# SALINITY TOLERANCE POTENTIAL OF TWO ACACIA SPECIES AT EARLY SEEDLING STAGE

Ghulam Abbas<sup>1,\*</sup> Muhammad Saqib<sup>1</sup>, Javaid Akhtar<sup>1</sup> and S.M.A. Basra<sup>2</sup>

<sup>1</sup>Institute of Soil and Environmental Sciences, University of Agriculture, Faisalabad, Pakistan; <sup>2</sup>Department of Crop Physiology, University of Agriculture, Faisalabad, Pakistan \*Corresponding author's e-mail: g.a92pk@gmail.com

Soil salinity is a major environmental issue in arid and semiarid regions of the world. Acacia has very important role for salt affected barren lands due to its high salinity tolerance potential. The aim of the present study was to explore the genetic differences among *Acacia ampliceps and Acacia nilotica* regarding their response to salinity. Three-weeks old seedlings of both species were transplanted in half strength Hoagland nutrient solution having five salt levels (control,100, 200, 300 and 400 mM NaCl) with four replications in completely randomized design with factorial arrangement. After eight weeks of transplantation, the plants were harvested and data for shoot and root length and their fresh and dry weights were recorded. Na<sup>+</sup>, K<sup>+</sup> and Cl<sup>-</sup>concentration of both root and shoot was determined. All the growth parameters of both species declined significantly in response to salinity. *Acacia ampliceps* was more salt tolerant than *Acacia nilotica* with better growth owing to higher K<sup>+</sup>: Na<sup>+</sup> ratio in plant tissues.

**Keywords:** Salinity, NaCl, K<sup>+</sup>: Na<sup>+</sup> ratio, acacia species, ionic composition

#### INTRODUCTION

Yield of many crops particularly for arid and semi-arid areas of the world is reduced due to salinity ((Majeed *et al.*, 2010; Murtaza *et al.*, 2011). About, 7% of the global land area and approximately 20% global cultivated land is affected by salinity (Zhu, 2001). About 6.67 m ha, approximately one third of the total cultivated area in Pakistan is having the problem of soil salinization (Khan, 1998).

The growth of most of the plants is badly affected due to salinity (Taiz and Zeiger, 2006) and if the level of salinity is too high the growing plants may be eliminated totally (Garg and Gupta, 1997). The provision of all the nutrients to plant is made through roots so the reaction of roots to salinity is very vital for understanding the whole plant response to salinity (Ahmed et al., 2012). Plant cells, tissues and organs display varied response to harsh environmental conditions at different levels of their growth (Munns, 1993). High cytoplasmic solute concentrations cannot be tolerated by most of the plants, so in order to carry on routine metabolic functions, plants have to dump higher concentrations of salts in their vacuoles or they have to restrict them in older tissues (Munns and Tester, 2008). Potassium ion is very vital for many cell functions like cell enlargement, osmoregulation and homeostasis (Schachtman et al., 1997). The plants with higher K<sup>+</sup>: Na<sup>+</sup> ratio are considered better tolerant to salinity stress (Saqib et al., 2005). Despite decline in growth due to salinity, there is marked genotypic difference within plant species regarding their sensitivity to higher concentrations of salts (Saqib et al., 2006).

According to Saboora *et al.* (2006), it is expected that salinity will increase at a rate of about 10% per year around the globe. Grassland cover and ultimately the feed production for animals is reduced due to salinization in the areas of arid and semiarid climate (El-kharbotly *et al.*, 2003). There is a great demand for such plant species which are naturally gifted with high salinity tolerance potential and at the same time can give reasonable economic yield from such barren lands.

Acacia species have the ability to survive in a varied range of territories and environments and are well acclimatized to the semi-arid and savannah type of climates and are tolerant to high pH and waterlogging (Benninson and Paterson, 1993). This genus is highly tolerant to salinity as well. Yokota (2003) compared five acacia species, viz A. ampliceps, A. salicina, A. ligulata, A. holosericea and A. mangium in solution culture for their salt tolerance and found that A. ampliceps was more tolerant and survived even at 428 mM NaCl where all the other species died. Marcar et al. (1991) found that some acacia species have the potential to grow up to 400 mM NaCl concentration. The objective of this study was to evaluate the salinity tolerance potential of two acacia species for their growth and ionic relationship, so that the most promising specie can be recommended for saltaffected lands.

### MATERIALS AND METHODS

Seeds of two acacia species (A. nilotica and A. ampliceps) were collected from Forest Research Institute, Peshawar, Pakistan and Nuclear Institute for Agriculture and Biology

(NIAB), Faisalabad, Pakistan. The seeds were sown in iron trays having two inch layer of sand washed with distilled water. The sand was kept moistened with water and with nutrient solution after seedling emergence. After three weeks, the seedlings were transplanted in foam plugged holes in polystyrene sheets floating over nutrient solution in 25L iron tubs.

After one week of transplantation, different levels of salinity (100, 200, 300 and 400 mM NaCl) were developed by the addition of calculated amount of NaCl in two increments (one per day) in the salinity treatment tubs, whereas no salt was added in control. The pH of the solution was maintained at 5.5±1 with dilute NaOH or HCl and the solution was changed weekly during the period of study. After eight weeks, the plants were harvested and the data regarding shoot and root length and fresh weights were recorded. Oven dry weight of shoot and root was recorded after drying the samples at 75°C for 48 hours. The shoot and root samples were ground in a grinding machine and digested with H<sub>2</sub>SO<sub>4</sub> and H<sub>2</sub>O<sub>2</sub> following the method of Wolf (1982) and were analyzed for ions. Na+ and K+ were determined by Sherwood- 410 Flame Photometer and Cl- was determined with Sherwood- 926 chloride analyzer. Data were subjected to statistical analysis (Steel et al., 1997) and significance of differences among treatments and species was determined using LSD test.

#### **RESULTS**

Shoot and root growth: The increasing levels of salinity resulted in reduction in shoot fresh and dry weights of both acacia species. The differences among species, treatments and their interaction were significant for both these parameters (Table 1). A. ampliceps survived even at 400 mM NaCl whereas A. nilotica could not survive at this salinity level (data not shown). In control, both species did not differ significantly; however, in the salt treatments they differed significantly from each other and this difference widened with increasing salt concentration in the growing medium. Salinity resulted in decline in the root growth as well. The main effects as well as their interaction were significant for both root fresh and dry weights. Reduction in fresh and dry weights of root was more in case of A. nilotica than A. ampliceps (Table 1). The length of both shoot and root also decreased due to salinity stress. Although treatments had significant effect; the species did not differ significantly from each other (Table 1). A. nilotica had higher root and shoot length up to 200 mM NaCl level. However, in 300 mM NaCl treatment, A. ampliceps had higher length of both root and shoot.

**Shoot and root ionic composition:** Salinity significantly increased the shoot and root Na<sup>+</sup> concentration in both species. (Table 2). The individual as well as interactive

Table 1. Effect of salinity on growth parameters of acacia species

	Control	100 mM NaCl	200 mM NaCl	300 mM NaCl	Mean
A. Shoot fresh weig	ht (g plant <sup>-1</sup> )				
A. ampliceps	3.91 a	3.40 b	2.52 d	1.25 f	2.77 A
A. nilotica	3.74 a	3.00 c	1.87 e	0.50 g	2.28 B
Mean	3.83 A	3.20 B	2.19 C	0.88 D	
B. Shoot dry weight	(g plant <sup>-1</sup> )				
A. ampliceps	0.74 a	0.63 b	0.49 d	0.24 f	0.53 A
A. nilotica	0.71a	0.55c	0.36 e	0.11 g	0.43 B
Mean	0.73 A	0.59 B	0.43 C	0.18 D	
C. Root fresh weigh	nt (g plant <sup>-1</sup> )				
A. ampliceps	1.78 a	1.60 b	1.20 d	0.65f	1.31 A
A. nilotica	1.74 a	1.44 c	0.95 e	0.33 g	1.12 B
Mean	1.76 A	1.52 B	1.08 C	0.49 D	
D. Root dry weight (	(g plant <sup>-1</sup> )				
A. ampliceps	0.31 a	0.28 b	0.22 d	0.12f	0.23 A
A. nilotica	0.30 a	0.24 c	0.17e	$0.06 \; \mathrm{g}$	0.19 B
Mean	0.31 A	0.26 B	0.20 C	0.09 D	
E. Shoot length (cm)	)				
A. ampliceps	30.0 b	26.5 c	18.25 d	9.50 e	21.06 A
A. nilotica	34.0 a	28.5 bc	19.0 d	5.60 f	21.78 A
Mean	32.0 A	27.5 B	18.63 C	7.55 D	
F. Root length (cm)					
A. ampliceps	24.0 b	21.5 c	16.0 d	8.10 e	17.4 A
A. nilotica	27.0 a	23.1 bc	16.5 d	4.70 f	17.83 A
Mean	25.5 A	22.3 B	16.3 C	6.40 D	

Means of a parameter sharing the same small letters are statistically similar at  $P \le 0.05$ .

Table 2. Effect of salinity on ionic composition of acacia species

	Control	100 mM NaCl	200 mM NaCl	300 mM NaCl	Mean
A. Shoot Na <sup>+</sup> co	oncentration (mmol g <sup>-1</sup>	dw)			
A. ampliceps	0.15 f	0.51 e	0.91 d	1.72 b	0.82 B
A. nilotica	0.15 f	0.76 d	1.30 c	2.30 a	1.13 A
Mean	0.15 D	0.64 C	1.11 B	2.01 A	
B. Root Na <sup>+</sup> cor	ncentration (mmol g <sup>-1</sup>	lw)			
A. ampliceps	0.19 g	0.65 f	1.30 d	2.10 b	1.06 B
A. nilotica	0.19 g	0.90 e	1.91 c	2.86 a	1.47 A
Mean	0.19 D	0.78 C	1.61 B	2.48 A	
C. Shoot Cl <sup>-</sup> cor	ncentration (mmol g <sup>-1</sup>	lw)			
A. ampliceps	0.19 g	0.60 f	1.01d	2.10 b	0.98 B
A. nilotica	0.18 g	0.76 e	1.41 c	2.62 a	1.24 A
Mean	0.19 D	0.68 C	1.21 B	2.36 A	
D. Root Cl <sup>-</sup> con-	centration (mmol g <sup>-1</sup> d	w)			
A. ampliceps	0.24 g	0.65f	1.34 d	2.47 b	1.18 B
A. nilotica	0.27 g	0.90 e	1.86 c	3.21 a	1.56 A
Mean	0.26 D	0.76 C	1.6 B	2.84 A	
E. Shoot K <sup>+</sup> cor	ncentration (mmol g <sup>-1</sup> c	lw)			
A. ampliceps	1.22 a	0.95 b	0.70 d	0.50 e	0.85 A
A. nilotica	1.20 a	0.82 c	0.55 e	0.29 f	0.71 B
Mean	1.21 A	0.89 B	0.60 C	0.42 D	
F. Root K <sup>+</sup> concentra	ation (mmol g <sup>-1</sup> dw)				
A. ampliceps	0.76 a	0.65 b	0.50 d	0.35 e	0.57 A
A. nilotica	0.75 a	0.58 c	0.38 e	0.19 f	0.48 B
Mean	0.76 A	0.62 B	0.44 C	0.27 D	

Means of a parameter sharing the same small letters are statistically similar at  $P \le 0.05$ .

effects of treatment and species were significant. In control, both species had same concentration of Na<sup>+</sup>. In saline treatments, A. ampliceps accumulated significantly lower Na<sup>+</sup> as compared to A. nilotica in both shoot and root. The difference between both species was maximum at 300 mM NaCl. In this treatment, maximum Na<sup>+</sup> concentration (2.3 mmol g<sup>-1</sup>dw) was found in the shoot of A. nilotica. At the same salinity level shoot Na+ concentration was 1.72 mmol g-1dw in A. ampliceps. The root Na+ concentration in 300 mM NaCl treatment was 2.86 and 2.1 mmol g<sup>-1</sup>dw for A. nilotica and A. ampliceps, respectively. Salinity significantly increased the shoot and root Cl concentration in both species (Table 2). The individual as well as interactive effects of treatment and species were found significant. In all saline treatments, A. ampliceps accumulated significantly lower Cl<sup>-</sup> as compared to A. nilotica in both shoot and root. At 300 mM NaCl, maximum Cl<sup>-</sup> concentration in the shoot of A. nilotica was 2.62 mmol g<sup>-1</sup>dw. Shoot Cl<sup>-</sup> concentration was 2.1 mmol g<sup>-1</sup>dw in A. ampliceps. At this salinity level the root Cl<sup>-</sup> concentration was 3.21 and 2.47 mmol g<sup>-1</sup>dw for A. nilotica and A. ampliceps respectively. The concentration

of K<sup>+</sup> as against of Na<sup>+</sup> decreased significantly at each increasing level of salinization (Table 2). The individual effects of treatments and species as well as their interaction were found significant. In control, K<sup>+</sup> concentration was the maximum whereas it was the minimum in 300 mM NaCl. The comparison of species at each salinity level showed that *A. ampliceps accumulated* more K<sup>+</sup> in both organs as compared to *A. nilotica*. At 300 mM NaCl, shoot K<sup>+</sup> concentration was 0.55 mmol g<sup>-1</sup>dw in *A. ampliceps* whereas in case of *A. nilotica* it was 0.29 mmol g<sup>-1</sup>dw. At this salinity level the root K<sup>+</sup> concentration was 0.35 and 0.19 mmol g<sup>-1</sup>dw for *A. ampliceps* and *A. nilotica*, respectively.

The K<sup>+</sup>: Na<sup>+</sup> ratio was maximum in control and decreased with increasing salinity levels. This ratio was minimum in 300 mM NaCl treatment. *A. ampliceps* had higher K<sup>+</sup>: Na<sup>+</sup> ratio than *A. nilotica* for both root and shoot under saline conditions (Fig.1&2). There was a strong negative correlation of Na<sup>+</sup> and Cl<sup>-</sup> concentration with shoot dry weight of both acacia species (Fig. 3&4). On the other hand a strong positive correlation was found between shoot K<sup>+</sup> concentration and shoot dry weight of both species (Fig. 5).

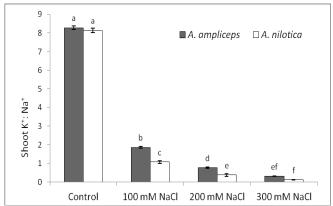
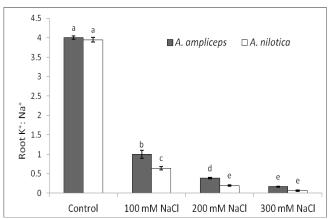


Figure 1. Effect of salinity on shoot K<sup>+</sup>: Na<sup>+</sup> ratio of acacia species. For both species vertical bars having a common

letter are not significantly different at P < 0.05.



**Figure 2. Effect of salinity on root K**<sup>+</sup>: Na<sup>+</sup> ratio of acacia species. For both species vertical bars having a common letter is not significantly different at P < 0.05.

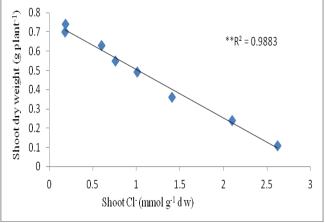


Figure 3. Correlation between shoot dry weight and Clconcentration of acacia species

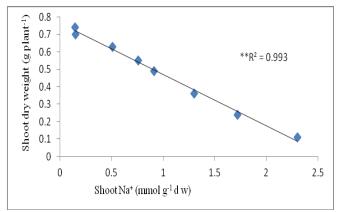


Figure 4. Correlation between shoot dry weight and Na<sup>+</sup> concentration of acacia species

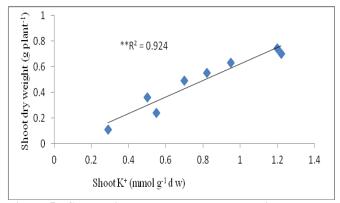


Figure 5. Correlation between shoot dry weight and K<sup>+</sup> concentration of acacia species

## DISCUSSION

This study indicated high salinity tolerance potential of the two acacia species, i.e. *A. amplicep* and *A. nilotica*. However, there was a distinct genetic difference between the species regarding salinity tolerance. *A. ampliceps* survived even at 400 mM NaCl whereas this salinity level proved to be fatal for *A. nilotica*. Such genetic variability in acacia was also observed by Marcar *et al.* (1991) who found that in a sand culture experiment, *A. ampliceps* and *A. auriculiformis* survived at 400 mM NaCl whereas *A. mangium* died at this salinity.

Shoot fresh and dry weights of both acacia species declined with increasing levels of salinization and maximum reduction was noticed at 300 mM NaCl. Beyond this salinity level, *A. nilotica* could not survive, however the seedlings of *A. ampliceps* survived. On relative basis, shoot dry weight was decreased up to 68 and 85% as compared to control in *A. ampliceps* and *A. nilotica* respectively. Salinity induced reduction in shoot growth in acacia was also reported by Farooq *et al.* (2010) and Yokota (2003). Reduction in the growth of plants due to salinity is mainly attributed to three

principle factors i.e. osmotic effects, ion toxicity and deficiency of necessary nutrients (Munns and Tester, 2008). These factors are operative at both cellular and whole plant level and manipulate all the metabolic activities of plants (Garg and Gupta, 1997). Ion toxicity and reduced osmotic potential of rooting medium are the main causes of reduced fresh and dry weights of shoot (Munns *et al.*, 1995).

Like shoot growth, the growth of root also decreased to a great extent due to salt treatments as compared to control. On relative basis, reduction in root dry weight as compared to control was 80 and 61% in A. nilotica and A. ampliceps, respectively. The length of both root and shoot was more in A. nilotica than A. ampliceps in control as well as in 100 and 200 mM NaCl levels. However, in these treatments the weight of both root and shoot was more in A. ampliceps than A. nilotica. The reason is that the leaves of A. ampliceps are much fleshy and heavier than those of A. nilotica and the roots of A. ampliceps have more lateral growth as compared to A. nilotica. In 300 mM NaCl level, there was severe reduction in length of both root and shoot; in this treatment A. ampliceps produced more shoot and root length than A. nilotica. Reduction in root growth due to salinity was also found by Hardikar and Pandey (2008) and Mahmood et al. (2010).

At higher levels of salinity both root formation (Kramer, 1983) and their elongation (Garg and Gupta, 1997) is reduced greatly. So both water and salt stress reduce the plant growth in an additive manner. The physical growth parameters like fresh and dry weights of both root and shoot are usually related to salinity tolerance potential of all plants at their early stages of growth so these are the tools of selection criteria for salinity tolerance (Larcher, 1995). Plants also have phytoremediation potential (Zulfiqar *et al.*, 2012).

The ionic composition revealed that shoot and root Na<sup>+</sup> and Cl<sup>-</sup> concentration of both species increased significantly in response to increasing salinity levels with more accumulation in A. nilotica than A. ampliceps. This higher accumulation of Na<sup>+</sup> in cells corresponds to the lower growth and dry weights of A. nilotica. These results are also supported by the findings of Marcar et al. (1991). The buildup of toxic ions in plant tissues is thought to be the major factor of decline in growth under salinity stress (Muscolo et al., 2003). There was a strong negative correlation of Na+ and Cl- concentration with shoot dry weight (Fig. 3&4). On the other hand, a strong positive correlation was found in case of shoot K+ concentration and shoot dry weight (Fig. 5). Ahmed et al. (2012) found an inverse correlation between the shoot growth and Na+ concentration in maize genotypes and Arshad et al. (2012) and Saqib et al. (2012) found similar relationship in wehat. Higher concentration of Na+ in tissues causes nutrient imbalance, osmotic effects and specific ion toxicity (Arzani, 2008).

K+ has a key role in salt tolerance where uptake of K+ is decreased by Na<sup>+</sup> (Fox and Guerinot, 1998). In our study, the reduction of K+ concentration was found in both root and shoot which indicated that Na<sup>+</sup> repressed the uptake of K<sup>+</sup>. The replacement of K<sup>+</sup> for Na<sup>+</sup> in the stele of the roots or in the vascular bundles in stems is thought as a mechanism for controlling transportation of salts from roots to shoots (Hardikar and Pandey, 2008). The higher concentration of Na+ in shoots of A. nilotica indicated that this mechanism was poorly operative in this species so it had low K+ concentration as compared to A. ampliceps. Decreased K+ uptake in response to salinity was also observed by (Ahmed et al., 2012; Marcar et al., 1991). The plants with higher K+: Na<sup>+</sup> ratio are considered more tolerant to salinity stress (Saqib et al., 2005). Similar was the case in our results where A. ampliceps owing to higher K+: Na+ ratio showed better salt tolerance. The higher Na<sup>+</sup>, Cl<sup>-</sup> and lower K<sup>+</sup>: Na<sup>+</sup> resulted in the reduction of root and shoot growth of A. nilotica more as compared to A. ampliceps.

**Conclusion:** Although salinity had a detrimental effect on all the growth parameters of the both acacia species, there was a distinct genetic difference between the tested species. On the basis of better ionic homeostasis *A. ampliceps* had more growth and survival at higher salinity levels than *A. nilotica*. Therefore, it can be successfully grown on highly salt-affected lands in order to get economic yield from otherwise barren lands.

#### REFERENCES

Ahmed, K., M. Saqib, J. Akhtar and R. Ahmad. 2012. Evaluation and characterization of genetic variation in maize (*Zea mays* L.) for salinity tolerance. Pak. J. Agri. Sci. 49:521-526.

Arshad, M., M. Saqib, J. Akhtar and M. Asghar. 2012. Effect of calcium on the salt tolerance of different wheat (*Triticum aestivum* L.) genotypes. Pak. J. Agri. Sci. 49: 497-504.

Arzani, A. 2008.Improving salinity tolerance in crop plants: a biotechnological view. Vitro Cell. Dev. Biol. Plant. 44:373-383.

Aslam, Z. 2006. Saline agriculture, p. 210-221. In: M.A.A. Quraishi, M.A. Zia and M.S. Quraishi (eds.), Pakistan Agriculture Management and Development. A-One Publishers, Lahore, Pakistan,

Benninson, J.J. and R.T. Paterson. 1993. Use of tree by livestock: Acacia Natural Resources Institute, United Kingdom. 32p.

El-kharbotly, A., O. Maghloub, A. Al-subhi, and A. Al-halhali. 2003. Indigenous grass species with potential for maintaining rangeland and livestock feeding in Oman. Agric. Ecosyst. Environ. 95:623-627.

- Farooq, H., N. Batool, J. Iqbal and W. Nouman. 2010. Effect of salinity and municipal wastewater on growth performance and nutrient composition of *Acacia nilotica*. Int. J. Agr. Biol. 12:591-596.
- Fox, T.C. and M.L. Guerinot. 1998. Molecular biology of cation transport in plants. Ann. Rev. Plant Physiol. Plant Mol. Biol. 49:669-696.
- Garg, B.K. and I.C. Gupta. 1997. Saline wastelands environment and plant growth. Scientific Publishers, Jodhpur, India. 287p.
- Hardikar, S.A. and A.N. Pandey. 2008. Growth, water status and nutrient accumulation of seedlings of *Acacia senegal* (L.) Willd.in response to soil salinity. Anal. Biol. 30:17-28.
- Khan, G.S. 1998. Soil salinity/sodicity status in Pakistan. Soil Survey of Pakistan, Lahore. 59p.
- Kramer, P.J. 1983. Water Relations of Plants. New York: Academic Press, Inc. 489p.
- Larcher, W. 1995. Physiological plant ecology: ecophysiology and stress physiology of functional groups. Springer-Verlag. Berlin. 540p.
- Mahmood, K., G. Sarwar, H. Schmeisky, N. Hussain and U. Saleem. 2010. Effect of various sodicity levels on growth parameters of *Acacia ampliceps*. J. Agric. Res. 48:381-90
- Majeed, A., M.F. Nisar and K. Hussain. 2010. Effect of saline culture on the concentration of Na<sup>+</sup>, K<sup>+</sup> and Cl<sup>-</sup> in Agrostistolonifera. Curr. Res. J. Biol. Sci. 2:76-82.
- Marcar, N.E., P. Dart and C. Sweeney. 1991. Effect of root-zone salinity on growth and chemical composition of *Acacia ampliceps* B. R. Maslin, *A. auriculiformis*, *A. cunn*. Ex Benth. and *A. mangium* Willd. at two nitrogen levels. New Phyt. 119:567-573.
- Munns, R. 1993. Physiological processes limiting plant growth in saline soils: some dogmas and hypotheses. Plant Cell Environ. 16:15-24.
- Munns, R., D.P. Schachtman and A.G. Condon. 1995. The significance of a two-phase growth response to salinity in wheat and barley. Aust. J. Plant Physiol. 22:561-569.
- Munns, R. and M. Tester. 2008. Mechanisms of salinity tolerance. Ann. Rev. Plant Biol. 59:651-681.
- Murtaza, B., G. Murtaza, M.Z. Rehman and A. Ghafoor. 2011. Use of brackish water for the reclamation of dense saline-sodic soils by auger hole technology and

- economic growth of wheat crop. Pak. J. Agri. Sci. 48:269-276.
- Muscolo, A., M. Sidari and M.R. Panuccio. 2003. Tolerance of kikuyu grass to long term salt stress is associated with induction of antioxidant defenses. Plant Growth Regul. 41:57-62.
- Saboora, A., K. Kiarostami, F. Behroozbayati, S.H. Hashemi. 2006. Salinity (NaCl) tolerance of wheat genotypes at germination and early seedling growth, Pak. J. Biol. Sci. 9:2009-2021.
- Saqib, M., C. Zorb, Z. Rengel and S. Schubert. 2005. Na<sup>+</sup> exclusion and salt resistance of wheat (*Triticumaestivum*) are improved by the expression of endogenous vacuolar Na<sup>+</sup>/H<sup>+</sup>antiporters in roots and shoots. Plant Sci. 169:959-965.
- Saqib, M., C. Zorb and S. Schubert. 2006. Salt-resistant and sensitive wheat genotypes do not show very different biochemical reaction at the level of proteins in the first phase of their response to NaCl salinity. J. Plant Nut. Soil Sci. 169:542-548.
- Saqib, Z.A., J. Akhtar, M.A. Ul-Haq and I. Ahmad. 2012. Salt induced changes in leaf phenology of wheat plants are regulated by accumulation and distribution pattern of Na<sup>+</sup> ion. Pak. J. Agri. Sci. 49:141-148.
- Schachtman, D.P., R. Kumar, J.I. Schroeder and E.L. Marsh.1997. Molecular and functional characterization of a novel low-affinity cation transporter (LCTI) in higher plants. Pro. Nat. Acad. Sci. USA. 94:11079-11084.
- Steel, R.G.D., J.H. Torrie and D.A. Dickey. 1997. Principles and Procedures of Statistics: A Biometrical Approach. 3rd ed. McGraw Hill Co., New York, USA.
- Taiz, L. and E. Zeiger. 2006. Plant physiology. 4<sup>th</sup> ed. Sinauer Associates, Inc., Publishers Sunderland, USA.
- Yokota, S. 2003. Relationship between salt tolerance and proline accumulation in Australian acacia species. J. For. Res. 8:89-93.
- Wolf, B.A. 1982. Comprehensive system of leaf analysis and its use for diagnosis crop nutrient status. Commun. Soil Sic. Plant Anal. 13:1035-1059.
- Zhu, J.K. 2001. Plant soil tolerance. Trends Plant Sci. 6:66-71.
- Zulfqar, S., A. Wahid, M. Farooq, N. Maqbool and M. Arfan. 2012. Phytoremediation of soil cadmium using *Chenopodium* species. Pak. J. Agri. Sci. 49:435-445.