of salinity on tomato plants by inducing the physiological and biochemical resistance. Different parameters responded to MeJA at various extents. Our findings illustrate that all the parameters responded to foliar application of MeJA and it is quite helpful creating physiological and biochemical resistance in salinity stressed tomato plants.

Keywords: Tomato, methyl jasmonate, photosynthesis, transpiration, antioxidants, proline, amino acids.

INTRODUCTION

Abiotic stresses are likely to be common due to climate change being emerging threat of 21st century in past decades to crop productivity. Drought stress heat stress, salinity stress and other abiotic stresses are soaring complexities to crop husbandry. Changing cropping pattern, irrigation with substandard water and soil deterioration finally lead to effect crop productivity (Nawaz *et al.*, 2015). Out of abiotic stresses salt stress has become an acute problem to farm land and productivity. Salt affected soils have large quantities of Na⁺ and Cl⁻ ions that hamper the plant physiological and biochemical processes, ultimately exert abiotic stress to plant (Munns and Tester, 2008). Overall 7% of total earth surface making 800 million hectares of land and 20% of irrigated farmland is exposed to salinity (Aoki *et al.*, 2005; Flowers and Yeo, 1995; Munns and Tester, 2008).

Phytohormones have been reported to cope stress in plants by producing proteins and causing resistance like abscisic acid (Jin *et al.*, 2000), salicylic acid (Poor *et al.*, 2011) and methyl jasmonate (MeJA) (Yoon *et al.*, 2009). MeJA has been reported to attenuate the drastic effects of salinity and water stress in different crop plants like soybean (Anjum *et al.*, 2011; Yoon *et al.*, 2009), barley (Tsonev *et al.*, 1998), strawberry (Wang, 1999), pea (Velitchkova and Fedina, 1998) and broccoli (Del Amor and Cuadra-Crespo, 2011).

Jasmonic acid (JA) and its methyl ester methyl jasmonate (MeJA) collectively termed as jasmonates are reported to cause cell signaling and regulatory phenomenon responsible to affect seed germination, tuberization, senescence, root

growth, reproductive growth and fruit ripening (Creelman and Mulpuri, 2002; Wasternack and Hause, 2002). MeJA under control conditions reduces gaseous exchange parameters by stomatal closure (Suhita *et al.*, 2003; Wang, 1999) facilitated by alkalization of the guard cell (Gehring *et al.*, 1997) but under stress conditions reveals significant improvement in physiological and biochemical processes (Popova *et al.*, 2003).

Tomato is a popular annual vegetable crop consumed fresh, cooked or processed: by various ways into various products. Deleterious effects of salinity on physiology and biochemistry of tomato is well documented (Cuartero and Fernandez-Munoz, 1998; Maggio *et al.*, 2007; Romero-Aranda *et al.*, 2001). In this way this research project was designed to check the possible effects of MeJA on salt stressed tomato plants. One of the key objectives of this study was to optimize the best dose of MeJA that can recover the most from the drastic effects of salinity on tomato.

MATERIALS AND METHODS

Plant material and growth condition: Present study was carried out in Institute of Horticultural Sciences (IHS), University of Agriculture, Faisalabad. Seeds of tomato were obtained from Ayub Agriculture Research Institute (AARI) Faisalabad. Seeds of tomato genotypes Rio Grande and Savera were surface sterilized before sowing with 5% sodium hypochlorite for ten minutes and then rinsed with pure water. Ten seeds were then sown in each sand filled

plastic pot (9 L volume) then thinned to five per pot. Half strength Hoagland solution was used as source of nutrition in the sand. There were three pots in each treatment and each pot represented as one experimental unit. Leaf samples and other parameters were taken at seedling stage (50 days after sowing). Thirty days after sowing the salinity treatments were applied and after one week MeJA treatments were applied as foliar application method.

Determination of gaseous exchange parameters; photosynthesis (A), transpiration (E) and stomatal conductance (gs): For the measurement of physiological attributes such as photosynthesis rate (A), transpiration rate (E) and stomatal conductance (gs) we selected the three young fully developed and healthy leaves. These selected leaves were placed one by one in the chamber of portable apparatus termed as Infra-Red Gas Analyzer (IRGA) (Analytical Development Company, Hoddesdon, England).

Determination of chlorophyll contents: In this study, we calculated the chlorophyll contents by following the method of Arnon (1949) and Davies (1976). Fresh leaves were cut in minute segments and one gram of these minute segments was put into small sterilized plastic bottles, having 80 % acetone solution. These plastic bottles were allowed to stay at room temperature overnight. The extract obtained from the plastic bottles was centrifuged at 14000 x g for 5 minutes and the supernatant thus obtained was taken at absorbance 645, 663 and 480 nm, using double beam Spectrophotometer (Hitachi-120, Japan).

$$\text{Chl a} = [12.7 (\text{OD } 663) - 2.69 (\text{OD } 645)] \times V/1000 \times W$$

$$\text{Chl b} = [22.9 (\text{OD } 645) - 4.68 (\text{OD } 663)] \times V/1000 \times W$$

Where V= Volume of extract and W= Weight of sample

Determination of total free amino acids (TFA) and total soluble protein (TSP) and leaf proline contents: Fresh leaf material of weight 0.5 g was taken and TFA, TSP and leaf proline contents were estimated by the protocol of Hamilton and Van Slyke, (1943), Lowry *et al.* (1951) and Bates *et al.* (1973), respectively.

Assay of catalase and peroxidase activity: Catalase (CAT) and peroxidase (POX) activities were measured by the

procedure of Chance and Maehly (1955) with some alteration. The CAT reaction solution (3 mL) was comprised of 50 mM phosphate buffer (pH 7.0), 5.9 mM H₂O₂ and 0.1 mL of enzyme extract. Changes in absorbance of the reaction solution were recorded after every 20s at 240 nm. One unit CAT activity was specified as an absorbance change of 0.01 units per min. The POX reaction solution (3 mL) was comprised of 50 mM phosphate buffer (pH 5.0), 20 mM guaiacol, 40 mM H₂O₂ and 0.1 mL of enzyme extract. Variations in absorbance of the reaction solution at 470 nm were calculated after every 20 seconds. One unit POX activity was assigned as an absorbance change of 0.01 units per min. The POX activity was determined and expressed as unit min⁻¹g⁻¹FW basis.

Statistical analysis and design: All the treatments were arranged in CRD (completely randomized design). Data recorded were subjected to statistical analysis by analysis of variance technique using STATISTIX computer program. LSD at 5% level of probability was used to compare the individual means.

RESULTS

Gaseous exchange attributes: Photosynthesis rate in both tomato genotypes reduced markedly due to the imposition of salt stress. However, the deleterious effect of salinity was more pronounced on ‘Savera’, as compared to ‘Rio Grande’. Overall, MeJA application resulted in 1.08-fold higher photosynthesis rate in salt tolerant genotype ‘Rio Grande’, as compared to ‘Savera’ under saline as well as non-saline conditions. Foliar spray of MeJA enhanced the photosynthesis rate under salt stressed but reduced under non-stressed plants (Fig. 1). Under non-saline conditions, MeJA caused the decrease in photosynthesis rate for salt tolerant genotype (Rio Grande) but minimum was recorded for savera (8.95 $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$). Overall, exogenous application of 50 micro molar MeJA was more helpful for attaining salt tolerance in tomato.

Transpiration rate was decreased in both tested genotypes

Table 1. Summary of ANOVA table of photosynthesis rate (A), transpiration rate (E), stomatal conductance (gs), chlorophyll contents “a” and “b”, total soluble proteins (TSP), total free amino acid (TFA), catalase (CAT) and peroxidase (POX).

SOV	A	E	gs	Chlorophyll “a”	Chlorophyll “b”	Proline contents	TSP	TFA	CAT	POX
MeJA (M)	***	***	***	***	***	***	***	***	***	***
Salinity (S)	***	***	***	***	***	***	***	***	***	***
Genotypes (G)	***	***	***	***	***	***	***	***	***	***
M×S	NS	NS	NS	***	*	NS	NS	***	NS	NS
M×G	NS	NS	NS	***	NS	NS	NS	NS	NS	NS
S×G	NS	NS	NS	***	NS	NS	NS	NS	NS	NS
M×S×G	NS	NS	NS	***	NS	NS	NS	NS	NS	NS
CV	2.89	11.11	6.55	4.41	8.57	5.98	3.84	5.60	2.11	2.01

***= $P \leq 0.0001$, *= $P \leq 0.01$, NS=Non-significant @ 0.05 LSD, SOV= Source of variation; CV=Coefficient of variance

under salt stressed conditions. The plants grown under non-saline conditions exhibited the highest transpiration rate than those grown under saline conditions. Under saline condition, highest value for transpiration rate was observed in 50 μ M MeJA-treated plants showing about 1.01-fold increase in transpiration, as compared to control plants in saline as well as non-saline conditions, respectively. Furthermore, 'Rio Grande' genotype exhibited significantly about 1.34-fold higher transpiration rate, as compared to 'Savera'. However, under saline as well as non-saline conditions 'Rio Grande' showed 1.40- and 1.29-fold higher transpiration rate, respectively, as compared to 'Savera' genotype (Fig. 2).

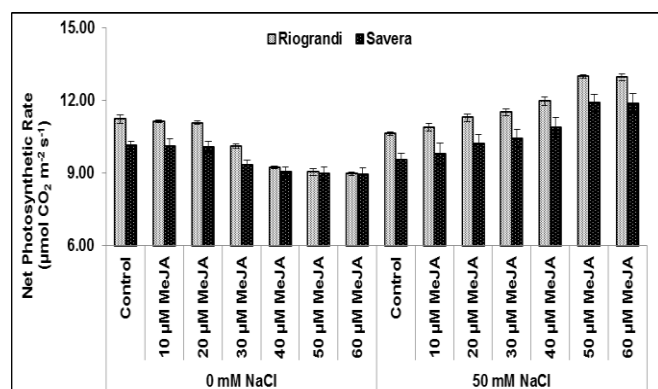


Figure 1. Effect of salinity stress and MeJA on net photosynthetic rate (A). All values are mean of three replications, vertical bars are \pm SE.

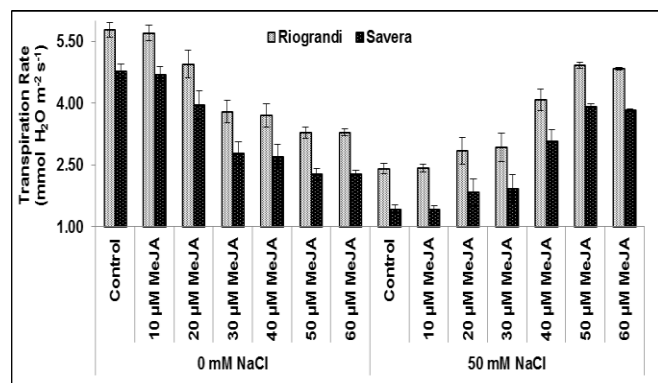


Figure 2. Effect of salinity stress and MeJA on transpiration rate (E). All values are mean of three replications, vertical bars are \pm SE.

Sodium chloride in growth medium reduced the stomatal conductance in both genotypes. Exogenous application of MeJA substantially enhanced stomatal conductance under saline but reduced under non-saline conditions. Mean stomatal conductance values of both genotypes observed in 50 μ M MeJA-treated plants was about 1.18- and 1.19-fold higher than control under saline conditions; whereas, stomatal conductance under non-saline condition was about

1.06-fold higher than saline conditions. Overall, salt tolerant genotype 'Rio Grande' exhibited about 1.11-fold higher stomatal conductance, as compared to 'Savera' (Fig. 3).

Chlorophyll contents: Chlorophyll "a" in both tomato genotypes reduced markedly due to the imposition of salt stress (Fig. 4). Deleterious effect of salinity was more in 'Savera', as compared to that on Rio Grande. Overall, chlorophyll "a" contents were 1.17-fold more in 'Rio Grande', as compared to 'Savera' genotype. Foliar spray of 60 μ M MeJA enhanced chlorophyll "a" contents upto 1.37-fold than control plants under salt stressed conditions. Under saline conditions both tomato genotypes positively responded to the MeJA application; however, 'Rio Grande' showed maximum increase in chlorophyll "b" contents, as compared to 'Savera'. Foliar application of MeJA on tomato plants showed increase in chlorophyll "b" contents under salt stress; whereas, chlorophyll "b" contents were continuously decreased in non-saline conditions, despite MeJA application. Overall, 'Rio Grande' exhibited significantly about 1.36-fold higher chlorophyll "b" contents, than 'Savera' genotype under both soil conditions (Fig. 5).

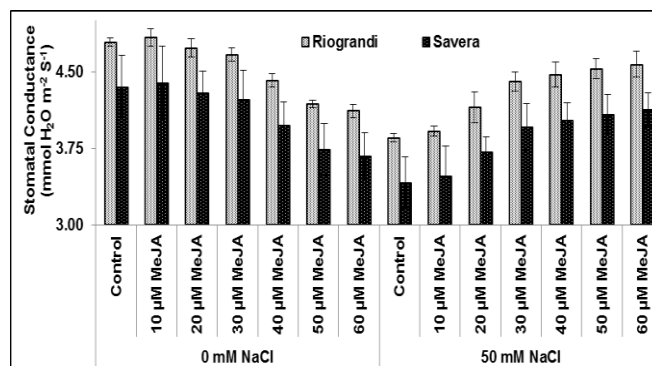


Figure 3. Effect of salinity stress and MeJA on stomatal conductance (gs). All values are mean of three replications, vertical bars are \pm SE.

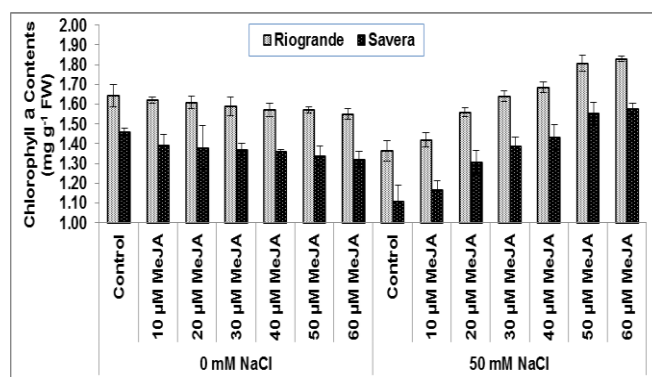


Figure 4. Effect of salinity stress and MeJA on chlorophyll "a" contents. All values are mean of three replications, vertical bars are \pm SE.

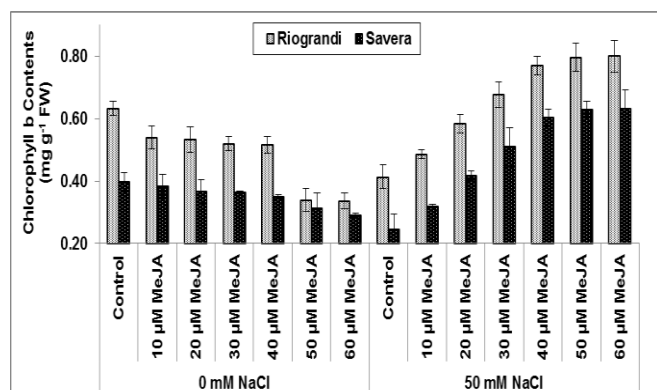


Figure 5. Effect of salinity stress and MeJA on chlorophyll “b” contents. All values are mean of three replications, vertical bars are \pm SE.

Proline contents: Proline contents were significantly higher 2.12-fold under salt stressed conditions due to application of MeJA, as compared to non-saline conditions. Under non-saline conditions, 50 μ M MeJA resulted in 1.68-fold higher proline contents; whereas, under salt stressed conditions 1.26-fold higher proline contents were observed in 50 μ M-treated tomato plants, as compared to control. Overall, salt tolerant genotype ‘Rio Grande’ exhibited 1.18-fold higher proline contents than ‘Savera’ genotype (Fig. 6).

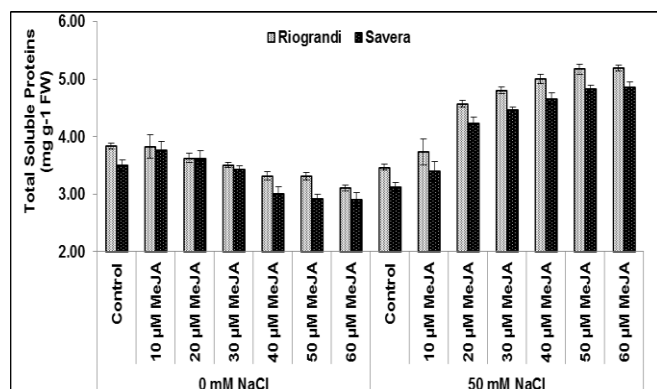


Figure 6. Effect of salinity stress and MeJA on total soluble proteins (TSP). All values are mean of three replications, vertical bars are \pm SE.

Total soluble proteins (TSP): Overall Rio Grande produced 1.07 fold more TSP than Savera both under saline and normal conditions. MeJA revealed a significant effect of TSP, foliar application of MeJA produced more than 66% TSP than control. Under non-saline conditions TSP decreased but under saline conditions TSP increased with MeJA application (Fig. 7).

Total free amino acids (TFA): A distinct change was observed in amino acid contents of the tomato genotypes, investigated under stressed and non-stressed conditions. Application of MeJA enhanced the level of amino acid in

salt affected soil about 3.02-fold higher than non-stressed conditions. Moreover, salt tolerant ‘Rio Grande’ genotype exhibited 1.14-fold higher amino acid contents than ‘Savera’. Exogenous application of 60 μ M MeJA resulted in 1.80- and 2.39-fold higher amino acids than control tomato plants in non-saline as well as saline conditions, respectively (Fig. 8).

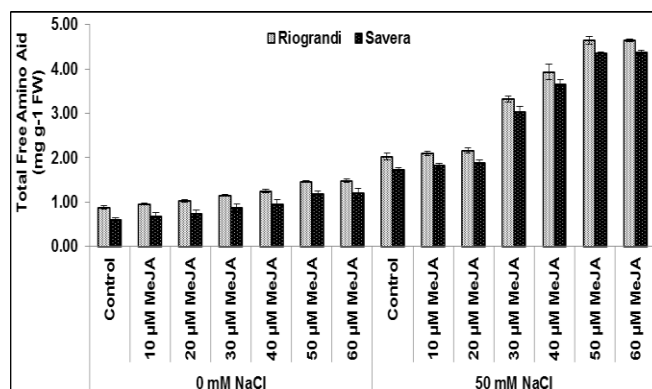


Figure 7. Effect of salinity stress and MeJA on total free amino acids (TFA). All values are mean of three replications, vertical bars are \pm SE.

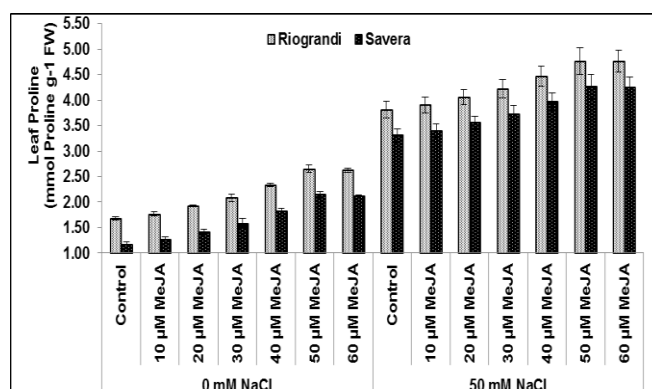


Figure 8. Effect of salinity stress and MeJA on leaf proline contents. All values are mean of three replications, vertical bars are \pm SE.

Antioxidants enzymatic: POX enzyme activity was gradually increased with increasing concentration of MeJA under non-saline and saline conditions. However, increase in POX activity was 1.07-fold more in saline conditions, as compared to non-saline conditions; whereas, salt tolerant genotype ‘Rio Grande’ exhibited about 1.05-fold increase in POX enzyme activity than ‘Savera’. Lowest activity of POX enzyme was observed in control plants, showing about 1.24- and 1.26-fold less POX enzyme activity under non-saline as well as saline condition, respectively, as compared to 60 μ M MeJA-treated tomato plants (Fig. 9).

In present study, salt stress increased the enzymatic activity of catalase (CAT) in the tested genotypes. Salt tolerant

genotype of tomato 'Rio Grande' exhibited significantly about 1.06-fold higher activity than 'Savera'. Whereas; application of MeJA resulted 1.09-fold higher CAT activity in saline conditions, as compared to non-saline conditions. Furthermore, CAT activity was 1.32- and 1.29-fold higher in 60 μ M MeJA-treated tomato plants in non-saline as well as saline condition, as compared to control (Fig. 10).

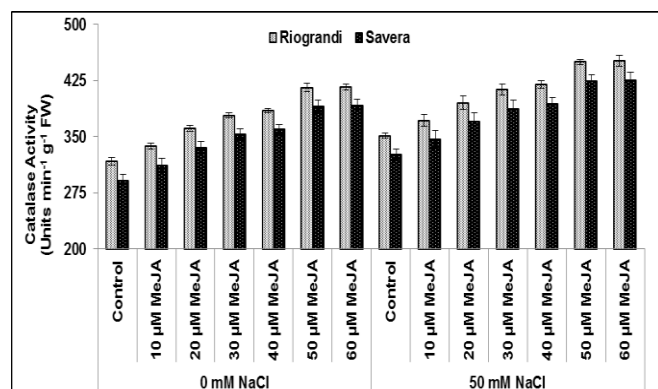


Figure 9. Effect of salinity stress and MeJA on catalase activity (CAT). All values are mean of three replications, vertical bars are \pm SE.

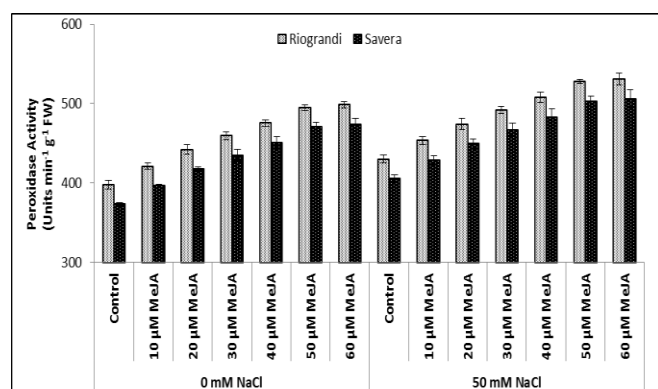


Figure 10. Effect of salinity stress and MeJA on catalase activity (CAT). All values are mean of three replications, vertical bars are \pm SE.

DISCUSSION

Gaseous exchange: Salinity significantly decreased photosynthesis rate (A) and other gaseous exchange parameters by the osmotic effect of salinity that leads to ABA accumulation (Jia *et al.*, 2002) that further leads to stomatal closure thus interrupts stomatal conductance and transpiration (Garcia-Mata and Lamattina, 2002), loss of pigments, activity of Rubisco (Lutts *et al.*, 1996). Under non-saline conditions MeJA application lead to reduction in gaseous exchange parameters but under salt stress conditions MeJA significantly recovered the photosynthesis rate. This could be due to the production of protective protein and

proper working of photosynthesis machinery (Velitchkova and Fedina, 1998; Yoon *et al.*, 2009). MeJA induced enhanced production of chlorophyll contents can be another reason under salt stress conditions for increased photosynthetic rate (Yoon *et al.*, 2009).

Chlorophyll contents, antioxidant enzymes and other biochemical attributes: Chlorophyll is described in the literature to be reduced under salinity stress in different crop plants like wheat (Mehta *et al.*, 2010), pea (Ahmad and Jhon, 2005), rice (Anuradha and Rao, 2003) and tomato (Al-aghabary *et al.*, 2005; Zribi *et al.*, 2009). Reactive oxygen species (ROS) are reported to be produced under stress conditions in different crop plants (Miller *et al.*, 2010; Zhu, 2001). ROS damage the macro-molecules including chlorophyll and some proteins (Tambussi *et al.*, 2000; Zhang *et al.*, 2009). So this can be a solid reason for the deterioration of chlorophyll contents under salinity stress. MeJA is reported to counteract the negative effects of salinity on chlorophyll contents (Fedina *et al.*, 2009; Kang *et al.*, 2005). Under saline conditions proline contents also increased in different crop plants like pea (Velitchkova and Fedina, 1998), soybean (Yoon *et al.*, 2009) and tomato (Claussen, 2005). MeJA application significantly increased free proline contents both under saline and non-saline conditions. Our findings are confirmed by the reports (Abdelgawad *et al.*, 2014; Fedina and Benderliev, 2000; Velitchkova and Fedina, 1998), who depicted an increase in proline contents by MeJA application. Proline contents are reported to be increased under drought stress in pepper and also play a role against oxidative stress (Anjum *et al.*, 2012). TSP and free amino acid contents were dramatically increased by the application MeJA under saline conditions but under normal conditions free amino acids gradually increased but TSP decreased. Our results are in line with Velitchkova and Fedina (1998) as they also recorded the same results in *Pisum sativum*. It is well documented (Abdelgawad *et al.*, 2014; Fedina *et al.*, 2009; Li *et al.*, 1998; Popova *et al.*, 2003) that MeJA augments antioxidants production under stress in different crop plants. In our results MeJA significantly enhanced the production of CAT and POX both under saline and non-saline conditions. Under stress conditions without foliar application of any chemical antioxidants are produced naturally against the response of oxidative stress of ROS (Gill and Tuteja, 2010). Thus in our study overproduction of CAT can be an adaptive mechanism of plants to stressful ecosystem (Nawaz *et al.*, 2015) and MeJA has added in it. Plant species lacking the endogenous production of POX also fail in optimum membrane permeability due to lipid peroxidation, required to cope stress (Bhardwaj *et al.*, 2009; Panda *et al.*, 2003).

Conclusion: A significant increase was observed in enzymatic antioxidants and osmoprotectants by MeJA application under salt stressed tomato. It leads the plants to

better withstand stress conditions by improved water relations and antioxidant system. A significant improvement in gaseous exchange parameters was also noted. There was a non-significant difference between 50 and 60 μ M MeJA application even in some cases 60 μ M performed better but if we optimize the best dose it remained 50 μ M MeJA foliar application for best recovery from salinity stress in tomato plants. Overall Rio Grande performed better than Savera.

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