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**Review Article** 

# A Review on Histopathological Alterations Induced by Heavy Metals (Cd, Ni, Cr, Hg) in Different Fish Species

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#### Authors' Contributions

SB and SS collected the data and wrote the manuscript. SN, AMMC and SS conceived the study and approved the article.

#### Keywords

Histopathology, Toxicity, Bioaccumulation, Toxic metals, Heavy metal pollution **Abstract** | Industrial, domestic and other human activities are responsible for heavy metal toxicity of aquatic bodies. Metals are the major stimulators for variety of diseases in aquatic organisms mainly fishes. Chromium (Cr), mercury (Hg), nickel (Ni) and cadmium (Cd) causes histopathological variations and several diseases in various fishes of Pakistan. Extensive histopathological variations in gills, liver, kidney and skin of different fishes were noticed when exposed to heavy metals, thus indicating severity of heavy metal toxicity. The current study was focused on the toxicological effect of Cr, Hg, Ni and Cd in different type of fishes and it would be useful for the scientific community to restore different metals contaminating water.

**Novelty Statement** | Histopathological alterations in target fish organs functioned as fundamental indicator to assess metal toxicity in different fish species as it provides accurate data on effects of different contami-nants on fish. Hence, histopathology must be consider to monitor contaminated aquatic systems.

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# Introduction

In natural environment, pollution owing to heavy metals is a major issue. Heavy metal toxicity and their bioaccumulation easily enter humans via food chain (Tahir *et al.*, 2017) and also affect other organisms. In an ecosystem, metal residual distribution revealed contaminated area. Toxicity can easily be determine by considering various factors as to measure the level of feed and water pollution, bioaccumulation and toxicity of heavy metals along with bio magnifications in living beings (Has-Schön *et al.*, 2006; Saleemi *et al.*, 2019).

Heavy metals intake is a main reason for metal accumulation in different fish organs (Javed, 2015).

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In aquatic environment, metal toxicity not only effect aquatic life (Fish) but also humans as they are consumers in food chain (Faiza *et al.*, 2015; Perera *et al.*, 2015). Various fish species are used to measure health conditions of aquatic life and their ecosystem, as metal pollution is increasing day by day (Abbas and Javed, 2016). Gills and food web are directly associated for the entry of heavy metals in aquatic life and also get entry in other organs. When accumulated, metals brings oxidative stress, thus causes genotoxicity (Khan *et al.*, 2015a, b). Different biological techniques are used to evaluate metal toxic levels and their impact on fish and other organism's behavior, their physiological sensitivity as well as morphological indices (Yang *et al.*, 2014; Kousar and Javed, 2015).

Different metals greatly effect cell organelles as well as metabolic and detoxifying enzymes (Wang and Shi, 2001). Metals also effect DNA and nuclear proteins,

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• Links • Researchers hence causes apoptosis or carcinogenesis (Beyersmann and Hartwig, 2008). Liver plays an important role in toxic material transformation (Udotong, 2015). Mercury (Hg) is a lethal metal and exists naturally in earth crust. Erosion, volcanic eruptions and anthropogenic activities (Industrial, agricultural and municipal wastes) may contaminate environment. As fish is on the top of aquatic food chain so metals from the water can easily enter the body. Increase mercury level in water brings more accumulation is fish organs.

More the amount of mercury in aquatic environment, greater will be its accumulation in fish tissues, and also in human beings (Gilbertson and Carpenter, 2004; Guilherme *et al.*, 2008). Due to methyl mercury rapid swimming was noticed in Zebra fish (Zamorano *et al.*, 2016). Cadmium (Cd) has teratogenic, carcinogenic and mutagenic effects on aquatic life. Freshwater ecosystem is effected by Cd owing to higher sensitivity towards aquatic life compared with mammals (Burger, 2008; Jia *et al.*, 2011).

Nickel (Ni) also possess toxic behavior and its greater concentration causes complications in blood, respiratory system, tissues as well as mucus membranes and effect different cells and chromosomes. Thus, the European Union had offered World Trade Organization (WTO) to consider nickel as a hazardous element (Purwanti et al., 2019). Nickel can also cause metal toxicity in organs, contact dermatitis, allergy, morphological alterations in cellular systems and even chromosomal abnormalities (Coen et al., 2001; Palaniappan and Karthikeyan, 2009). Literature on nickel accumulation in fish is limited compared with other metals (Palermo et al., 2015). Chromium (Cr) toxicity may cause mucus discharge, color changes, erratic motion and even abrupt swimming as in Danio rerio (Zebra fish) (Nisha et al., 2016). Thus, the aim of current work was to estimate the effect of different heavy metals i.e. mercury, cadmium, nickel and chromium on fishes that brings histopathological variations and also causes various diseases. In Pakistan, fish species are going to reduce due to heavy metal accumulation and present study reveals that heavy metal exposure in aquatic life increases damage to fish tissues and organs.

# Materials and Methods

Present study was designed to review the studies from 2000-2020 and it was based on the histopathological alterations on fish. Various articles were searched from Google Scholar and PubMed using keywords as histopathology, heavy metals, toxicity, bioaccumulation, aquatic life, fish species, and histopathological abnormalities. Latest research papers and articles were selected to avail some relevant data. All the papers were cited along with their proper references. Aquatic toxicity Industrial effects

Countries being developed are in front of many issues and one of them is water pollution. This is just because of industrialization which is spreading very quickly and civilization as well. Very huge number of pollutants and polluting products are released from these industries. From these pollutants, various heavy metals are on top of the list which is continually being released into adjacent water bodies without treating them. Hazardous impacts of heavy metals on water ecosystem have been considered as a global issue (Yousafzai *et al.*, 2008). These toxic metals are allowed to be released in the environment through anthropogenic and natural activities i.e., industrial and urban discharges, mining combustion processes and mining etc. (Barone *et al.*, 2013; Ambreen *et al.*, 2015).

#### Biochemical toxicity

Chromium is considered to be exerting its toxic effects on intestinal epithelial cells and brings changes in functions regarding glucose transportation. A study on the intestine of rainbow trout revealed shortage of glucose absorption by the epithelial cells of intestine (Stokes and Fromm, 1965).

Various fish species' osmoregulatory functions have been affected by the impact of trivalent chromium ( $Cr^{3+}$ ). A fish species named *Cyprinus carpio var. communis* upon chromium sulphate [ $Cr_2(SO_4)_3$ ] exposure brought variations in osmoregulation functioning in fish (Subashini *et al.*, 2005). Heavy metals exposure up to different levels might cause various behavioral and morphological aberrations in fish (Qadir and Malik, 2011). Cd is considered as a toxic heavy metal and also have disrupting properties of endocrine (Afridi *et al.*, 2011; Chouchene *et al.*, 2011).

Some metals like Cd, Cr, Pb, Hg and As being non-essential elements can exert toxic effects even at very low concentration. While Co, Mn, Zn, Cu, and Ni are the essential metals producing lethal effects at high concentrations (Couture and Rajotte, 2003).

#### Bioaccumulation of heavy metals

Fish attained the ability of heavy metals bioaccumulation in their tissues via absorption, while humans can come into exposure of these metals by the food chain. As fish is the prior in human's diet thus affecting humans in different ways (Dogan and Yimaz, 2007). Heavy metal's bioaccumulation and their bio-magnification is linked with the pathway of transformation of pollutants from one trophic level to another (Ghannam *et al.*, 2015).

In marine biota, the accumulation of Cr needs to be discovered. Marine biota which uses gill tissues for absorption is more affected by its concentration. As per known, marine organisms uses gills for the absorption process as major and main part of their body. Cr might deposit at different concentrations in different tissues. Fish gills, kidney and liver were found to be containing high concentration of Cr and muscles were not found to be affected and accumulated by the metal. However, Cr cannot be accumulated at higher trophic levels in marine food webs (Pourahmad *et al.*, 2005).

#### Histopathological alterations

Histopathological studies are thought to be consistent and comprehensive biomonitoring tool to analyze the fish health and its survival rate in accumulated aquatic ecosystems (Deore and Wagh, 2012). Effects of metals i.e. Cd, Cr, Ni and Hg in different fish species present in Pakistan were noticed along with their histopathological changes (Table 1).

# Heavy metals and diseases

#### Cadmium

Metals uptake primarily occurs by gills but it can also occur through intestinal epithelium (Mohamed, 2008). Some of the aberrations were observed in the intestine of *Oreochromis nioticus* and *Lates niloticus*. It was noticed that there were various severe degenerative and necrotic alterations in their intestinal mucosa. Kaoud *et al.* (2011) studied some pathological changes in the intestine of *Oreochromis niloticus* exposed with Cd along with atrophy in muscularis, necrotic and degenerative alterations in mucosa and sub mucosa. As the liver is associated with biotransformation and detoxification processes so due to these functions, its location, and its access to the body supply, it is an organ which is mostly affected by the water contamination (Camargo and Martinez, 2007; Mohamed, 2009).

## Nickel

Generally, Ni present in trace amounts in the fish, if its concentration increases; it would be a risk for the survival of water organisms (Rauf *et al.*, 2009). An extensive use of Ni, in various activities like, ceramic, steel industries and electroplating which are making Ni containing products, releasing waste materials into adjacent water channels without treating them (Jabeen and Javed, 2011). Fish accumulates heavy metals in its body by various ways i.e., through respiratory system, digestive system and skin directly (Ma'ruf, 2007). One of a fish named *Barbonymus gonionotus*, is considered to be sensitive to the environmental fluctuations and that's why they act as bio indicator of environmental changes in the aquatic ecosystem (Wajiya and Yazid, 2009).

## Chromium

Cr has deteriorating effects on fish survival and its growth (Shaheen *et al.*, 2012). It just not only damage fish fertility but also produces aberrations in their progeny as well. Breeders' exposure before spawning

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induced cumulative impact on fish gonads and it results in meager, weak and retarded off-springs just because of the malfunctioning of trophic hormones (Kobayashi and Okamura, 2005). It was noticed that, any decrease in activation of mitogen induced lymphocytes and a noticeable change in shape and phagocytic functions of neutrophils. These alterations may reflect the reduced power of resistance to pathogens chronic exposure of Cr in fish (Steinhagen *et al.*, 2004).

# Mercury

Studies revealed that, Hg ions increases the formation of vacuole in hepatocytes and hepatic disintegration occurs in severe cases. Damage increases with persistent exposure of Hg, indicating its accumulation in fish body. This study also in the favor of Kuang *et al.* (2007), who studied the roles on heavy metal's ions in *A. japonica* under the water. Toxicity of heavy metals may lead to various pathological alterations in various fish tissues and these changes also has been addressed in *Labeo rohita* when exposed to mercuric chloride and *Channa puntatus* when exposed to phenyl mercuric acetate (Karuppasamy, 2000). Some major diseases caused by heavy metal toxicity are:

## Necrosis

Hepatocytes necrosis and its degeneration might be due to the cumulative effects of metals and an increase in their concentrations in liver. Liver cellular degeneration might be due to oxygen deficiency as a result vascular dilation along with intravascular haemolysis was noticed in blood vessels (Mohamed, 2001). Lesions as degeneration, blood vessels dilation and necrosis in hepatocytes were detected in *Mugil cephalus*, *Tilapia mossambicus* and *Clarias gariepinus* (Ibrahim and Mahmoud, 2005).

## Oedema

Labeo rohita gills when exposed to tannery effluent indicated fusions and primary lamellar epithelium clumping (Fanta *et al.*, 2003). Some degenerative alterations in edema and lamellar were observed in fish gills when exposed to heavy metals (Osman *et al.*, 2009). The lamellar epithelium lifting is a change of tissues, might be induced by any incidence of severe edema (Pane *et al.*, 2004; Schwaiger *et al.*, 2004).

## Hyperplasia

Epithelial cells which undergo the lamella fusion and hyperplasia can go on to cause a huge decrease in gill surface area for respiration. It also disrupts blood flow in gills, alters the functioning of metabolism and may cause fish mortality (Purwanti *et al.*, 2019). Recently, some of the researchers demarcated the gill injuries of two types. First type of injury comes from defense response and it includes hyperplasia of gill epithelium filaments while the second type is direct injuries which include shedding and necrosis of gill epithelium (Fanta *et al.*, 2003). S. Bibi et al.

Fish species	Metal	Concentration	Exposure type	Exposure time	Organ	Histopathological changes	References
Wallago attu	Cadmium	1/3, 1/4, 1/5 of LC50	Sub-lethal	14days	Liver	Protein level decreased	(Batool <i>et al.</i> , 2018)
Catla catla	Nickel	70.40 mg/L	Sub-lethal	60days	•••••	Total length, fork and weight increased	(Safina <i>et</i> <i>al.</i> , 2020)
Labio rohita		71.99 mg/L				Weight increment	
Cirrhinus mrigala		79.9 mg /L				Weight increment	
Channa striata	Nickel	1/3, 1/4, 1/5 of LC50	Sub-lethal	14days		CAT activity reduced	(Arshad <i>et</i> <i>al.</i> , 2018)
Catla catla	Cadmium		Acute	96hrs	Blood	Hemoglobin, hematocrit and RBCs decreased	(Hassan <i>et</i> <i>al.</i> , 2018)
Channa marulius	Cadmium	101.25mg/L	Acute	96hrs	Liver	Liver sensitivity increased	(Javed et
Mystus seenghala							al., 2016)
Wallago attu							
Ctenopharyngodon idella	Chromium	25.5mg/L	Acute	24hrs	Gills, muscle tissues	Swelling in gills cells, fusions, in- flammation. muscle cells necrosis, RBCs and hemoglobin decline.	(Shah <i>et al</i> ., 2020a)
		22.5mg/L		48hrs			
		20mg/L		72hrs			
		18mg/L		96hrs			
Oreochromis niloticus	Mercury	0.044 mg/L	Sub-lethal	48hrs	Liver	Hepatocytes destruction, sinu- soids, pyknotic nuclei	(Pervaiz <i>et</i> <i>al.</i> , 2019)
				96hrs		Cellular necrosis, melano-mac- rophages centers appearance, pyknotic and karyolosis nucleus, hepatic cords disorganization.	
		0.055 mg/L		48hrs		Tissue degeneration, hepatic cords disorganzation, bile ducts degeneration, pyknosis	
				96hrs		Bile duct inflammation, cellular necrosis, pyknotic nuclei, hemo- poietic tissue degeneration.	
		0.073 mg/L		48hrs		Pyknotic nuclei, hemopoietic tissue degeneration, bile duct degeneration, vacuolization, necrosis.	
				96hrs		Pyknosis, necrosis, degeneration of tissue, bile duct inflammation, vacuolization.	
Carassius auratus	Chromium	4ppm	pm Sub-lethal pm	24hrs	Gills Intestine Skin	Increased breathing rate. Increased mucus secretion. Tissue residues.	(Fawad <i>et</i> <i>al.</i> , 2017)
		6ppm		48hrs			
		8ppm		72hrs			
		12ppm		96hrs			
Oreochromis niloticus	Cadmium mixture	183.4 ml	Chronic	2weeks	Kidney	Renal CAT activity increased firstly, decreased finally.	(Ahmed <i>et al.</i> , 2016)
Ctenopharyngo- donidella	Chromium	45.5, 43.5, 41.5, and 39 mg /L	Lethal	96hrs	Blood	RBCs and hemoglobin level decreased	(Shah <i>et al</i> ., 2020)
Labeo rohita	Nickel	0.32 - 1.22 mg/L	Sub-lethal	River water	Gills	Hyperplasia, necrosis, Blood vessels congestion	(Sultana <i>et</i> <i>al.</i> , 2016)
					Intestine	Atrophy,	
					Liver	Necrosis	
						Table continue on next page	

Fish species	Metal	Concentration	Exposure type	Exposure time	Organ	Histopathological changes	References
Tilapia nilotica	Cadmi- um-cobalt mixture	37.93 mg/L	Acute	96hrs	Tissues	Exhibited sensitivity higher	(Ambreen and Javed, 2016a)
Cyprinus carpio	Chromium	5 mg/Land 10mg/L	Sub-lethal	16 days	Liver	Blood congestion, necrosis, pa- renchyma cell degeneration.	(Rana <i>et al.</i> , 2015)
					Kidney	Pyknotic nuclei reduction, haemolysis.	
Gibelion catla	Chromium	1/3rd of LC50	Chronic sub-lethal	12 weeks	Liver	Growth performance affected.	(Azmat <i>et</i> <i>al.</i> , 2018)
Labeo rohita					Kidney		
Cirrhinus mrigala					Gills		
Cyprinus carpio	Chromium	25 to 150mg/L	Sub-lethal	30days		Deleterious effects on growth produced.	(Shaheen and Jabeen, 2015)
Cyprinus carpio	Cadmium	10.42, 7.81, 6.25 and 5.21mg/L	Sub-lethal	30days	Blood	DNA damage	(Ambreen and Javed, 2016b)
Cyprinus carpio	Cadmi- um-lead mixture	78.94 and 82.22 $\mu gg^{-1}$	Lethal	96hrs	Kidney	Higher accumulation, increase sensitivity.	(Ambreen <i>et al.</i> , 2015)

Histopathological variations due to heavy metals in fish species

#### **Results and Discussion**

Different metals (Hg, Cr and Cd) are considered as systemic toxicants, thus affect various organs even at low level (Olsson *et al.*, 1998). Cellular organelles as well as enzymes involved in detoxification, damage repair and metabolic process are greatly influenced by heavy metals (Wang and Shi, 2001). Metallic ions effect nuclear proteins and DNA molecule, hence brings apoptosis or carcinogenesis (Beyersmann and Hartwig, 2008).

Histological biomarkers regarding fish organs toxicity are important indicators of environmental pollution. Biomarkers permits to examine specific target organ (gills) that are responsible for their vital function as respiration (Gernhöfer et al., 2001; Peebua et al., 2008). Gills are mainly involved in entry of heavy metals that brings lesions and gill damage and structural variations in gills of exposed fish are responsible for deaths (Bols et al., 2001; Obomanu et al., 2007). A study indicated that histopathological changes in gill tissues of Channa might cause hypoxic disorders, thus affecting their locomotory ability (Begum et al., 2006). Various organic compounds causes toxicopathic lesions in liver of fish. Cloudy swelling, hydropic degenerations as well as karyolysis, pyknosis and karyorrhexis of nuclei occurs due to acute toxic injury (Paul et al., 2014; Jiraungkoorskul et al., 2003). Cytoplasmic vacuoles of hepatocytes containing glycogen and lipids are associated with normal liver functioning (Camargo and Martinez, 2007). Hepatocytes vacuolation brings metabolic changes when exposed to contaminated water. Similarly, histopathological variations with Cu and Cd were noticed in liver of teleostean fishes (Pacheco and Santos, 2002; Ibrahim and Mahmoud, 2005; Tayel et al., 2008).

Fish can even cope with increased level of heavy metals in their bodies but is not safe for human consumption. Thus, time duration of contact with toxic elements and fish species as well as concentration rate must be considered before establishing basics for tolerable levels in ecological studies (Usero et al., 2004). The oedema can lead to tissue swelling due to fluid accumulation and cause cell organelles to decrease cell permeability (Rennika and Nurlita, 2013). Oedema may cause hyperplasia occurrence which may lead to interlamellae being clogged, thus causing the entire space to be filled with new cells and thicken the epithelium present at lamellae base (Robert, 2001). Epithelial degeneration, inflammatory cell infiltration in sub mucosa along with sub mucosal edema was detected in the intestine of tilapia when exposed to carbofuran (Soufy et al., 2007). Histopathological changes in the intestine of Oreochromis niloticus and Lates niloticus indicated severe degenerative and necrotic variations in the intestinal mucosa. Edema may be due to absorption of toxic metals (Hanna et al., 2005). Thus, various studies reported heavy metal accumulation in different fish organs and changes were noticed regarding particular heavy metal.

#### **Conclusions and Recommendations**

The current review indicates that exposure of heavy metals such as Cr, Cd, Ni and Hg at various concentrations brings histopathological changes in different fish organs. Such histopathological variations would be supportive to determine effect of heavy metals on fish species. Hence, present research served as experimental tools for the estimation of environmental pollution.

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## Conflict of interest

The authors have declared no conflict of interest.

# References

- Abbas, S. and Javed, M., 2016. Growth performance of *Labeo rohita* under chronic dual exposure of waterborne and dietary cobalt. *Pak. J. Zool.*, **48**: 257–264.
- Afridi, H.I., Kazi, T.G., Kazi, N., Kandhro, G.A., Baig, J.A., Jamali, M.K., Arain, M.B. and Shah, A.Q., 2011. Interactions between cadmium and zinc in the biological samples of Pakistani smokers and nonsmokers cardiovascular disease patients. *Biol. Trace Elem. Res.*, 139: 257–268. https://doi. org/10.1007/s12011-009-8607-3
- Ahmed, T., Abdullah,S., Abbas, K. and Zia, M.A., 2016. Catalase enzyme response to chronic Pb+ Cd metal mixture exposure, its purification and partial characterization from the kidney of freshwater fish, *Oreochromis niloticus. Pak. J. Zool.*, 48: 1733-1740.
- Ambreen, F., Javed, M. and Batool, U., 2015. Tissue specific heavy metals uptake in economically important fish, *Cyprinus carpio* at acute exposure of metals mixtures. *Pak. J. Zool.*, 47: 399-407.
- Ambreen, F. and Javed, M. 2016a. Metal mixtures toxicity and bioaccumulation in *Tilapia nilotica* at 96-Hr LC 50 exposure. *J. Bioresour. Manag.*, 3: 14-21. https://doi.org/10.35691/JBM.6102.0058
- Ambreen, F. and Javed, M., 2016b. Effects of 30 day sub-lethal exposure of cadmium and lead mixture on dna damage in fish. *J. Anim. Plant Sci.*, 26: 674-679.
- Arshad, R., Abdullah, S., Naz, H. and Abbas, K., 2018. Catalase activity as a bio-indicator of lead+ nickel toxicity in carnivorous fish, *Channa striata. Proc. Pak. Acad. Sci. B. Life Environ. Sci.*, 55: 37–43.
- Azmat, H., Javed, M., Abdullah, S., Javid, A. and Hussain, S., 2018. Growth responses of major carps reared under chronic stress of chromium. *J. Anim. Plant Sci.*, 28: 604-609.
- Barone, G., Giacominelli, R.S. and Storelli, M.M., 2013.
  Comparative study on trace metal accumulation in the liver of two fish species (Torpedinidae): Concentration-size relationship. *Ecotoxicol. Environ. Saf.*, 97: 73-77. https://doi.org/10.1016/j.

#### ecoenv.2013.07.004

- Batool, Y., Abdullah, S., Naz, H. and Abbas, K., 2018. Sub-lethal effect of waterborne cadmium exposure on glutathione S-transferase and total protein contents in liver of carnivorous fish, *Wallago attu. Proc. Pak. Acad. Sci: B. Life Environ. Sci.*, 55: 21–25.
- Begum, G., Venkateswara, R.J. and Srikanth, K., 2006. Oxidative stress and changes in locomotor behavior and gill morphology of *Gambusia affinis* exposed to chromium. *Toxicol. Environ. Chem.*, 88: 355-365. https://doi.org/10.1080/02772240600635985
- Beyersmann, D. and Hartwig, A., 2008. Carcinogenic metal compounds: recent insight into molecular and cellular mechanisms. *Arch. Toxicol.*, 82: 493-512. https://doi.org/10.1007/s00204-008-0313-y
- Bols, N.C., Brubacher, J.L., Ganassin, R.C. and Lee, L.E.J., 2001. Ecotoxicology and innate immunity in fish. *Dev. Comp. Immunol.*, **25**: 853-873. https://doi.org/10.1016/S0145-305X(01)00040-4
- Burger, J., 2008. Assessment and management of risk to wildlife from cadmium. *Sci. Total Environ.*, **56**: 471-502. https://doi.org/10.1016/j. scitotenv.2007.08.037
- Camargo, M.M. and Martinez, C.B. 2007. Histopathology of gills, kidney and liver of a Neotropical fish caged in an urban stream. *Neotrop Ichthyol.*, 5: 327-336. https://doi.org/10.1590/ S1679-62252007000300013
- Chouchene, L., Banni, M., Kerkeni, A., Saïd, K. and Messaoudi, I., 2011. Cadmium-induced ovarian pathophysiology is mediated by change in gene expression pattern of zinc transporters in zebrafish (*Danio rerio*). *Chem. Biol. Interact.*, **193**: 172–179. https://doi.org/10.1016/j.cbi.2011.06.010
- Coen, N., Mothersill, C., Kadhim, M. and Wright, E.G., 2001. Heavy metals of relevance to human health induce genomic instability. *J. Pathol.*, 195: 293-299. https://doi.org/10.1002/path.950
- Couture, P. and Rajotte, J.W., 2003. Morphometric and metabolic indicators of metal stress in wild yellow perch (*Perca flavescens*) from Sudbury, Ontario: A review. J. Environ. Monit., 5: 216-221. https://doi. org/10.1039/b210338a
- Deore, S.V. and Wagh, S.B., 2012. Heavy metal induced histopathological alterations in liver of *Channa gachua* (Ham). *J. Exp. Sci.*, **3**: 35–38.
- Dogan, M. and Yimaz, A.B., 2007. Heavy metals in water and in tissues of himir (*Carasobarbus luteus*) from Orontes (Asi) River, Turkey. *Environ. Monit.* Assess., 53: 161-168. https://doi.org/10.1007/s10661-007-0005-8
- Faiza, A., Javed, M. and Ummara, B., 2015. Tissue specific heavy metals uptake in economically important fish, *Cyprinus carpio* at acute exposure of metals mixtures. *Pak. J. Zool.*, 47: 399-407.

- Fanta, E., Rios, F.S., Romao, A., Vianna, S. and Freiberger, S., 2003. Histopathology of the fish *Corydoras paleatus* contaminated with sublethal levels of organophosphorus in water and food. *Ecotoxicol. Environ. Saf.*, 54: 119-130. https://doi. org/10.1016/S0147-6513(02)00044-1
- Fawad, M., Yousafzai, A.M., Haseeb, A., Rehman, H.U., Jan, A.A., Akhtar, N., Saeed, K. and Usman, K., 2017. Acute toxicity and bioaccumulation of chromium in gills, skin and intestine of goldfish (*Carassius auratus*). J. Entomol. Zool. Stud., 5: 568-571.
- Gernhöfer, M., Pawet, M., Schramm, M., Müller, E. and Triebskorn, R., 2001. Ultrastructural biomarkers as tools to characterize the health status of fish in contaminated streams. J. Aquat. Ecosyst. Stress Recovery, 8: 241–260. https://doi. org/10.1023/A:1012958804442
- Ghannam, H.E., El Haddad, E.S.E. and Talab, A.S., 2015. Bioaccumulation of heavy metals in tilapia fish organs. *J. Biol. Environ. Sci.*, **7**: 88-99.
- Gilbertson, M. and Carpenter, D.O., 2004. An ecosystem approach to the health effects of mercury in the Great Lakes basin ecosystem. *Environ. Res.*, 95: 240-246. https://doi.org/10.1016/j. envres.2004.02.015
- Guilherme, S., Valega, M., Pereira, M.S., Santos, M.A. and Pacheco, M., 2008. Antioxidant and biotransformation responses in *Liza aurata* under environmental mercury exposure relationship with mercury accumulation and implications for public health. *Mar. Pollut. Bull.*, 56: 845-859. https://doi. org/10.1016/j.marpolbul.2008.02.003
- Hanna, M.I., Shaheed, I.B. and Elias, N.S., 2005. A contribution on chromium and lead toxicity in cultured *Oreochromis niloticus*. *Egypt. J. Aquat. Biol. Fish.*, **9**: 177-209.
- Hassan, W., Abdullah, S., Afzal, M. and Hussain, M., 2018. Assessment of acute metals toxicity in *Catla catla* through hematological and biochemical blood markers. *Pak. J. Agric. Sci.*, **55**: 449-454. https://doi. org/10.21162/PAKJAS/18.6002
- Has-Schön, E., Bogut, I. and Strelec, I., 2006. Heavy metal profile in five fish species included in human diet, domiciled in the end flow of River Neretva (Croatia). Arch. Environ. Contam. Toxicol., 50: 545-551. https://doi.org/10.1007/s00244-005-0047-2
- Ibrahim, S.A. and Mahmoud, S.A., 2005. Effect of heavy metals accumulation on enzyme activity and histology of liver of some Nile fish in Egypt. *Egypt. J. Aquat. Biol. Fish.*, 9: 203-219. https://doi. org/10.21608/ejabf.2005.1824
- Jabeen, G. and Javed, M., 2011. Evaluation of arsenic toxicity to biota in river Ravi (Pakistan) aquatic ecosystem. *Int. J. Agric. Biol.*, **13**: 929-934.

- Javed, M., 2015. Chronic dual exposure (waterborne and dietary) effects of cadmium, zinc and copper on growth and their bioaccumulation in *Cirrhina mrigala. Pak. Vet. J.*, **35**: 143-146.
- Javed, M., Abbas, S. and Latif, F., 2016. Acute toxicity of cadmium and its bio-accumulation in the carnivorous fish species *Channa marulius*, *Mystus seenghala* and *Wallago attu. Int. J. Agric. Biol.*, **18**: 1169-1173. https://doi.org/10.17957/IJAB/15.0221
- Jia, X., Zhang, H. and Liu, X., 2011. Low levels of cadmium exposure induced DNA damage and oxidative stress in the liver of Oujiang colored common carp *Cyprinus carpio* var. color. *Fish Physiol Biochem.*, 37: 97-103. https://doi.org/10.1007/ s10695-010-9416-5
- Jiraungkoorskul, W., Upatham, E.S., Kruatrachue, M., Sahaphong, S., Vichasri-Grams S. and Pokethiyiyook, P., 2003. Biochemical and histopathological effects of glyphosate herbicide on Nile tilapia (*Oreochromis niloticus*). *Environ. Toxicol.*, 18: 260-267. https://doi.org/10.1002/tox.10123
- Kaoud, H.A., Zaki, M.M., El-Dahshan, A.R., Saeid, S. and El-Zorba, H.Y., 2011. Amelioration the toxic effects of cadmium-exposure in Nile tilapia (*Oreochromis Niloticus*) by using *Lemnagibba L. J. Life Sci.*, 8: 185-195.
- Karuppasamy, R., 2000. Tissue Histopathology of *Channa punctafus* (Bloch) under phenyl mercuric acetate toxicity. *Bull. Pure Appl. Sci.*, **19**: 109-116.
- Khan, M.S., Jabeen, F., Asghar, M.S., Qureshi, A.N., Shakeel, M., Noureen, A. and Shabbir, S., 2015a.
  Role of nao-ceria in the amelioration of oxidative stress: Current and future applications in medicine. *Int. J. Biosci.*, 6: 89-109. https://doi.org/10.12692/ ijb/6.8.89-109
- Khan, M.S., Jabeen, F., Qureshi, A.N., Asghar, S.M., Shakeel, M. and Noureen, A., 2015b. Toxicity of silver nanoparticles in fish. A critical review. *J. Biodiver. Environ. Sci.*, **6**: 211-227.
- Kobayashi, N. and Okamura, H., 2005. Effects of heavy metals on sea urchin embryo development. Part 2. Interactive toxic effects of heavy metals in synthetic mine effluents. *Chemosphere*, **61**: 1198-1203. https:// doi.org/10.1016/j.chemosphere.2005.02.071
- Kousar, S. and Javed, M., 2015. Diagnosis of metals induced DNA damage in fish using comet assay. *Pak. Vet. J.*, **35**: 168-172.
- Kuang, W.H., Zhang, D.Y. and Huang, X.Y., 2007. Study on residual rules of heavy metal ions Cd, Hg, and Pb in eels. *Sci. Bull.*, **23**: 689-692.
- Ma'ruf, M., 2007. Analysis of heavy metal concentrations in Baronang (Siganus sp.) and aquatic environment for the management of Bontang coastal areas. Thesis. Graduate program. Mulawarman University.
- Mohamed, F.A., 2001. Impacts of environmental

pollution in the southern region of Lake Manzalah, Egypt, on the histological structures of the liver and intestine of *Oreochromis niloticus* and *Tilapia zilli. J. Egypt. Acad. Soc. Environ. Dev.*, **2**: 25-42.

- Mohamed, F.A.S., 2008. Bioaccumulation of selected metals and histopathological alterations in tissues of *Oreochromis niloticus* and *Lates niloticus* from Lake Nasser, Egypt. *Glob. Vet.*, **2**: 205-2018.
- Mohamed, F.A.S., 2009. Histopathological studies on *Tilapia zillii* and *Solea vulgaris* from Lake Qarun, Egypt. *World J. Fish Marine Sci.*, **1**: 29-39.
- Nisha, J.C., Sekar, R.R.J. and Chandran, R., 2016. Acute effect of chromium toxicity on the behavioral response of Zebra fish *Danio rerio. Int. J. Plant Anim. Environ. Sci.*, **6**: 6-14.
- Obomanu, F.G., Ogbalu, O.K., Gabriel, U.U. Fekarurhobo, G.K. and Abadi, S.U., 2007. Piscicidal effects of *Lepidagathis alopecuriodes* on mudskipper, Periophthalmus papillio form the Niger Delta. Res. J. Appl. Sci., 2: 282-387.
- Olsson, P.E., Kling, P. and Hogstr, C., 1998. Mechanisms of heavy metal accumulation and toxicity in fish. *Metal Metab. Aquat. Environ.*, pp. 321-350. https:// doi.org/10.1007/978-1-4757-2761-6\_10
- Osman, M.M., El-Fiky, S.A., Soheir, Y.M. and Abeer, A.I., 2009. Impact of water pollution on histopathological and electrophoretic characters of *Oreochromis niloticus* Fish. *Res. J. Environ. Toxicol.*, **3**: 9-23. https://doi.org/10.3923/rjet.2009.9.23
- Pacheco, M. and Santos, M.A., 2002. Biotransformation, genotoxic and histopathological effects of environmental contaminants in European eel (*Anguilla anguilla L.*). *Ecotoxicol. Environ. Saf.*, 53: 331-347. https://doi.org/10.1016/S0147-6513(02)00017-9
- Palaniappan, P.L.R.M. and Karthikeyan, S., 2009. Bioaccumulation and depuration of chromium in the selected organs and whole body tissues of freshwater fish *Cirrhinus mrigala* individually and in binary solutions with nickel. *J. Environ. Sci.*, 21: 229-236. https://doi.org/10.1016/S1001-0742(08)62256-1
- Palermo, F.F., Risso, W.E., Simonato, J.D. and Martinez, C.B.R., 2015. Bioaccumulation of nickel and its biochemical and genotoxic effects on juveniles of the neo tropical fish *Prochilodus lineatus*. *Ecotoxicol. Environ. Saf.*, **116**: 19-28. https://doi.org/10.1016/j. ecoenv.2015.02.032
- Pane, E.F., Haque, A. and Wood, C.M., 2004. Mechanistic analysis of acute, Ni-induced respiratory toxicity in the rainbow trout (*Oncorhynchus mykiss*): An exclusively branchial phenomenon. *Aquat. Toxicol.*, 69: 11-24. https://doi.org/10.1016/j. aquatox.2004.04.009
- Paul, R., Guite, L.L. and Ramanujam, S.N., 2014.

Copper and Cadmium induced histopathological alterations in liver of *Heteropneustes fossilis* (Bloch) at varying water pH. *Int. J. Fish Aquat. Studies.*, **1**: 38-42.

- Peebua, P., Kruatrachue, M., Pokethitiyook, P. and Singhakaew, S., 2008. Histopathological alterations of Nile tilapia, Oreoechromis niloticus Tilapia zillii and Synodontis schall from EI Salam canal, Egypt. Egypt. J. Aquat. Biol. Fish., 7: 99-138. https://doi. org/10.21608/ejabf.2003.1770
- Perera, P.A.C.T., Kodithuwakku, P.S., Sundarabarathy, V.T. and Edirisinghe, U., 2015. Bioaccumulation of cadmium in freshwater fish. An environmental perspective. *Insight Ecol.*, 4: 1-12. https://doi. org/10.5567/ECOLOGY-IK.2015.1.12
- Pervaiz, A., Afridi, R., Pervaiz, Z., Masood, R. and Pervaiz, H., 2019. Examination of morphological, behavioral and histopathological effects on *Oreochromis niloticus* after acute exposure to methylmercury. J. Innov. Sci., 5: 16-24. https://doi. org/10.17582/journal.jis/2019/5.1.16.24
- Pourahmad, J., Rabiei, M., Jokar, F. and O'brien, P.J., 2005. A comparison of hepatocyte cytotoxic mechanisms for chromate and arsenite. *Toxicology*, 206: 449-460.https://doi.org/10.1016/j.tox.2004.08.002
- Purwanti, I., Arroisi, W., Rahardja, B. and Sulmartiwi, L., 2019. Bioaccumulation and histopathological effect on the gills and liver of silver barb (*Barbonymus* gonionotus) exposed to the heavy metal nickel. *IOP* Conf. Ser. Earth Environ. Sci., 236: 012-098. https:// doi.org/10.1088/1755-1315/236/1/012098
- Qadir, A. and Malik, R.N., 2011. Heavy metals in eight edible fish species from two polluted tributaries (Aikand Palkhu) of the River Chenab, Pakistan. *Biol. Trace Elem. Res.*, **143**: 1524–1540. https://doi. org/10.1007/s12011-011-9011-3
- Rana, M.A., Jabeen, F., Shabbir, S., Naureen, A., Sultana, K., Ahmad, I. and Shabnam, M., 2015.
  Histopathological study of liver and kidney in common carp (*Cyprinus carpio*) exposed to different doses of potassium dichromate. *Int. J. Biol. Sci.*, 6: 108-116.
- Rauf, A., Javed, M. and Ullah, U., 2009. Heavy metals level in three major carps (*Catla catla, Labeo rohita* and *Cirrhina mrigala*) from the river Ravi, Pakistan. *Pak. Vet. J.*, **29**: 24-26.
- Rennika, A. and Nurlita, A., 2013. Concentration and length of exposure of organic and inorganic compounds on the Mujair fish gill network (*Oreochromis mossambicus*) in sub lethal condition. J. Sci. Art. Pomits, **2**: 2337-3520.
- Robert, R.J., 2001. *Fish Pathology*. 3<sup>rd</sup> ed. London: W.B. Saunders.
- Safina, K.M.J., Akhter, M. and Nadeem, S., 2020. Effect of dietary nickel concentrations on the growth and

feed conversion efficiency of fish. J. Entomol. Zool. Stud., 8: 420-424.

- Saleemi, M.K., Tahir, W.M., Abbas, Z.R., Akhtar, M., Ali, A., Javed, T.M., Fatima, Z., Zubair, M., Bhatti, A.S. and Hassan, U.Z., 2019. Amelioration of toxicopathological effects of cadmium with silymarin and milk thistle in male Japanese quail (*Coturnix japonica*). *Environ. Sci. Pollut. Res.*, 26: 21371-21380. https://doi.org/10.1007/s11356-019-05385-7
- Schwaiger, J., Ferling, H., Mallow, U., Wintermayr, H. and Negele, R.D., 2004. Toxic effects of the nonsteroidal anti-inflammatory drug diclofenac. Part I. Histopathological alterations and bioaccumulation in rainbow trout. *Aquat. Toxicol.*, 68:141-150. https://doi.org/10.1016/j.aquatox.2004.03.014
- Shah, N., Khan, A., Ali, R., Marimuthu, K., Uddin, M.N., Rizwan, M., Rahman, K.U., Alam, M., Adnan, M., Jawad, S.M. and Hussain, S., 2020a. Monitoring bioaccumulation (in gills and muscle Tissues), hematology, and genotoxic alteration in *Ctenopharyngodon idella* exposed to selected heavy metals. *BioMed. Res. Int.*, Volume 2020: Article ID 6185231, 16 pages. https://doi. org/10.1155/2020/6185231
- Shah, N., Khisroon, M. and Shah, S.S.A., 2020.
  Assessment of copper, chromium, and lead toxicity in fish (*Ctenopharyngodon idella* Valenciennes, 1844) through hematological biomarkers. *Environ. Sci. Pollut. Res.*, 27: 33259-33269. https://doi. org/10.1007/s11356-020-09598-z
- Shaheen, T. and Jabeen, F., 2015. Effect of various doses of Cr (VI) on survival and growth of *Cyprinus carpio*. *Pak. J. Zool.*, 47.
- Shaheen, T., Akhtar, T., Chughtai, M.I., Khan, M., Pervaiz, K. and Ashraf, M., 2012. Teratological effects of various sublethal concentration of chromium hexavalent [Cr(VI)] on the gills of *Cyprinus carpio. Int. J. Agric. Biol.*, 14: 318–320.
- Soufy, H., Soliman, M.K., El-Manakhly, E.M. and Gaafar, A.Y., 2007. Some biochemical and pathological investigations on monosex Tilapia following chronic exposure to carbofuran pesticides. *Glob. Vet.*, **1**: 45-52.
- Steinhagen, D., Helmus, T., Maurer, S., Michael, R.D., Leibold, W., Scharsack, J.P., Skouras, A. and Schuberth, H.J., 2004. Effect of hexavalent carcinogenic chromium on carp *Cyprinus carpio* immune cells. *Dis. Aquat. Org.*, 62: 155-161. https:// doi.org/10.3354/dao062155
- Stokes, R.M. and Fromm, P.O., 1965. Effects of chromate on glucose transport by the gut of rainbow trout. *Physiol. Zool.*, 38: 202-205. https:// doi.org/10.1086/physzool.38.3.30152831

Subashini, P., Manavalaramanujam, R., Ramesh, M. and

Geetha, N., 2005. Changes in selected biomarkers in freshwater teleost fish, *Cyprinus carpio var. communis* exposed to sublethal concentrations of chromium sulphate toxicity. *J. Environ. Sci. Eng.*, **47**: 65-68.

- Sultana, T., Butt, K., Sultana, S., Al-Ghanim, K., Mubashra, R., Bashir, N., Ahmed, Z., Ashraf, A. and Mahboob, S., 2016. Histopathological changes in liver, gills and intestine of *Labeo rohita* inhabiting industrial waste contaminated water of river Ravi. *Pak. J. Zool.*, 48: 1171-1177.
- Tahir, M.W., Saleemi, K.M., Khan, A., Yousaf, M., Butt, L.S., Siriwong, W., Muhammad, F., Bhatti, A.S. and Qureshi, S.A., 2017. Hematobiochemical effects of cadmium intoxication in male Japanese quail (*Coturnix japonica*) and its amelioration with silymarin and milk thistle. *Toxin Rev.*, 36: 187-193. https://doi.org/10.1080/15569543.2017.1287088
- Tayel, S.I., Yacoub, A.M. and Mahmoud, S.A., 2008. Histopathological and hematological responses to freshwater pollution in the Nile catfish *Clarias* gariepinus. J. Egypt. Acad. Soc. Environ. Dev., 9: 43-60.
- Udotong, J.I.R., 2015. Histopathological changes in liver and muscle of Tilapia fish from QIRE exposed to concentrations of heavy metals. *Int. J. Biol. Biomol. Agric. Food Biotechnol. Eng.*, **9**.
- Usero, J., Izquierdo, C., Morillo, J. and Gracia. I., 2004. Heavy metals in fish (*Solea vulgaris, Anguilla anguilla* and *Liza aurata*) from salt marshes on the southern Atlantic coast of Spain. *Environ. Int.*, **29**: 949-956. https://doi.org/10.1016/S0160-4120(03)00061-8
- Wang, S. and Shi, X., 2001. Molecular mechanisms of metal toxicity and carcinogenesis. *Mol. Cell. Biochem.*, 222: 3-9. https://doi.org/10.1007/978-1-4615-0793-2\_1
- Wijaya, G.S. and Yazid, M., 2009. Microanatomic structure of Ren and nutrition value coefficient (NVC) bio indicator of Tawes (*Puntius javanicus Blkr*) living in integrated ponds of PT. APB Batan. *Proc. Nucl. Saf. Semin.*, pp. 13.
- Yang, X.F., Zhang, H.T., Fan, G.Y., Liu, D.Y., Ge, Y.M., Jiang, J.Q. and Wang, Z.L., 2014. DNA damage of lung cells from immature cadmium ingested mice. *Pak. Vet. J.*, 34: 73-77.
- Yousafzai, A.M., Khan, A.R. and Shakoori, A.R., 2008. Heavy metal pollution in River Kabul affecting the inhabitant fish population. *Pak. J. Zool.*, **40**: 331-339.
- Zamorano, F.X.M., Klingler, C.A.R., Head, J. and Carvan, M.J., 2016. Parental whole life cycle exposure to dietary methylmercury in Zebra fish (*Danio rerio*) affects the behavior of offspring. *Environ. Sci. Technol.*, **50**: 4808-4816. https://doi. org/10.1021/acs.est.6b00223