IMPROVED HORTICULTURAL PRACTICES AGAINST LEAF WILTING, ROOT ROT AND NUTRIENT UPTAKE INMANGO (Mangiferaindica L.)

Muhammad Nafees^{1,*}, Saeed Ahmad², Raheel Anwar², Ishtiaq Ahmad¹, Maryyam² and R.R. Hussnain³

¹University College of Agriculture and Environmental Sciences, The Islamia University of Bahawalpur-63100, Pakistan; ²Institute of Horticultural Sciences, University of Agriculture, Faisalabad-38040, Pakistan; ³Pakistan Science Foundation, Constitution Avenue G-5/2, Islamabad, Pakistan Corresponding author's e.mail: nafeescaes@gmail.com

Poor plant health condition due to various known biotic and abiotic stresses; becoming a disaster in each mango growing country of the world including Pakistan. On the basis of previous researches on the identification of pathogen and several abiotic factors; Soil drenching and foliar spray of various concentrations of Topsin M (TMIC), Aliette (ATP) and Ridomil Gold (ACE) in combination with CuSO₄ (Copper sulphate) was done on mango plants of cv. S.B. (Samar Bahist) Chaunsa showing wilting of leaves and shoots. Foliar application of micro-nutrients (Fe B and Zn) (Iron, Boron and Zinc) was also practiced to improve general health of experimental plants Month-wise emergence of flushes was significantly higher in all treated plants compared with control. Percentage of wilted leaves and root rot in plants, which received drenching and foliar treatments, was significantly reduced (≥50%) compared with untreated plants. Nitrogen, phosphorus and potassium (N, P and K) levels in leaves were significantly improved in treated plants compared with control. Sigmoid relationship was observed between fungicides and copper sulphate concentrations and uptake of N, P and K in treated plants. Application of 250g ATP fungicide by foliar spray plus 125g by soil drench, each along with 50g CuSO₄ proved to be the best against leaf wilting and it improved the N and P level in leaves. While, application of 250g TMIC by foliar spray and 125g by soil drench, each with 50g CuSO₄, was found to be the best to reduce the spread of root rot in experimental plants. Preliminary spray of TMIC along with Copper sulphate is effective to improve plant health of mango cv. S.B. Chounsa.

Keywords: Soil drenching, fungicide spray, copper sulphate, wilting, root rot, mango

INTRODUCTION

Mango (Mangifera indica L.) is an important fruit crop in tropical and sub-tropical climates of the world with an annual production of more than 26 million metric tons on an area of approximately 3.44 million hectare (FAO, 2011). It remained the part of religion and cultural heritage in Indo-Pakistan, and being cultivated in this region of world for the last four thousand years (Candolle, 1984) or even six thousand years (Hills, 1952). It is famous as the king of fruits for its excellent taste and good nutritive qualities (Purseglove, 1972) in Indo-Pak. Per hectare production of mango in Pakistan is quite low (10.54 tons) (Anonymous, 2011) than actual potential of cultivars. This is because of certain diseases which directly lower the production (Purseley, 1993) which was improved up to 28% through protecting crop against diseases (Rawal, 1998; Huang et al., 2012). Since the late nineties, mango decline or dieback disease has become one of the most severe problems in orchard management of the Sindh province (Khanzada et al., 2004b; Khanzada et al., 2004c). Malformation of inflorescence, anthracnose and mango decline (slow and quick) diseases are well known in Pakistan (Nafees et al., 2010; Anwar et al., 2011; Akhtar et al., 2002; Kazmi et al., 2005). In most cases, the disease has been characterized by

the exudation of gum, wilting, dieback, vascular browning and death of the whole tree (Narasimahudu and Reddy, 1992; Khanzada et al., 2004c). Among these, quick decline is more destructive (Sial et al., 2002) and is the combination of several diseases referred as decline disorders (Mahmood and Gill, 2002). Their characteristic symptoms are drying of twigs/branches from top to downward with scratchy appearance (Parkash and Singh, 1976; Ragab et al., 1971), gummosis, bark splitting (Rios-Castano and Reuther, 1968), stem bleeding (Malik et al., 2004). In severe condition, branches start drying one after other in a sequence, resulting in death of the whole plant (Khanzada et al., 2004a). Only pest control work on mango has been done so far (Khan, 1996) and casual organisms are identified by pathologists while disease management through fungicides application on fruit bearing plants is still missing. This is the reason that problem aggravates over the years and even progressive growers have to uproot highly productive mango plants which adversely affect the mango industry of the country (Anonymous, 2011). Keeping in view the above financial crunch on mango industry through decline syndrome, experiments were planned to determine the most effective fungicide among Topsin-M, Aliette and Ridomil Gold with their recommended dose in combination with CuSO₄ and best method of application to minimize the spread of mango

leaf wilting, root rot and to improve plant health by increasing the intensity of new flushes and nutrient uptake.

MATERIALS AND METHODS

Experiment was planned on declining mango plants cv. S.B. Chaunsa of 12 year old, being grown in commercial orchard of uniform sub-tropical agro-climatic conditions (30°18'0N71°56'0E) during 2004-06. Active ingredients in Topsin-M, Aliette and Ridomil Gold is Thiophanate methyl (dimethyl [1-2-phenylene) -bis (iminocarbonotheyl) (TMCI), Aluminiumtris (*O*-ethyl phosphanate (ATP) and Acid methyl ester (ACE), respectively.

	Description
T_1	Water spray only (Control)
T_2	400g TMCI and 100g CuSO ₄ by soil drenching and
	250g TMCI and 100g CuSO ₄ by foliar spray
T_3	250g TMCI and 50g CuSO ₄ by soil drenching and
	125g TMCI and 50g CuSO ₄ by foliar spray
T_4	400g ATP and 100g CuSO ₄ by soil drenching and
	250g ATP and 100g CuSO ₄ by foliar spray
T_5	250g ATP and 50g CuSO ₄ by soil drenching and
	125g ATP and 50g CuSO ₄ by foliar spray
T_6	400g ACE and 100g CuSO ₄ by soil drenching and
	250g ACE and 100g CuSO ₄ by foliar spray
T_7	250g ACE and 50g CuSO ₄ by soil drenching and
	125g ACE and 50g CuSO ₄ by foliar spray

For both soil drenching and foliar spray, chemicals were dissolved in 100 L of water. Soil drenching and foliar spray practices were carried out in April and May respectively during each year. All dry shoots along with 4 inch healthy shoots were removed from experimental plants including control during April by using autoclave sterilized sharp axe and pruning scissors to minimize further spread of disease. Micronutrient mixture of (FeSO₄ 400g +Bordeaux 200g and ZnSO₄ 100g) per 20 L of water was sprayed on all experimental plants including control during the month of May after neutralizing the solution with limestone. Plants were also supplied with 3kg NPK (17:17:17), a synthetic fertilizer of Engro Chemicals, under the plant canopy of all experimental plants including control to improve plant health. Ten terminals of 1.5 to 2 cm thickness were randomly selected on each plant from shoulder height by moving around the plant to record month wise emergence of flushes. Percentage of wilted leaves was recorded after nine months of its emergence. Number of rotten roots, was recorded by digging one square feet hole at six different places under the plant canopy. N, P and K levels in 5-7 month old leaves (Chadha et al., 1986) were analyzed by using the method described by (Chapman and Parker, 1961). Experiment was conducted in a randomized complete block design (RCBD), with seven treatments and three replications

considering whole plant as independent experimental unit. Data was analyzed with software (MSTAT-C, 1989). The least significant differences between treatments were evaluated at α =0.05. Recovery of declining mango plants was assessed on the basis of healthy shoot emergence, percentage of wilted leaves, number of rotten roots and improvement in leaf N, P and K status in treated and control plants.

RESULTS AND DISCUSSION

Effect of fungicides and CuSO₄ soil drenching and foliar spray on vegetative growth: There was significant difference between treated and control plants for April to July flushes, however, LSD value (1.08) for T2 to T7 showed nonsignificant differences for the emergence of April and July flushes (Fig. 1). Significantly higher May flushes recorded in response to low dose of Ridomil Gold (T₇) followed by its high dose (T_6) and least May flushes were recorded in T_2 . Significantly higher June flushes were recorded for high dose of Aliette (T₄) which was at par with high and low dose of Topsin-M (T₂and T₃). Results in this experiment showed that there was no difference between type and dose of fungicide and copper sulphate application for the emergence of April and July flushes. This might be due to reason that we treated experimental plants in April so less time for the emergence of flushes in this month. Significant difference for May and June flushes was due to the reason that percentage of root rot was reduced and efficiency of nutrient uptake was improved. Significantly less emergence of flushes in control plants proved that decline syndrome could be minimized with timely application of proper fungicide in orchard as it was earlier reported by (Ahmed et al., 1995) who evaluated various fungicides against dieback and canopy volume in mango at their recommended doses and reported that three foliar sprays of each of Topsin-M and Benlate at the rate of 1.5 g l⁻¹ and 0.5 g l⁻¹, respectively, were effective in increasing leaf shoot area. Our results also support the findings of Narasimhnudu and Reddy (1992) who showed that maximum disease control with more healthy flushes was achieved by applying 1% Bordeaux mixture followed by 0.8% Bordeaux mixture carbendazim during active growth (spring) season.

Effects of soil drenching and spray of fungicides and $CuSO_4$ on leaf wilting and root rot: Significant difference was recorded between treated and control plants for wilting leaves (Fig. 2). Highly significant wilted leaves (65.44%) (from tip and margins) were recorded in control (T_1) plants followed by low dose of Topsin-M (T_3) and high dose of Ridomil Gold (T_6), respectively. The lowest percentage of wilted leaves was recorded in plants received treatment (T_5) and (T_7) and (T_4) for wilted leaves.

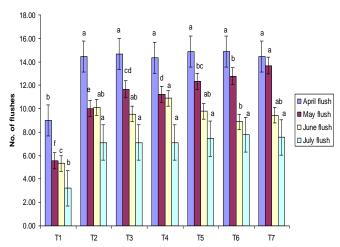


Figure 1. Effect of soil drenching and foliar spray on month wise emergence of flushes. Means with different letters are significantly different at p < 0.05.

Experimental plants showed significant difference in the reduction of root rot as a result of soil drenching and foliar spray of different concentrations of selected fungicides and CuSO₄. Significantly higher number rotten roots (22.11) was recorded in control plants which was followed by 19.78 and 19.33 in plants which received low dose of Aliette (T₅) and high dose of Ridomil Gold (T₆), respectively. Low dose of Topsin-M (T₃) was best with significantly less number of rotten roots (19.22) which was statistically same with high dose of Topsin M (T₂), high dose of Aliette (T₄) and low dose of Ridomil Gold (T₇), respectively. Our results also reveal that wilted/dry leaves were significantly reduced in treated plants compared with control. The highest percentage of wilted leaves was recorded in control (T₁) and least percentage was counted in plants drenched and sprayed with lower doses of Alliete (T_5) and Ridomil Gold (T_7) . These finding are in line with the results of Ahmed et al. (1995) who reported that spray of different fungicides helped in reduction of drying and anthracnose in mango. The results also correlate with the findings of (Narasmnudu and Reddy, 1992) that the rate of leaf wilting can be reduced by using a paste of copper based Bordeaux mixture or Carbendazime as fungicides were copper based in our experiments. Results are in line with the finding of Weinert et al. (1997) who stated that blossom blight and leaf drying could be well controlled with 0.025% Benomyl spray. Root rot was effectively controlled in plants drenched and sprayed with different doses of Topsin-M, Aliette and Ridomil Gold compared with untreated plants. There was maximum spread of root rot in control plants while plants treated with Topsin-M contained minimum number of rotten roots. Intensity of root rot was significantly reduced with continued soil drenching and spraying of fungicides as done in three year

of this study. These findings were supported by several earlier reports (Pegg et al., 1987; Guest et al., 1995; Whiley et al., 1995) which suggests that trunk injection, soil drenching and foliar spray of different fungicides could control *Phytophthora* root rot in avocados and other fruit plants. Risk factors in soil drenching must also be taken into account as Kaiser and Whiley (1998) reported that application of fungicide (Phosphonate) through soil drenching was uneconomical and moreover, drenching may increase the risk for development of pathogen resistance, thus, we used best methods having minimal side effect on plant health.

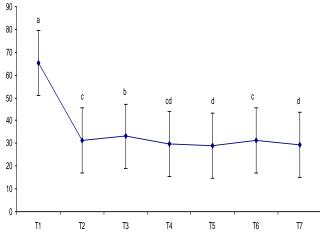


Figure 2. Effect of soil drenching and spray of fungicides and copper sulphate on leaf wilting in mango. Means with different letters are significantly different at p < 0.05.

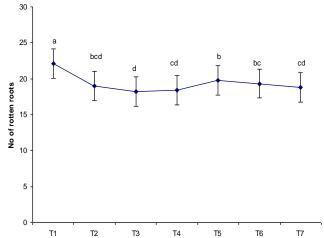


Figure 3. Effect of soil drenching and spray of fungicides and copper sulphate on root rot. Means with different letters are significantly different at p < 0.05.

Effect of soil drenching and spray of fungicides and CuSO₄ on NP and K level in leaves: Nitrogen, phosphorus and potassium concentration was estimated in mature and healthy leaves of treated and control plants. The results are presented in Figure 4. Highly significant effect of soil drenching and foliar spray of fungicides and CuSO₄ was recorded for nitrogen (N) uptake in leaves compared with control. Statistically highest N (1.33%) was recorded in leaves of plants receiving low dose of Aliette (T₅) followed by high and low dose of Ridomil Gold (T₆ and T₇) with 1.27% and 1.26% N, respectively. Statistically significant effect of soil drenching and spray practices was also recorded on phosphorus (P) level in leaves of treated plants compared with control which showed the lowest level of P (0.13%) in leaves. The highly significant level of P (0.17%) was recorded in plants treated with fungicides and CuSO₄ with low dose of Aliette (T5) which was statistically similar with low dose of Topsin-M (T_3) , high dose of Aliette (T_4) and high dose of Ridomil Gold (T₆). Significant effect of treatments was recorded for K uptake in leaves with highest value (0.53%) in leaves of plants receiving low dose of Ridomil Gold (T_7) followed by high dose of Aliette (T_4) , (0.52%). In our research N, P and K level in leaves of treated plants was significantly higher compared with control. This proved that drenching and foliar spray of fungicides and copper sulphate was effective against mango decline by improving plant health. Our results are in accordance with the findings of Mahmood et al. (2002) who recorded that two sprays of Topsin-M @ 1g l⁻¹ effectively reduced mango decline and improved plant nutrient level in leaves. Similar results were reported by Li et al. (1995) by working on dieback of Japanese apricot. The results support to the findings of Werner (1993) who stated that application of paclobutrazol improved the level of several nutrient contents of the leaves with increase N, Ca, Mn, Zn. Our results proved that combination of soil drenching and foliar spray of fungicides plus copper sulphate improved plant health by reducing leaf wilting, root rot and increased the intensity of new flushes and nutrient uptake. This was supported by Rawal (1998) who observed that dieback of mango could be controlled by the spray of 0.1% Carbendazim, 0.1% Methyl ethiophanate or 0.2% Chlorothalonil at fortnightly intervals. Lonsdale and Kotze (1993) also confirmed our findings that these broad-spectrum systemic fungicides (Topsin M, Aliette and Ridomil Gold) were beneficial for the control of mango die-back disease. The results in this experiment were also in line with the work of Parkash and Raoof (1989) that effectively controlled mango decline by pruning the affected portions and spraying with 5:5:50 Bordeaux mixture. Similarly Khanzada et al. (2005) stated that proper sanitation, irrigation and fertilization with at least three fortnightly sprays of copper base fungicides Carbendazim could control mango decline under field conditions. Findings from this experiment may play a significant role in

management of mango decline where poor orchard management practices such as improper irrigation, intercropping, root injuries by deep ploughing and presence of infested plants have been reported to be predisposing factors of decline malady (Saeed *et al.*, 2007).

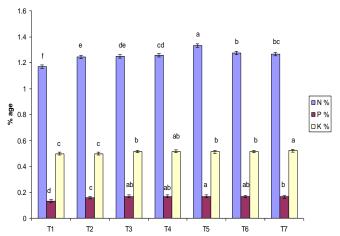


Figure 4. Effect of soil drenching and spray of fungicides on NP & K uptake in Mango. Means with different letters are significantly different at p<0.05.

Conclusion: Poor plant health is leading as a major factor in reducing production and causing huge economic losses to mango industry of the country. On the basis of research worked by pathologists on mango decline, soil drenching and foliar spray of copper-based systemic fungicide could be helpful to reduce spread of leaf wilting, root rot and improve plant health in mango if these practices are started at early stage of decline intensity. Our findings suggest that soil drenching of 250g of any fungicide (Aliette, Topsin M or Ridomil Gold) and 50g of CuSO₄ along with foliar application of 125g of the respective fungicides with 50g of CuSO₄ was the best to improve plant health including leaf wilting, root rot and improving leaf nutrient status in mango cv. S.B. Chounsa.

REFERENCES

Ahmed, I., A., Mahmood, K. Majeed and A. Saleem. 1995. Evaluation of various Fungicides against die-back disease caused by *Diplodianatalensis*in mango. Pak. J. Phytopathol. 7:208-209.

Akhtar, K.P. and S. Alam. 2002. Assessment keys for some important diseases of mango. Pak. J. Biol. Sci. 5:246-250.

Anonymous. 2011. FAO production year book. Food and Agriculture Organization of the United Nations, Rome.

- Anonymous. 2011. Pakistan Statistical Year Book. Federal Bureau of Statistical Year Book. Statistical Division, Govt. of Pakistan.
- Anwar, R., S. Ahmad, I.A. Rajwana, A.S. Khan, N. Memon and M. Nafees. 2011. Phenological growth patterns and floral malformation of mango (*Mangiferaindica* L.) tree under subtropical climate. Pak. J. Agri. Sci. 48:109-115.
- Candolle, D.E. 1984. Origin of cultivated plants, chap. II. K. Paul publisher company London. Chapman, H.D. and F. Parker. 1961. Determination of NPK method of analysis for soil, plant and waters. Pvt. Div. Agri. Univ. California, USA. pp.150-179.
- Chadha, K.L. and R.N. Pal. 1986. *Mangifera indica*. In: A.H. Halevy (ed.), CRC Handbook of flowering. CRC Press Inc., Florida, pp.211-230.
- FAO. 2011. FAO Stat. Food and Agricultural Organization of the United Nation. Available online with updates at http://www.fao.org
- Guest, D.I., K.G. Pegg and A.W. Whiley. 1995. Control of *Phytophthora* diseases of tree crops using trunk-injected phosphonates. Hort. Rev. 17:299-330.
- Hills, A.F. 1952. Economics Botany. McGraw Hill, New York.
- Huang, S., G. Zhu, L. Qin, X. Zhou, F. Huang, Q. Li, W. Yan, H. Huang, Z. Cen, G. Fu and C. Hu. 2012. Enhancement of efficacy in controlling postharvest decays and extending shelf life of mangoes by combined pre- and post-harvest chemical applications. Int. J. Agric. Biol. 14:176-182.
- Kaiser, C. and A.W. Whiley. 1998. Effects of phosphonate soil drenching on avocado (*Perseaamericana*Mill.) root phosphonate concentration. Talking Avocados 9:15.
- Kazmi, M.R., F.S. Fateh, K. Majeed, A.M. Kashkhely, I.Hussain, I. Ahmad and A. Jabeen. 2005. Incidence and etiology of mango sudden death phenomenon in Pakistan. Pak. J. Phytopathol. 17:154-158.
- Khan, I.A., M.A. Kan, M. Asif, M.J. Jaskani, K. Daud, N.H. Labar, M.R. Kazmi, and L.A. Khan. 1996. National coordinated research project on mango: First annual report. Deptt. Of Horti., Uni. of Agri. Faisalabad, Pakistn. p.240.
- Khanzada, M.A., A.M. Lodhi and S. Shahzad.2004a. Mango dieback and gummosis in Sindh, Pakistan caused by *Lasiodiplodiatheobromae*. Plant Health Progress doi:10.1094/PHP-2004-0302-01-DG.
- Khanzada, M.A., A.M. Lodhi and S. Shahzad.2004b. Pathogenicity of *Lasiodiplodiatheobromae* and *Fusariumsolani* on mango. Pak. J. Bot. 36:181-189.
- Khanzada, M.A., A.M. Lodhi and S. Shahzad.2004c. Decline and gummosis diseases of mango in Sindh caused by *Lasiodiplodiatheobromae*.Plant Health Progress: 10.1094/PHP- 204-0302-01-DG.

- Khanzada, M.A., M. Lodhi and S. Shahzad. 2005. Chemical control of *lasiodiplodiatheobromae*, the causal agent of mango Decline in Sindh. Pak. J. Bot. 37:1023-1030.
- Li, H.Y., R.B. Cao and Y.T. Mu. 1995. *In vitro* inhibition of *Botryospheriadothideae* and *Lasiodiplodiatheobromae*, and chemical control of gummosis diseases of Japanese apricot and peach plants in Zhejiang province, China. Crop Prot. 14:187-191.
- Lonsdale, J.H. and J.M. Kotze. 1993. Chemical control of mango blossom disease and the effect on fruit set and yield. Plant Dis. 77: 558-562.
- Mahmood, A. and M.A. Gill. 2002. Quick decline of mango and in vitro response of fungicides against the disease. Int. J. Agri. Biol. 4:39-40.
- Mahmood, A., A. Saleem and K.M. Akhtar. 2002. Mango decline in Pakistan and its management. Pak. J. Phytopathol.14:40-43.
- Malik, M.T., A.A. Dasti and S.M. Khan. 2004. Some manageable predisposing factors of collar/stem rot of mango. Pak. J. Phytoipathol. 16:37-42.
- MSTAT. 1989. MSTAT user' guide: A microcomputer program for the design management and analysis of agronomic research experiments. Michigan State Univ. East Lansing, USA.
- Nafees, M., R. Anwar, M. Jameel, M.N. Aslam, S. Ahmad, F.Z. Akhtar and N. Memon. 2010. Flushing pattern of mango (*Mangiferaindica* L.) cultivars in response to pruning of panicles and its effect on carry over effect of floral malformation. Pak. J. Agric. Sci. 47:13-18.
- Narasimhnudu, Y. and P.S.N. Reddy. 1992. A note on gummosis of mango. Indian Phytopathol.45: 261-262.
- Parkash, O. and M.A. Raoof. 1989. Dieback disease of mango (*MangiferaindicaL.*), its distribution, incidence, cause and management. Phytopathologia Brasiliensis 14: 207-215.
- Parkash, O. and U.N. Singh. 1976. New disease of mango. Proc. of Fruit Research Workshop, Hyderabad, India. 24-28 May. pp.300-302.
- Pegg, K.G., A.W. Whiley, P.W. Langdon and J.B. Saranah. 1987. Comparison of phosetyl-Al, phosphorous acid and metalaxyl for the long-term control of Phytophthora root rot of avocado. Aust. J. Experi. Agri. 27: 471-474.
- Purseglove, J.W. 1972. Mangoes West of India. Acta Hortic. 24:107-174.
- Purseley, D. 1993. Disease of fruit corps.Deptt.of Primary Industries, Brisbane, Queensland, Australia.
- Ragab, M.M., K.A. Sabet and N.A. Dawood. 1971. *Botrydiplodiatheobromae* Pat. The cause of fruit rote and dieback of mango in A.R.E. Agricultural Research Review, Cairo 49: 81-97.
- Rawal, R.D. 1998. Management of fungal diseases in tropical fruits. In: R.K. Arora and V.R. Rao (eds.),Tropical. Fruits in Asia: Diversity, Maintenance,Conservation and Use. Proc. of the IPGRI-

- ICARUTFANET regional training course on the conservation and use of germplasm of tropical fruits in Asia held at Indian Institute of Horticultural Research, 18-3J May 1997, Bangalore, India.
- Rios-Castano, D. and W. Reuther. 1968. Bark cracking of mango trunk. Proc. of Tropical Research. J. Amer.Soc. Hort. Sci. 11:16-22.
- Saeed, S., N. Hussain and R. Attique. 2007. Etiology and management of sudden death phenomenon in mango. Second Annual Report.Deptt.ofEntomolo. Uni.College of Agri. BahuddinZakariya Uni., Multan.pp.12-40.
- Sial, A.K. 2002. Mango: a fruit for the world market. Business of Finance Review, the News.10th April, 2002, Lahore, Pakistan.
- Weinert, M.P. A. Drenth, S.H. Soo, J.A.G. Irwin and K.G. Pegg. 1997. Different phosphorous acid sensitivity levels in *Phytophthoracinnamomi* isolates from treated and untreated avocado trees. In: Proc. of Australasian Plant Pathology Society, 11 Biennial Conference. p.35.
- Werner, H. 1993. Influence of paclobutrazol on growth and leaf nutrient content of mango (cv. Blanco). Acta Hort. 341:225.
- Whiley, A.W., P.A. Hargreaves, K.G. Pegg, V.J. Doogan, L.J. Ruddle, J.B. Saranah and P.W. Langdon. 1995. Changing sink strengths influence translocation of phosphonate in avocado (*Perseaamericana*Mill.) trees. Aust. J. Agri. Res. 46:1079-90.