

EFFECTS OF NaCl AND Na₂SO₄ SALINIZATION ON GERMINATION AND EARLY SEEDLING GROWTH OF FIFTEEN GERMPLASMS OF GUAR (*CYAMOPSIS TETRAGONOLOBA* (L.) TAUB.) – *IN VITRO*

Ishrat Jahan, D. Khan and M. Javed Zaki

Department of Botany, University of Karachi, Karachi- 75270, Pakistan.

ABSTRACT

Germination of fifteen germplasms of guar (four varieties, BR-2/1, BR-90, BR-99 and BR-2017) and 11 lines (S-5733, S-5742, S-5747, S-5759, S-5761, S-5765, S-5784, S-5785, S-5797, S-5798 and S-5825) has been investigated *in vitro* under NaCl and Na₂SO₄ salinities (0 to 10 bar) corresponding to 0.203 to 22.20 dS.m⁻¹ in case of NaCl and 0.203 to 23.0 dS.m⁻¹ in case of Na₂SO₄. Final germination percentage (FGP) was of quite high order reaching ≈ 90 % in several germplasms after four days of incubation in 10 bar NaCl except few germplasms in which germination significantly declined. Sequentially descending order of salt tolerance in terms of increasing *per cent* reduction of final germination in 10 bar NaCl and Na₂SO₄ over control was found to be as follows. FGP was generally more affected under Na₂SO₄.

NaCl (Germination reduction in 10 bar over control):

BR-2/1 (-6.67%) = BR-99 (-6.67%) > S-5785 (-11.11%) > BR-90 (-13.33%) > S-5742 (-17.24%) > S-5761 (-19.99%) ≈ S-5733 (-20.0%) > S-5759 (-24.14%) ≈ S-5747 (-24.99%) > S-5765 (-27.58%) > S-5785 (-33.33%) > S-5797 (-46.67%) ≈ S-5798 (-48.18%) > BR-2017 (-51.85%) > S-5784 (-68.97%)

Na₂SO₄ (Germination reduction in 10 bar over control)

S-5825 (-17.24%) > S-5759 (-31.03%) ≈ S-5747 (-32.14%) ≈ S-5785 (-33.33%) = S-5797 (-33.33%) > BR-99 (-40.0%) ≈ BR-2017 (-40.74%) > S-5798 (-40.74%) ≈ S-5765 (-41.38%) > BR-90 (-46.67%) > BR-2/1 (-50.0%) > S-5761 (-60.00%) > S-5784 (-62.07%) > S-5733 (-66.67%) > S-5742 (-82.76%)

Germination velocity was greatly impeded under both types of salinities. Fifty per cent reduction in GVI on the basis of best fit linear or curvilinear predictive equations associated with in-hand germplasms as follows:

NaCl salinity (50% Reduction in GVI:

S-5784 (6.66 bar) < S-5733 (7.50 bar) < S-5798 (8.55 bar) ≈ S-5742 (8.95 bar) = S-5761 (8.95 bar) < BR-2017 (9.36 bar) < S-5797 (9.92 bar) < S-5765 (10.02 bar) < S-5785 (11.73bar) < S-BR-2/1 (11.83 bar) < S-5759 (12.42 bar) < S-5747 (13.55 bar) < S-5825 (19.56 bar) < BR-90 (20.36 bar) < BR-99 (exhibited promotion by 6.25%).

Na₂SO₄ salinity (50% Reduction in GVI:

S-5733 (5.18 bar) < S-5761 (5.79bar) < BR-2/1 (6.16 bar) ≈ BR-90 (6.27 bar) ≈ S-5742 (6.29 bar) ≈ S-5759 (6.57bar) ≈ BR-99 (6.61bar) ≈ BR-2017 (6.69 bar) < S-5784 (7.34 bar) ≈ S-5797 (7.46 bar) ≈ S-5747 (7.76 bar) < S-5765 (8.74 bar) < S-5825 (9.47 bar) < S-5785 (10.25 bar) < S-5798 (12.49 bar)

Root and shoot length and cumulative seedling weight declined with salinity. Fifty per cent reduction in seedling weight in 10 bar salinities associated with the germplasms as given below:

NaCl salinity (Promotion / Reduction in seedling weight in 10 bar over control:

S-5733 (+163.33%) > S-5785 (+51.16%) > S-5825 (+ 28.57%) > BR-99 (+21.95%) ≈ S-5742 (+ 21.74%) > BR-2/1 (+14. 38%) > S-5759 (-8.24%) > S-5747 (-9.18%) > S- 5766 (-17.68%) > S-5797 (-18.60%) > BR-90 (-23.08%) > S-5784 (-39.39%) > S-5761 (-57.30%) > BR-2017 (-62.15%) > S-5798 (-70.19%)

Na₂SO₄ salinity (Promotion / reduction in seedling weight on 10 bar over control:

S-5733 (+ 192.22%) > S-5785 (+12.79%) > S-5825 (+6.72%) > BR- 90 (-5.98%) > BR- 2/1 (-7.29%) > S-5765 (-12.71%) > S-5797 (-17.83) > S-5761 (-20.22%) ≈ BR-99 (-21.19%) ≈ S-5742 (-22.46%) > S- 5747 (-30.61%) ≈ S-5759 (-30.77%) > BR-2017 (-49.72%) > S-5784 (-61.04 > S-5798 (-74.04%).

Agglomerative clustering of germplasms at dissimilarity distance of 5% on the basis of FGP, GVI and seedling dry weight in two type of salinities indicated 5, 2 and 8 groups of germplasms in NaCl and 9, 5 and 7 groups in Na₂SO₄, respectively. Obviously the amplitude of behavioural dispersion in Na₂SO₄ salinity was wider than in NaCl. The extracted groups were differentially tolerant to salinity. Since, there was variation in salt tolerance of germplasms on the basis of parameters studied; a composite performance index was designed according to which there were three groups each in NaCl and Na₂SO₄ salinities.

CPI -NaCl salinity:

- I. Germplasms with CPI: 70-80 (S-5761, S-5785 and S-5797)
- II. Germplasms with CPI: 81-90 (S-5825, BR-2017, BR-90, S-5798, BR- 2/1, and BR-99)
- III. Germplasms with CPI: 98-103 (S-5747, S- 5742, S-5784, S- 5759, S-5733 and S-5765) – relatively more tolerant.

CPI - Na₂SO₄ salinity:

- I. Germplasms with CPI: 71-79 (S-5785, BR-99, S-5798, S-5797, BR-2017, BR- 2/1). All varietal germplasms except BR-90 entered the composition of this group.
- II. Germplasms with CPI: 84-88 (BR-90, S-5825, S-5765, S-5761, S-5784
- III. Germplasms with CPI: 95-96 (S-5733, S-5747, S-5742) – obviously, this group was relatively more tolerant.

On the basis of promotion or reduction of CPI in Na₂SO₄ over NaCl following generalizations may be made:

- Germplasms showing CPI reduction: - 0.82 to -1.95% (BR- 90, S-5742 and S-5747). These germplasms were more or less equally tolerant to NaCl and Na₂SO₄ salinities.
- Germplasms with CPI reduction: -5.23 to - 6.95% (S-5797, S-5733, S-5785, and BR-2017)
- Germplasms with CPI reduction: -11.08 to - 15.48% (BR- 2/1, S-5784, S-5759, S- 5798 and S-5765) – more sensitive to Na₂SO₄ than above groups.
- Germplasms with CPI reduction: -20.99% (BR-99) – most sensitive to Na₂SO₄ amongst the tested germplasms.
- Germplasms with CPI promotion: + 2.58 to +17.06% (S-5825 and S-5761, respectively) – most tolerant to Na₂SO₄ salinity which was promotory to them over NaCl- particularly S-5761.

The results are discussed in view of the available literature pertaining to guar.

Key Words: Guar (*Cyamopsis tetragonoloba* (L.) Taub., Pakistan's guar germplasms, NaCl and Na₂SO₄ salinity, Germination, Germination velocity, agglomerative cluster analysis, Salinity tolerance

INTRODUCTION

Guar or cluster bean (*Cyamopsis tetragonoloba* (L.) Taub.), is a legume crop, grown for three major purposes primarily as cattle forage, as green manure and raw material for industry (AlShameri *et al.*, 2017). It is expected that genus *Cyamopsis* originated from the genus *Indigofera* due to aneuploidy (Senn, 1938). Guar is mainly grown in India (80% of the World production), Pakistan (15%), Sudan and USA (Sharma, 2010). Guar grows best in sandy soil (Raychaudhuri, 1952). It is an excellent soil-binding crop due to potential *Bradyrhizobium* nodulation (ElSheikh and Ibrahim, 1999). Being a valuable crop economically, it has been extensively studied for various biological aspects and its agronomic characteristics (to cite a few: Whistler and Hymowitz, 1979; Omer *et al.*, 1993; Lahiri *et al.*, 1996; Afria *et al.*, 1998; Ashraf *et al.* 2002; Weixin, 2003; Chaudhary *et al.*, 2007 a and b; Morris, 2010; Pathak, 2010; Rao and Shahid, 2011; Rai *et al.*, 2012; Sultan *et al.*, 2012; Singh, 2014; Deepika and Dhingra (2014); Khalid *et al.*, 2017; Muftahizade *et al.*, 2017; Shehata *et al.*, 2017; Amin *et al.*, 2018; AlShameri *et al.*, 2017; 2019). Deepak *et al.* (2003) have reported regeneration of guar shoot from cotyledonary nodes. Amin *et al.* (2018) reported that it may grow in Pb-accumulated soil (1000mg/ kg soil). Gresta *et al.* (2018) tested nine guar genotypes (India1, India2, India3, Kinman, Lewis, Metador, monument, etc.) from India, Pakistan, USA and South Africa in Sicily, Italy for germination capability and speed in response to constant temperature (5 to 35 °C, with 5 °C increments) and two alternating temperature 15/10 °C and 20/15 °C. Optimal temperatures were 30 and 35 °C but Indian genotype also demonstrated substantial germination percentage (33-43%) at constant temperature as low as 15 °C. Final germination percentages in all genotypes increased with temperature from 10-35 °C except Pakistan germplasm which showed decline in germination at higher temperature of 35 °C. Germination time was generally higher at low temperature. Germination significantly improved when seeds were exposed to 20 °C for 6h a day. Earlier, in another paper, Sortino and Gresta (2007) evaluated growth and yield of five cultivars in a Mediterranean environment. Singla *et al.* (2016) studied planting time of guar in semi-arid Southern plains of USA and found that delaying planting beyond mid-June is detrimental to guar productivity. Guar may survive in fairly high salinity (Mehta and Desai, 1953; Yadava *et al.*, 1974). Many guar genotypes have been tested for salt tolerance in recent years, and some of them have turned out to be indeed salt tolerant. The potential of some guar accessions for saline soils was tested by Teolis *et al.* (2009) by evaluating seed germination of 42 accessions of guar in sodium chloride solution. The seeds were germinated in 200 mM NaCl or double-distilled water (control). There were significant differences in salt tolerance among the accessions tested. Percentages of seeds germinating in the salt solution ranged from 7.7 to 90.3% of the control, with the higher percentages suggesting greater salt tolerance. Twelve of the forty-two accessions were further selected to study the effect of salinity on plant growth. Significant differences were obtained between the accessions for seed yield per pod, but not for seed yield per plant, plant height, branch number, and pod number (percentage of control). The presence of variation in the germplasm suggests some potential for selecting cultivars with increased salt tolerance. Salinity studies should include both germination and plant trials, because germination assays for salt tolerance do not necessarily predict plant growth and yield in saline soils.

Abdou *et al.* (2017) studied the effects of Compost and some bio-stimulants treatments on guar. Khafagy *et al.* (2010) have reported that presoaking of guar seeds in selected phytohormones (ABA or SA) and vitamins (Ascorbic acid and Thiamine) exhibited significant effect on seed germination as well as seedling growth under both normal and NaCl saline conditions compared to control. Generally, phytohormone (Salicylic acid) and plant vitamin (Ascorbic Acid or Thiamine) counteracted the harmful effect of salinity on guar seedling growth. Jat *et al.* (2015) studied two varieties of guar ('Neelam – 51' and 'Naveena') for seed quality enhancement by seed priming. Kumar *et al.* (2017) employed conventional and biotechnological approaches for genetic improvement of cluster bean and Arora and Pahuja (2008) employed chemical and physical mutagens to extend the range of genetic diversity of guar plants but economically important traits were not found in mutants. Ambika and Balakrishnan (2015) investigated the effects of cow urine priming of guar seeds on germination and seedling growth.

Some genotypes of guar from Pakistan were investigated by Rashid *et al.* (2015) using agronomic traits (shoot fresh biomass, shoot dry biomass and plant height) as well as stress tolerance indices of these traits). At adult vegetative growth, genotypes 41671, Khushab White, 5597, 24320, Sillanwali White, 24321, Mardan White, Br-99,

Karor White, Hafizabad White, BWP-5611 and Klorkot Black were rated as salt tolerant, while 24323, Khanewal Local 2, Kalorkot White, BWP5589, Chiniot White, 27340, BWP-5599, Bhowana White and BWP-5596 were rated as highly salt sensitive. A significant genotypic variation was found amongst the 31 genotypes tested. They opined that further study is necessitated to elucidate physiological basis of salt tolerance in guar (Rasheed *et al.*, 2015). Alshameri *et al.* (2017) have tested drought and salt tolerance of some guar germplasms and reported that accessions BWP 5595, 24320, Chiniot 1, Chiniot 2, Kaloorkot 2, and BWP 5599 were more drought tolerant than BR-99, BR-90, 027340, 24333, 24332, Khanewal Local 2, and Bhawana 2. Accessions 24320, BWP 5595, Chiniot 1, Chiniot 2, and 22159 were considered to be more salt-tolerant than BWP 5589, 24333, Bhowana 2, 24287, 027340 and BWP 5596. Overall, BWP 5595 and 24320 were found to be drought- and salt-tolerant, respectively, while Khanewal Local 2 and BWP 5589 were drought-sensitive and salt-sensitive, respectively. Thirty six accessions of guar from Pakistan were tested by Ali *et al.* (2015) for enhanced forage production on hot dry lands of Pakistan. Andrade (1985) worked for salt tolerance of three varieties of guar (Kinman, RGC 518 and Santa Cruz. Percent germination of seeds of the cultivars decreased in the media from 5bar to 15 bar. Compared to the control, germination started declining from 5 bar and was minimal around 20-25% in 15 bar medium. Kinman was relatively less affected.

Khan *et al.* (1989) reported that threshold EC_{iw} in case of Lahiya (Punjab) accession of guar with amended Seawater dilution inducing 50% reduction in seed production was 5.76dS.m⁻¹. They found that salinity decreased number of pods and seeds per plant. The number of seeds per pod remained unaffected (7- 8 seeds per pod). All cations (Na, K, Ca and Mg) were more allocated to leaf. Abusuwar and Abbaker (2009) *in vitro* investigated forage species including guar for germination in various dilutions of Red Seawater (control = 0.4 dS.m⁻¹, and the treatments- 3.095, 5.54, and 16.57 dS.m⁻¹ salinity).

Guar is grown with seeds and in this respect germination studies have been undertaken by many workers around the World. To cite a few – Yadava *et al.* (1974), Andrade (1985); Datta and Dayal (1988); Vinisky and Ray (1988); Francois *et al.* (1989); Kumar *et al.* (1988); Miyamoto (1988); Khan *et al.* (1989); Kumar *et al.* (1990); Chunmei *et al.* (2002); Teolis *et al.* (2009); Francois and Kleiman (1990); Khanzada *et al.* (2001); Ashraf *et al.* (2005); Khafagy *et al.* (2010); Rasheed *et al.* (2015); Iqbal (2015); Gul *et al.* (2015); Sambangi and Rani (2016); Alshameri *et al.* (2017); Joshi and Datta (2017); Bina and Bostani (2017); Zheng *et al.*, 2017; Gresta *et al.* (2018); Sudhar *et al.* (2018 a & b); Prakash *et al.* (2019), etc. Guar collection founded in the Institute of Plant Genetic Resources (VIR), Russia, by N.I. Vavilov lists more than 100 accessions. Germination of guar seeds from VIR collection, after 40-year storage at the room temperature, remained high. The research data and the practice of guar growing in Russia testify that the crop could be successfully cultivated in the southern regions of Russia (Dzyubenko *et al.*, 2017).

In present studies, the effects of NaCl and Na₂SO₄ salinities (0 to 10 bar) have been *in vitro* investigated on germination of 15 guar germplasms of Pakistan.

MATERIALS AND METHODS

Freshly collected seeds from the crops of guar, four varieties - BR-2/1, BR-90, BR-99 and BR-2017 and eleven lines including S-5733, S-5742, S-5747, S-5759, S-5761, S-5765, S-5784, S-5785, S-5797, S-5798 and S-5825 were received from Agricultural Research Station, Bahawalpur by the courtesy of Dr. Lal Hussain Akhtar. The biological traits of the guar germplasms employed in this study are presented in Table 1. Germplasms- BR-2/1, BR-90, and BR-99 were released in 1984, 1991 and 2000, respectively whereas var. BR-2017 is recently released. Other germplasms are the accessions collected by the Agricultural Research station and experimented for various agronomic aspects. Germplasms BR-99 and BR-2017 and accessions such as S-5733, S-5747, S-5761, S-5765 and S-5785 are quite high-yielding. All germplasms are cultivated for fodder. S-5733 is an advance line under testing at Agricultural Research Station, Bahawalpur, Pakistan. The seeds were stored at room temperature in dry cabinet not more than 20 days. Longer storage of guar seeds was observed to cause dormancy due to dry hardy seed coat. They are reported to germinate even after 40-year storage (Dzyubenko *et al.*, 2017). Healthier seeds with suitable weights were screened for their salt tolerance (see Khan *et al.*, 2018). The selected seeds were surface sterilized using 1% bleach (NaOCl) and rinsed with sterilized distilled water. Ten seeds of each variety/line were placed on Whatman filter paper in sterilized 9 cm Petri plates. NaCl and Na₂SO₄ solutions were used to investigate the effect of salt stress on guar germination. Ten aqueous concentrations (1 to 10 bar) of NaCl and Na₂SO₄ (Meiri *et al.*, 1971) were prepared in sterilized distilled-deionized water. For the purpose to facilitate comparison, Electrical conductivities of the prepared solutions were measured on EC meter and expressed as (dS.m⁻¹) which further ascertained their salinity levels (Fig. 1).

Each Petri plate containing seeds was irrigated with five mL of aqueous solution of NaCl or Na₂SO₄ (1 to 10 bar) and plates were incubated at 28-30 °C in a growth chamber to germinate. The optimum temperature for guar germination is reported to 25-30°C (Zheng *et al.*, 1980). The sterilized distilled water was used as control for each germplasm. Each treatment had three replicates. Seed germination was recorded every 24 h (AOSA, 1990) until four days. A Seed was considered germinated when its radicle showed at least 2 mm of growth. On 4th day, root length, shoot length and dry weight of healthy seedlings were recorded discarding the seedlings which died (particularly in

high concentration of Na_2SO_4). Promotion or reduction was calculated as: treatment - control / control expressed in percent. There are several parameters or indices of germination employed in germination studies (Ranal *et al.*, 2009; Aflaki *et al.*, 2017). It is opined by Al- Mudaris, 1998) that one single parameter is in itself not sufficient to fully describe germination. The 'final germination percentage' is an end phase parameter which only reflects the capacity of a seed lot to reach germination. Since it does not reflect Speed, synchrony or spread of germination - all vital factors from horticultural and agronomic standpoints- it should be accompanied by a measure of germination velocity. In present studies the germination Velocity Index (GVI) was calculated following Woodstock (1976) - $\text{GVI} = \text{N1}/1 + \text{N2}/2 + \text{N3}/3 + \text{N4}/4 + \dots + \text{Nn}/\text{n}$, where N1 , N2 , N3 , N4 ... Nn are the number of new germinants on a particular day of the test and 1, 2, 3, 4...n are the number of days of observation. GVI is higher if more seeds germinate in lower number of days.

Seed germination involves not only qualitative responses of individual seeds but also population responses which are distributed over time. Standard analysis of variance or regression methods are appropriate when some viable seeds fail to germinate (Scott *et al.*, 1984). Beside variance and regression analyses of the data, varieties /lines were agglomerated in form of a cluster diagram (Ward, 1963) by calculating Euclidean distances on the basis of germination, GVI and seedling dry weight to investigate inter-germplasm similarity of behavior against NaCl and Na_2SO_4 salinities. The technique of cluster analysis has been successfully used by Khalid *et al.* (2017) to delimit clusters on the basis of various agronomic traits amongst 100 accessions of guar from Pakistan into eight clusters of which cluster I (Accessions - 5778, 22267, 22229, 5597, 5588, 5590, 5596, 22228, 5765, 22165, 5789, BR-99, 6036, 6056, 5557, 5752, 5885, 5824, 5823, 5825, 5747, 5601, 5743, 5733 and 21897) and cluster V (6500, 6498, 6499, 6497 and 6496) were considered to be the best performer as regards to grain yield, plant height, number of pods.plant⁻¹ and number of grains.pod⁻¹ having compact plant type.

Since germination, velocity of germination and the seedling growth varies amongst the species and various cultivars and accessions of a species (Asana and Kale, 1965; Maliwal and Paliwal, 1970), a composite performance index (CPI) was calculated as:

$$\text{CPI} = \frac{\text{Final mean germination all over the Treatment Range} + \text{Mean velocity of germination all over the Treatment Range} + \text{Seedling dry mass at the last day of incubation all over the Treatment Range (0 to 10 bar)}}{3}$$

To compare CPI in two types of salinities, promotion or reduction of CPI in two salinities was calculated as:

$$\begin{aligned} \text{\% CPI Promotion or Reduction} &= \frac{\text{CPI}_{\text{Na}_2\text{SO}_4} - \text{CPI}_{\text{NaCl}}}{\text{CPI}_{\text{NaCl}}} \times 100 \text{ and} \\ \text{\% CPI Promotion or Reduction} &= \frac{\text{CPI}_{\text{NaCl}} - \text{CPI}_{\text{Na}_2\text{SO}_4}}{\text{CPI}_{\text{Na}_2\text{SO}_4}} \times 100 \end{aligned}$$

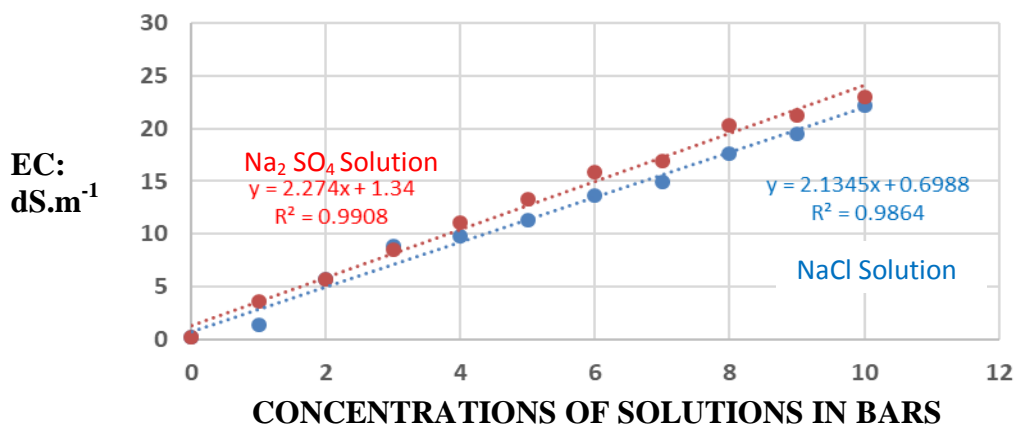


Fig. 1. Relationship of ECs of NaCl and Na_2SO_4 solutions prepared in concentrations expressed in bar following Meiri *et al.* (1971). For ease and convenience bar values were expressed as positive number bearing "bar" as suffix.

RESULTS AND DISCUSSION

Relationship between bar concentrations of NaCl and Na₂SO₄ solutions with their electrical conductivities:

There was statistically highly significant positive correlation between bar and EC units of concentration of NaCl and Na₂SO₄ solutions (Fig. 1). EC values for different bar concentrations of Na₂SO₄ were slightly higher than EC values for NaCl solutions. The EC of NaCl solutions (1 to 10 bar) corresponded to 1.322 -22.20dS.m⁻¹ and that of Na₂SO₄ corresponded to 3.58 to 23.0 dS.m⁻¹. Control solutions had EC value equal to 0.203dS.m⁻¹. The equations posted on the curves (Fig.1) may be useful in estimating EC values on the basis of bar concentrations of these salts. It may be mentioned that the osmotic potentials of salt solutions decreases as their concentrations increase. Osmotic potential when represented in bar, they are shown with negative numbers. Higher bar values indicate proportionately lower osmotic potential, with respect to the plant, of a solution i.e., more saline in terms of electrical conductivity of the solutions and consequently lesser and lesser availability of water to seeds, seedlings or plant (see Carrow and Duncan, 2017). Rangaswamy (2010) reported that soil salinity of 25 dS.m⁻¹ resulted in an osmotic potential of -9 bar which greatly reduced plant available water in case of wheat with 89% to 96% of the field capacity unavailable for plant uptake.

Germination

Freshly collected seeds of various guar germplasms exhibited no dormancy in the present experiment. They germinated quite rapidly and synchronously under control conditions. Germination became assynchronized as salinity increased. The final germination percentage (FGP) under NaCl and Na₂SO₄ salinity is presented in Fig. 2 and 3, respectively. FGP was generally of high order reaching to 90-100% in majority of germplasms after four days of incubation under control as also reported by Thorner (1909) and Ganai Al Awad El-Daw (1998) for guar. Thorner (1909), however, stated that guar seed germination percentage with an initial value of 98% can drop to 2% when submerged in water for 38 days.

FGP of germplasms BR-2017, S-5765 and S-5797 significantly declined under high salinity of NaCl (10 bar) (Fig. 2). There was comparatively larger inhibition of FGP under Na₂SO₄ salinity (Fig. 3). The mean FGP of germplasms under 10 bar concentration and over the entire range of the treatments and control for both salts is presented in Table 2a. FGP of guar germplasms in 10 bar NaCl averaged to 68.88 ± 5.0% which was significantly higher than that (53.78 ± 4.02%) in Na₂SO₄ ($t = 2.24$, $p < 0.05$). Under 10 bar NaCl FGP was higher than the average FGP in 9 germplasms (BR-2/1, BR-90, BR-99, S-5733, S-5742, S-5747, S-5765, S-5759 and S-5825) and was lesser than the average FGP in six germplasms (BR-2017, S-5761, S-5784, S-5797, S-5798 and S-5785). The germination in 10 bar NaCl was maximum in BR-2/1 (93.33 ± 3.33%) and BR-99 (93.33 ± 3.33%) and minimum in S-5784 (30 ± 15.28%) followed by BR-2017 (43.33 ± 8.82%).

FGP on 4th day in 10 bar Na₂SO₄ was higher than the average FGP in 8 germplasms (BR-99, S-5747, S-5759, S-5761, S-5784, S-5885, S-5797 and S-5825), lower than the average FGP in four germplasms (BR-2/1, S-5633, S-5742 and S-5765) and at par with the average FGP in three germplasms (BR-90, BR-2017 and S-5798).

FGP on the fourth day over the entire range of treatments (0 to 10 bar) was also higher in NaCl (88.52 ± 1.62) than that in Na₂SO₄ (82.21 ± 1.39) ($t = 2.96$, $p < 0.05$). FGP over the entire range of NaCl treatments was the maximum in BR-99 (96.97 ± 1.14%) and the minimum in BR-2017 (75.46 ± 4.88%). In NaCl, nine germplasms had higher FGP than the grand mean and six germplasms had lower FGP than the grand mean. In Na₂SO₄ environment maximum FGP was exhibited by BR-90 (91.82 ± 4.13) and minimum by S-5797 (75.45 ± 4.63). Seven germplasms had FGP higher than the grand mean and eight germplasms had lower FGP than the grand mean.

The adverse effects of salinity on germination are well-known even in drought tolerant species (Khan *et al.*, 1984; Myers and Morgon, 1989; Ismail, 1990; De Villiers *et al.*, 1994; Shaukat and Burhan, 2000). Our results are more or less in agreement with Datta and Dayal (1988) who investigated 25 genotypes of guar (CG-1, HGS-25, HGS-18, CG-4, PLG-174, HFG-314, DSE-IJ, HGS -46, PLG-85, PLG-119, HGS-47, DSE -16J, HFG-156, PNB, FS-277, HGS-43, HFG-119, Guara, HG-182, HGS-75, HFG-189, Suvidha, HGS-3, HG-17-1 and Hg-258) for salinity tolerance under composite salts (NaCl, CaCl₂, MgCl₂ and MgSO₄ at 30 ± 1 °C in Petri dishes irrigated with 10 mL solution of EC: 0, 4, 8, 12, or 16dS.m⁻¹. The final germination level didn't vary much with salinity level in their experiment also. Similarly, Zucchini (*Cucurbita pepo* L.) has been reported to germinate more than 90% at 40dS.m⁻¹ of NaCl (Bina and Bostani, 2017). Higher salinities differentially reduced germination in all varieties tested of *Vigna unguiculata* (Thiam *et al.* (2013), rice cultivars (Zafar *et al.*, 2015) and maize cultivars (Hassan *et al.*, 2018).

Table 1. Morphological traits of some guar varieties / lines. *

| Variety/ Lines | Year of Release | Yield Potential (Kg ha ⁻¹) | Salient Features |
|-------------------|--------------------------------------------|--------------------------------------------------------------------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| BR-2/1 | 1984 | Grain Yield = 1200 Kg ha ⁻¹ Fodder Yield = 26 t ha ⁻¹ | Hairy, Long stature, long duration, branched, grain bold & dark brown, suitable for fodder purpose. |
| BR-90 | 1991 | Grain Yield = 1400 Kg ha ⁻¹ Fodder Yield = 28 t ha ⁻¹ | Hairy, short stature, long duration, profusely branched, golden grain colour, suitable for fodder & grain purpose |
| BR-99 | 2000 | Grain Yield = 1900 Kg ha ⁻¹ Fodder Yield = 30 t ha ⁻¹ | Hairy, Single stemmed, no branching, medium duration, seed colour-grey, tolerant to sucking pests and diseases, suitable for grain, fodder and vegetable purposes. |
| BR-2017 | 2017 | Grain Yield = 2400 Kg ha ⁻¹ Fodder Yield = 35 t ha ⁻¹ | Hairy, erect type, 0-1 branch, higher gum & protein contents, early maturing and short duration variety with heavy fruiting, requires very low inputs, tolerant to sucking pests and diseases, suitable for grain, fodder and vegetable purposes. |
| S-5274 | - | | Approved as BR-2017 |
| S-5733 | Advance line under testing in yield trials | Grain Yield = 1800 Kg ha ⁻¹ Fodder Yield = 29 t ha ⁻¹ | Hairy, erect type with 1-3 branches, long duration, tolerant to insect pests & diseases suitable for fodder and seed purposes. |
| S-5742 | -do- | Grain Yield= 1750 Kg ha ⁻¹ Fodder Yield= 32 t ha ⁻¹ | Hairy, single stemmed with no branch, early maturing, tolerant to insect pests & diseases, suitable for seed and vegetable purposes. |
| S-5747 | -do- | Grain Yield= 1900 Kg ha ⁻¹ Fodder Yield= 26 t ha ⁻¹ | Hairy, erect type with no branch, short duration, tolerant to insect pests & diseases, suitable for seed and vegetable purposes. |
| S-5759 | -do- | Grain Yield= 1700 Kg ha ⁻¹ Fodder Yield= 27 t ha ⁻¹ | Hairy, erect type with no branch, short duration, tolerant to insect pests & diseases, suitable for seed and vegetable purposes. |
| S-5761 | -do- | Grain Yield= 1900 Kg ha ⁻¹ Fodder Yield= 30 t ha ⁻¹ | Hairy, 0-1 branch, early maturing & short duration, tolerant to insect pests & diseases, suitable for seed, fodder and vegetable purposes. |
| S-5765 | -do- | Grain Yield= 2000 Kg ha ⁻¹ Fodder Yield= 34 t ha ⁻¹ | Hairy, no branch, erect type, early maturing & short duration, tolerant to insect pests & diseases, suitable for seed, fodder and vegetable purposes. |
| S-5784 | -do- | Grain Yield= 1800 Kg ha ⁻¹ Fodder Yield= 24 t ha ⁻¹ | Hairy, non-branched, early maturing, tolerant to insect pests & diseases, suitable for seed purpose. |
| S-5785 | -do- | Grain Yield = 2200 Kg ha ⁻¹ Fodder Yield = 22 t ha ⁻¹ | Hairy, branches 2-4, long duration, tolerant to insect pests & diseases, suitable for seed purpose. |
| S-5797 | -do- | Grain Yield= 1400 Kg ha ⁻¹ Fodder Yield= 34 t ha ⁻¹ | Hairy, branches 6-10, long duration, tolerant to insect pests & diseases, suitable for fodder purpose. |
| S-5798 | -do- | Grain Yield = 1300 Kg ha ⁻¹ Fodder Yield = 33 t ha ⁻¹ | Hairy, branches 8-10, long duration, tolerant to insect pests & diseases, suitable for fodder purpose. |
| S-5825 | -do- | Grain Yield = 1500 Kg ha ⁻¹ Fodder Yield = 32 t ha ⁻¹ | Hairy, branched, long duration, tolerant to insect pests & diseases, suitable for fodder purpose. |

*, Courtesy – Regional Agricultural Research Station, Bahawalpur, Pakistan.

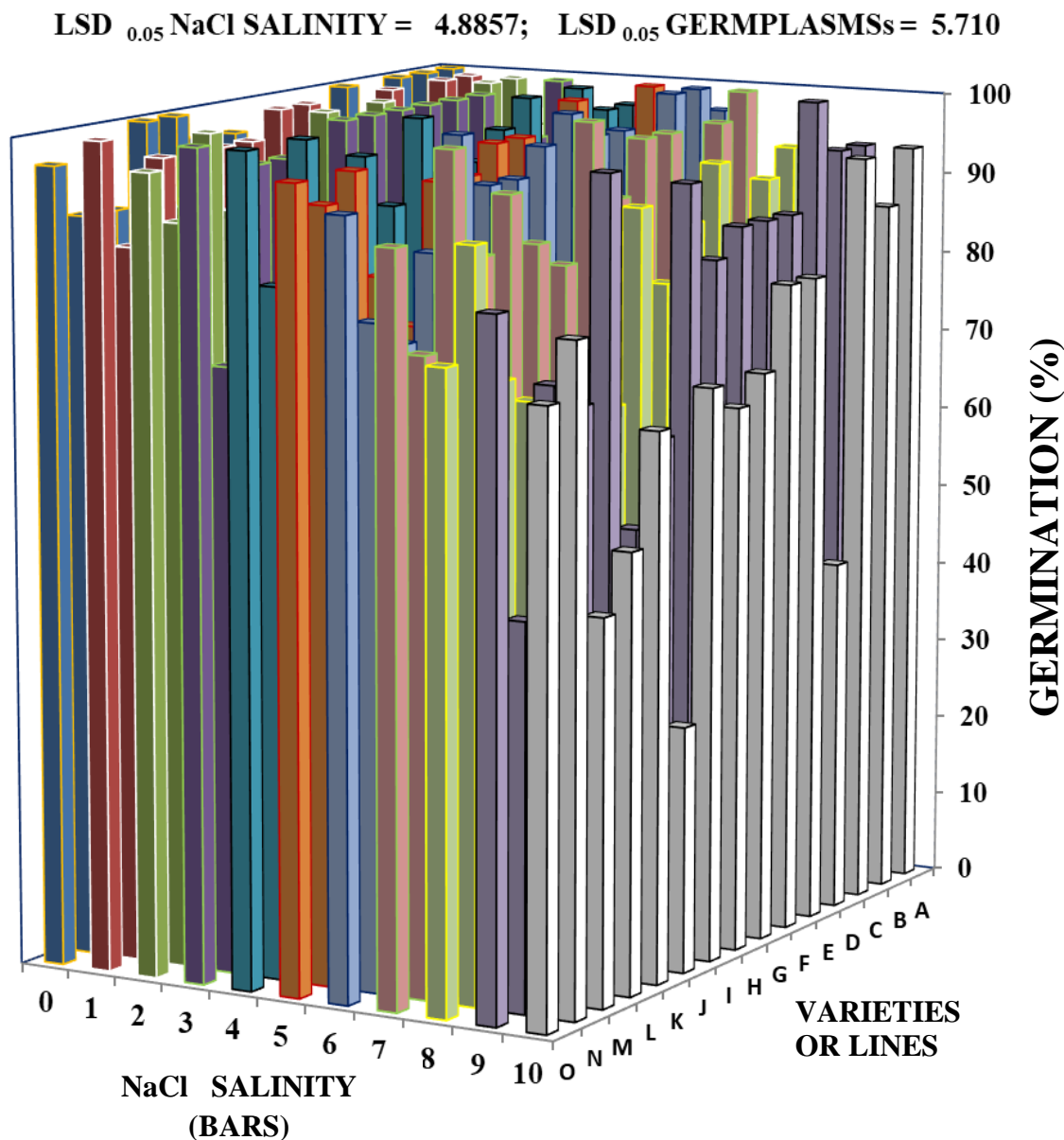


Fig. 2. Per cent final mean germination of Guar varieties / lines under influence of NaCl salinization (0-10 bars).

Varieties / Lines: A, BR-2/1, B, BR-90, C, BR-99, D, BR-2017, E, S-5733, F, S-5742, G, S-5747, H, S-5759, I, S-5761, J, S-5765, K, S-5784, L, S-5785, M, S-5797, N, S-5798 and O, S-5825.

The sequential order of FGP of various germplasms in descending order of relative tolerance to NaCl and Na₂SO₄ salinities is given in Table 2b. **Clearly, BR-2/1, BR-99 and BR-90 were highly tolerant to high NaCl salinity (10 bar) but not to Na₂SO₄.** In Na₂SO₄, S-5825 had the highest FGP. Over the entire range of NaCl treatments, BR-99, S-5733, S-5742, BR-2/1, BR-90, S-5747 and S-5761 were comparatively more tolerant. In Na₂SO₄ treatment range, BR-90 and S-5747 exhibited > 90 % germination. The local in-hand germplasm of guar appeared to be more tolerant to NaCl than to Na₂SO₄.

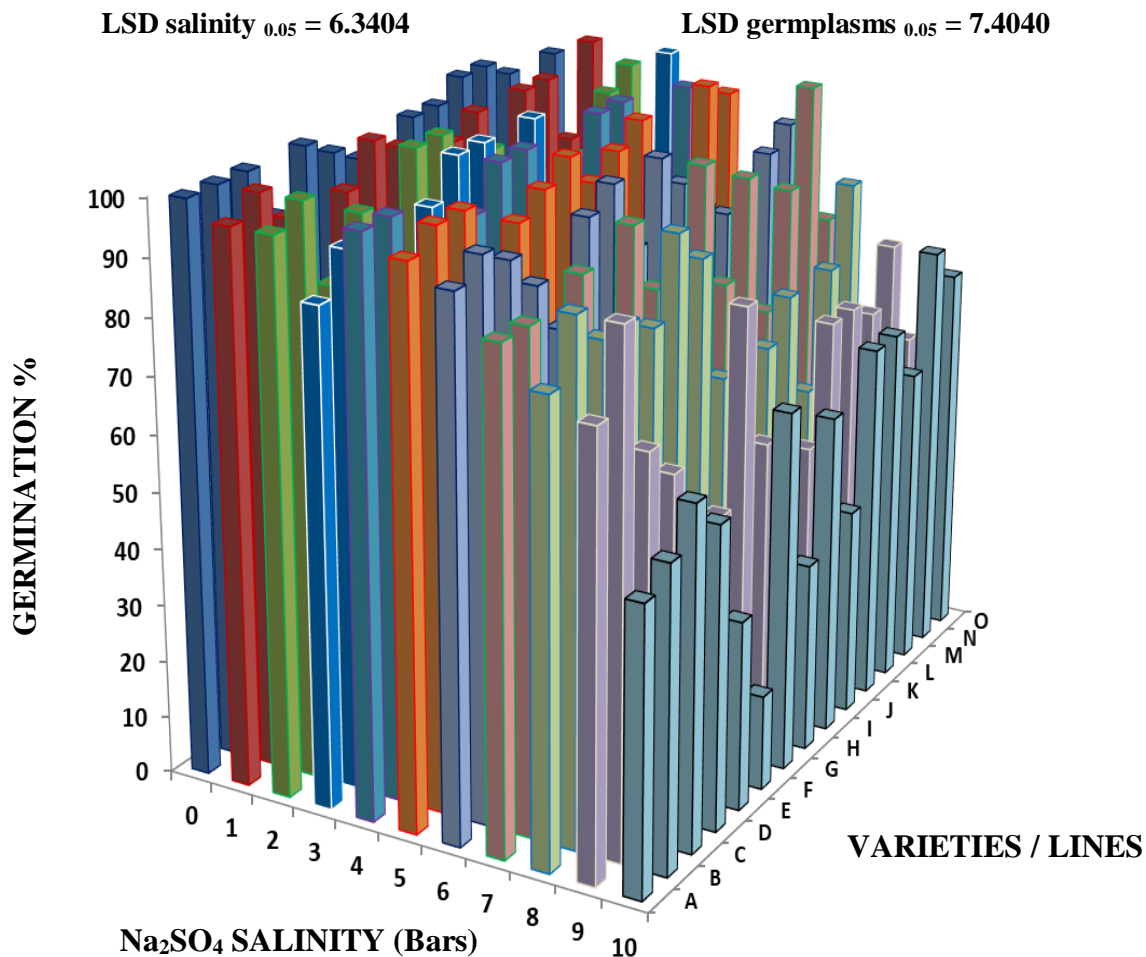


Fig. 3. Per cent final mean germination of Guar varieties / lines under influence of Na₂SO₄ salinization (0-10 bars). Varieties / Lines: A, BR-2/1, B, BR- 90, C, BR- 99, D, BR-2017, E, S-5733, F, S-5742, G, S-5747, H, S-5759, I, S-5761, J, S-5765, K, S-5784, L, S-5785, M, S-5797, N, S-5798 and O, S-5825.

Table 2a. Mean Final germination (%) of guar cultivars and lines under salinity.

| Genotype | Final germination (Day 4) in 10 Bar salinity | | Final germination over the treatment range (0 to 10 Bar) | |
|----------|----------------------------------------------|---------------------------------|-----------------------------------------------------------|---------------------------------|
| | NaCl | Na ₂ SO ₄ | NaCl | Na ₂ SO ₄ |
| BR-2/1 | 93.33 ± 3.33 | 50.00 ± 5.77 | 92.72 ± 1.671 | 85.58 ± 4.516 |
| BR-90 | 86.67 ± 13.33 | 53.33 ± 24.04 | 92.27 ± 1.335 | 91.82 ± 4.129 |
| BR-99 | 93.33 ± 3.33 | 60.00 ± 11.55 | 96.97 ± 1.142 | 83.34 ± 4.669 |
| Br-2017 | 43.33 ± 8.82 | 53.33 ± 6.67 | 75.46 ± 4.88 | 80.91 ± 3.973 |
| S-5733 | 80.00 ± 0.00 | 33.33 ± 16.67 | 94.85 ± 2.03 | 78.18 ± 7.126 |
| S-5742 | 80.00 ± 10.00 | 16.67 ± 16.67 | 93.33 ± 1.85 | 85.15 ± 8.267 |
| S-5747 | 70.00 ± 5.77 | 63.33 ± 21.86 | 91.21 ± 2.925 | 90.61 ± 3.591 |
| S-5759 | 73.33 ± 3.33 | 66.67 ± 20.28 | 91.52 ± 2.89 | 78.18 ± 6.52 |
| S-5761 | 66.67 ± 12.02 | 56.67 ± 3.33 | 91.52 ± 2.96 | 77.27 ± 5.875 |
| S-5765 | 70.00 ± 25.17 | 36.67 ± 13.33 | 87.88 ± 4.387 | 84.25 ± 5.845 |
| S-5784 | 30.00 ± 15.28 | 63.33 ± 6.67 | 82.12 ± 6.579 | 78.48 ± 6.580 |
| S-5785 | 66.67 ± 3.33 | 63.33 ± 6.67 | 88.49 ± 3.633 | 76.06 ± 4.920 |
| S-5797 | 53.33 ± 12.02 | 63.33 ± 3.33 | 82.43 ± 4.072 | 75.45 ± 4.633 |
| S-5798 | 46.67 ± 8.82 | 53.33 ± 8.82 | 86.36 ± 5.133 | 81.21 ± 4.387 |
| S-5825 | 80.00 ± 10.00 | 73.33 ± 6.67 | 80.91 ± 3.92 | 86.67 ± 2.880 |
| Mean | 68.88 ± 5.04 | 53.78 ± 4.02 | 88.56 ± 1.62 | 82.21 ± 1.39 |

Table 2b. Sequential order of final germination of guar germplasms in 10 bar NaCl and 10 bar Na₂SO₄ and also over the treatment range inclusive control. Germination is shown in parenthesis (see Table 2a).

| | |
|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|--|
| Final Germination (%) in 10 bar NaCl | |
| BR-2/1 (93.3) = BR-99 (93.3) ≈ BR-90 (86.7) > S-5733 (80.0) = S-5742 (80.0) = S-5825 (80.0) > S-5759 (73.3) > S-5747 (70.0) = S-5765 (70.0) > S-5761 (66.67) = S-5785 (66.67) > S-5797 (53.33) > S-5798 (46.67) > BR-2017 (43.33) > S-5784 (30.0) | |
| Final germination 10 bar Na ₂ SO ₄ | |
| S-5825 (73.3) > S-5759 (66.7) > S-5747 (63.3) = S-5784 (63.3) = S-5785 (63.3) = S-5797 (63.3) > BR-99 (60.0) > S-5761 (56.7) > Br-90 (53.3) = BR-2017 (53.3) = S-5798 (53.3) > BR-2/1 (50.0) > S-5765 (36.37) > S-5733 (33.33) > S-5742 (16.7) | |
| Final Germination over the treatment Range inclusive control (NaCl) | |
| BR-99 (96.7) = S-5733 (94.9) ≈ S-5742 (93.3) ≈ BR-2/1 (92.7) ≈ BR-90 (92.3) ≈ S-5759 (91.5) ≈ S-5747 (91.5) ≈ S-5761 (91.5) > S-5785 (88.5) ≈ S-5765 (87.9) ≈ S-5798 (86.4) > S-5797 (82.4) ≈ S-5784 (82.1) = > S-5825 (80.9) > BR-2017 (75.5) | |
| Final Germination over the treatment Range inclusive control (Na ₂ SO ₄) | |
| BR-90 (91.8) ≈ S-5747 (90.06) > S-5825 (86.7) ≈ BR-2/1 (85.6) ≈ S-5742 (85.2) S-5765 (84.3) ≈ BR-99 (83.3) ≈ S-5798 (81.2) = S-5761 (81.2) ≈ BR-2017 (80.9) > S-5784 (78.5) ≈ S-5733 (78.2) = SW-5759 (78.2) > S-5785 (76.1) ≈ S-5797 (75.5) | |

Table 3. Three way ANOVA of germination data of 15 guar varieties tested against concentrations of NaCl.

| Source | SS | df | MS | F | p |
|--------------------------|------------|------|-----------|---------|------------|
| NaCl concentrations(bar) | 267576.97 | 10 | 26757.69 | 191.68 | 0.0001 *** |
| Varieties | 1483263.59 | 14 | 494421.19 | 3541.80 | 0.0001 *** |
| Days | 332884.65 | 3 | 23777.47 | 170.33 | 0.0001 *** |
| Bar x varieties | 63008.08 | 140 | 2100.27 | 15.045 | 0.0001 *** |
| Bar x Days | 88494.25 | 30 | 632.10 | 4.528 | 0.0001 *** |
| Varieties x Days | 227949.29 | 42 | 5427.36 | 38.879 | 0.0001 *** |
| Bar x Varieties x Days | 135054.04 | 420 | 321.55 | 2.30 | 0.0001 *** |
| Error | 184266.67 | 1320 | 139.596 | - | - |
| Total | 2782497.52 | 1979 | - | - | - |

| NaCl Concentrations LSD _{0.05} : 2.4430 | | | | | Varieties or lines, LSD _{0.05} : 2.8530 | | | | |
|--------------------------------------------------------------------|---------|-------|-----|-----|--------------------------------------------------|---------|-------|-----|------|
| Rank | TRT | Mean | n | NSR | Rank | TRT | Mean | n | NSR |
| 1 | Control | 77.55 | 180 | a | 1 | BR-90 | 89.02 | 132 | a |
| 2 | 1 Bar | 76.05 | 180 | ab | 2 | BR-99 | 88.64 | 132 | a |
| 3 | 2 bar | 74.28 | 180 | bc | 3 | BR-2/1 | 80.23 | 132 | b |
| 4 | 3 Bar | 72.06 | 180 | c | 4 | S-5733 | 71.14 | 132 | c |
| 5 | 4 Bar | 69.33 | 180 | d | 5 | S-5742 | 64.47 | 132 | d |
| 6 | 5 bar | 66.22 | 180 | e | 6 | S-5747 | 64.09 | 132 | d |
| 7 | 7 Bar | 63.50 | 180 | f | 7 | S-5825 | 60.61 | 132 | e |
| 8 | 6 Bar | 63.00 | 180 | f | 8 | S-5759 | 60.08 | 132 | ef |
| 9 | 9 Bar | 50.72 | 180 | g | 9 | S-5797 | 59.47 | 132 | efg |
| 10 | 8 Bar | 49.39 | 180 | g | 10 | S-5765 | 59.24 | 132 | efgh |
| 11 | 10 Bar | 40.28 | 180 | h | 11 | BR-2017 | 57.27 | 132 | fgh |
| TRT, Treatments (salt concentrations or varieties; NSR, NS ranges) | | | | | 12 | S-5798 | 56.52 | 132 | gh |
| | | | | | 13 | S-5761 | 56.21 | 132 | h |
| | | | | | 14 | S-5785 | 50.53 | 132 | i |
| | | | | | 15 | S-5784 | 40.30 | 132 | j |

| Days of incubation LSD 0.0-5: 1.4733 | | | | |
|--------------------------------------|-------|-------|-----|-----|
| Rank | TRT | Mean | n | NSR |
| 1 | Day 4 | 89.64 | 495 | a |
| 2 | Day 3 | 52.51 | 495 | b |
| 3 | Day 2 | 63.96 | 495 | c |
| 4 | Day 1 | 19.31 | 495 | d |

Mean germination-based tolerance order (NaCl):
N = 132, Range: 89.0 – 40.3% (8 groups)
[Br-90 = Br-99] > [BR-2/1] > [S-5733] > [S-5742 = S-5747] > [S-5825 = S-5759 = S-5797 = S-5765 = BR-2017 = S-5798] > [S-5761] > [S-5785] > [S-5784]

Table 4. Three way ANOVA of germination data of 15 guar varieties tested against concentrations of Na₂SO₄.

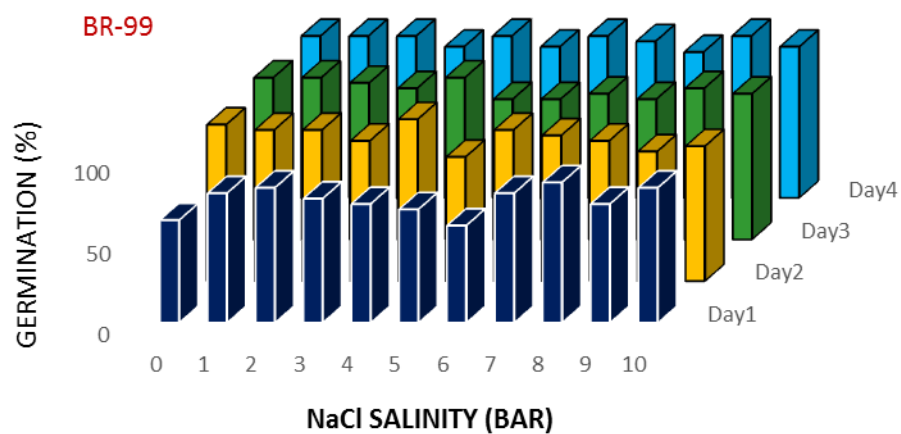
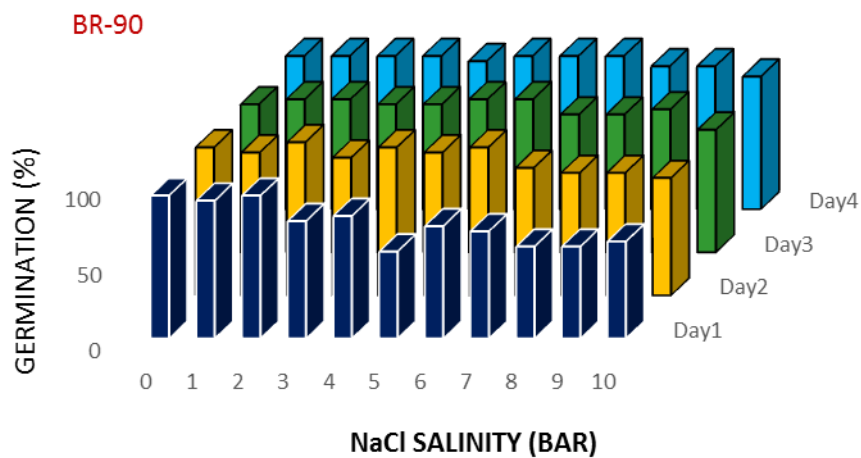
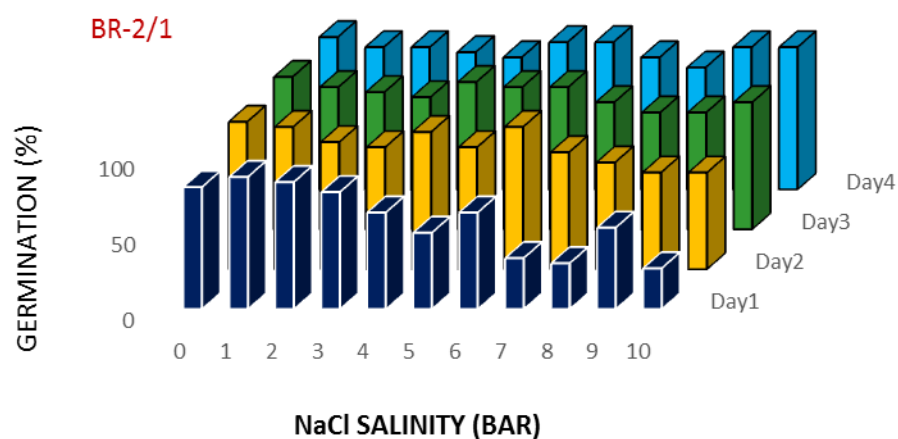
| Source | SS | df | MS | F | p |
|------------------------------------------------------|------------|------|-----------|---------|------------|
| Na ₂ SO ₄ concentrations (bar) | 604383.33 | 10 | 60438.33 | 330.30 | 0.0001 *** |
| Varieties | 145568.69 | 14 | 485156.23 | 2651.42 | 0.0001 *** |
| Days | 99131.31 | 3 | 7080.81 | 38.70 | 0.0001 *** |
| Bar x varieties | 127770.20 | 140 | 4259.01 | 23.28 | 0.0001 *** |
| Bar x Days | 143905.58 | 30 | 1027.91 | 5.62 | 0.0001 *** |
| Varieties x Days | 59919.19 | 42 | 1426.65 | 7.80 | 0.0001 *** |
| Bar x Varieties x Days | 181175.25 | 420 | 431037 | 2.36 | 0.0001 *** |
| Error | 241533.33 | 1320 | 182.78 | - | - |
| Total | 2913289.89 | 1979 | - | - | - |

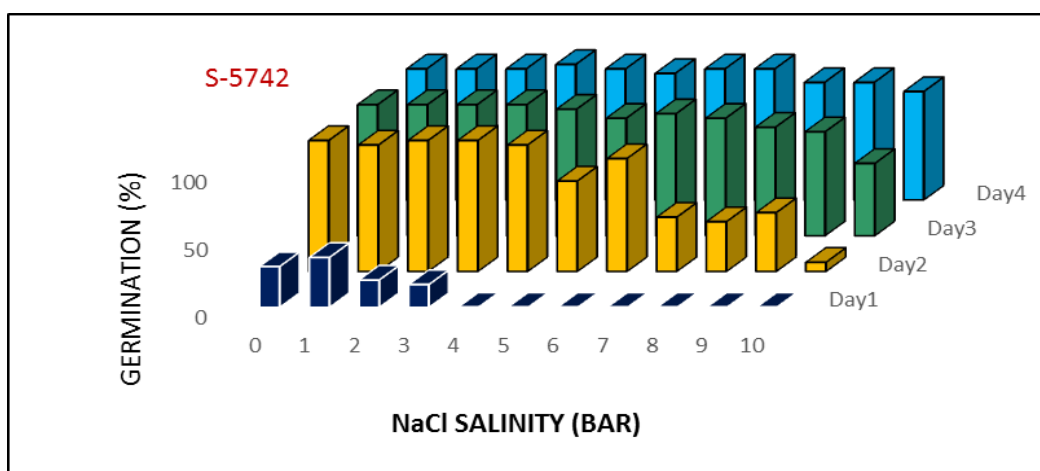
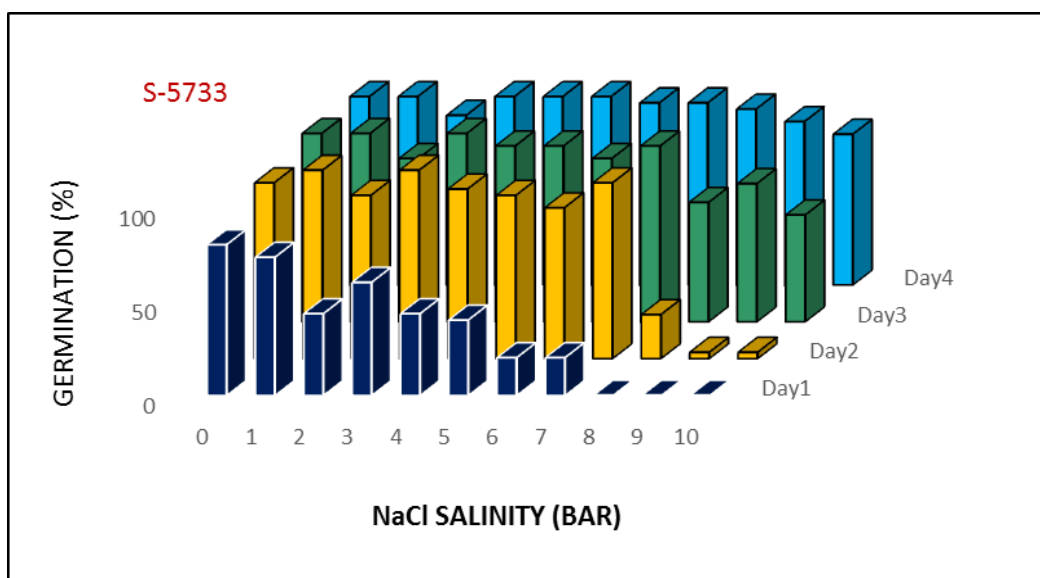
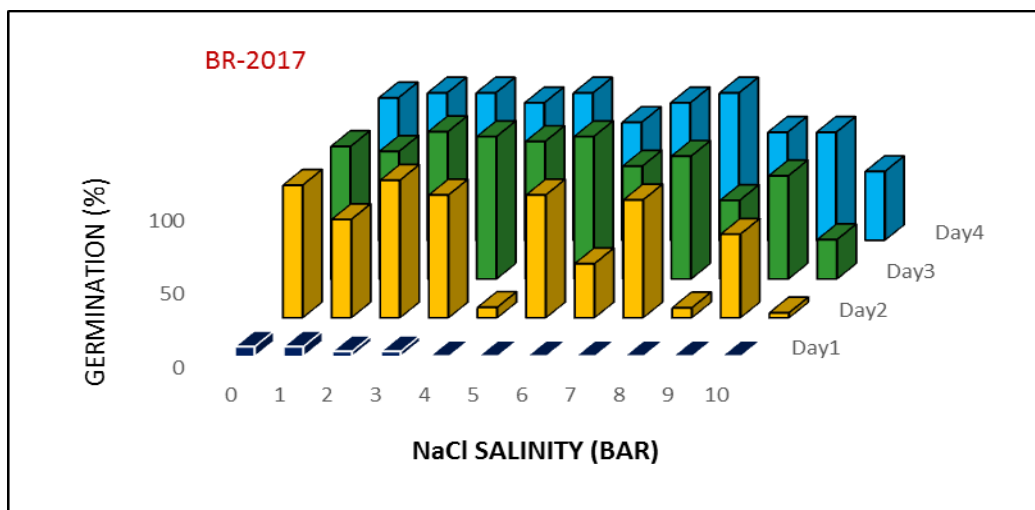
| Na ₂ SO ₄ Concentrations LSD _{0.05} : 2.7972 | | | | | Varieties or lines, LSD _{0.05} : 3.2664 | | | | |
|-----------------------------------------------------------------------------|---------|-------|-----|-----|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|---------|-------|-----|-----|
| Rank | TRT | Mean | n | NSR | Rank | TRT | Mean | n | NSR |
| 1 | Control | 77.78 | 180 | a | 1 | BR-90 | 69.39 | 132 | a |
| 2 | 1 Bar | 74.22 | 180 | b | 2 | BR-2/1 | 65.38 | 132 | b |
| 3 | 2 bar | 68.28 | 180 | c | 3 | S-5825 | 59.85 | 132 | c |
| 4 | 3 Bar | 65.89 | 180 | c | 4 | S-5798 | 57.12 | 132 | cd |
| 5 | 4 Bar | 61.11 | 180 | d | 5 | S-5742 | 56.81 | 132 | cd |
| 6 | 5 bar | 59.17 | 180 | de | 6 | BR-99 | 56.06 | 132 | d |
| 7 | 7 Bar | 57.83 | 180 | e | 7 | S-5765 | 55.98 | 132 | d |
| 8 | 6 Bar | 46.39 | 180 | f | 8 | S-5733 | 54.92 | 132 | d |
| 9 | 9 Bar | 35.61 | 180 | g | 9 | S-5747 | 54.02 | 132 | de |
| 10 | 8 Bar | 29.33 | 180 | h | 10 | S-5784 | 51.29 | 132 | ef |
| 11 | 10 Bar | 23.28 | 180 | i | 11 | S-5785 | 51.21 | 132 | ef |
| TRT, Treatments (salt concentrations or varieties; NSR, NS ranges) | | | | | 12 | S-5797 | 50.23 | 132 | f |
| | | | | | 13 | S-5759 | 48.94 | 132 | fg |
| | | | | | 14 | S-5761 | 45.68 | 132 | g |
| | | | | | 15 | BR-2017 | 39.77 | 132 | h |
| Days of incubation LSD 0.0-5: 1.687 | | | | | Mean germination-based tolerance order (Na ₂ SO ₄): N = 132, Range: 69.4 – 39.8% (six groups) [BR-90] > [BR-2/1] > [S-5825 = S-5742 = BR-99 = S-5765 = S-5733 = S-5747 = S-5798] > [S-5784 = S-5785 = S-5797 = S-5759] > [S-5761] > [BR-2017] | | | | |
| Rank | TRT | Mean | n | NSR | | | | | |
| 1 | Day 4 | 81.13 | 495 | a | | | | | |
| 2 | Day3 | 72.76 | 495 | b | | | | | |
| 3 | Day 2 | 52.91 | 495 | c | | | | | |
| 4 | Day 1 | 10.97 | 495 | d | | | | | |

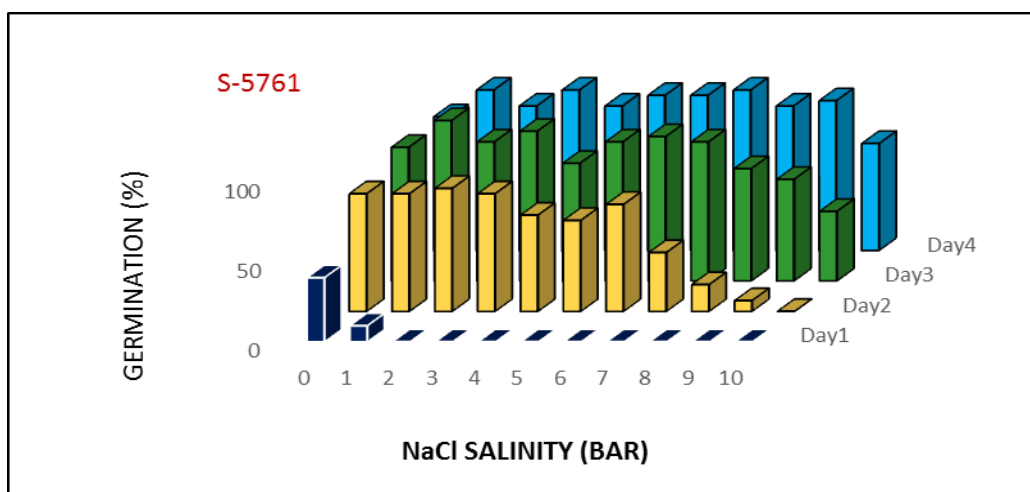
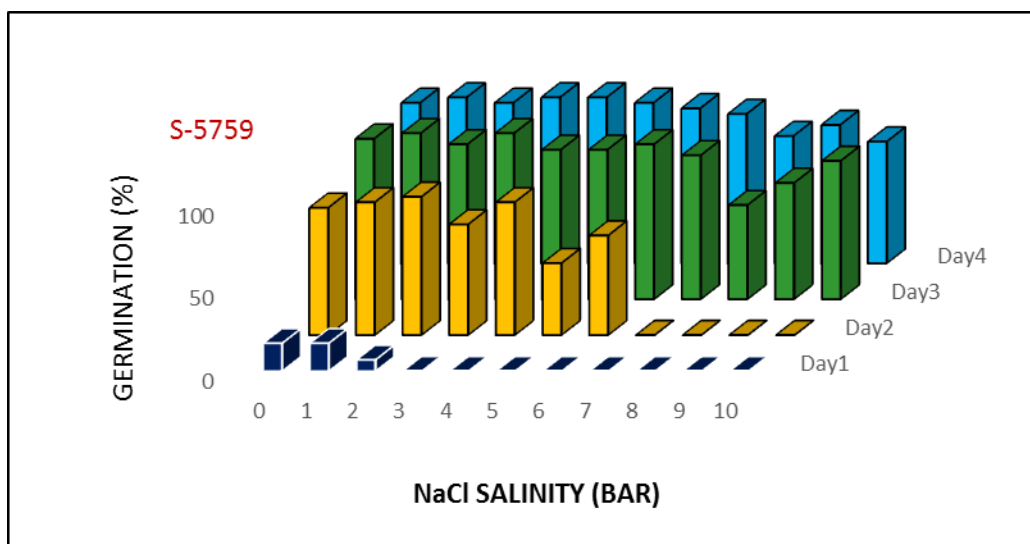
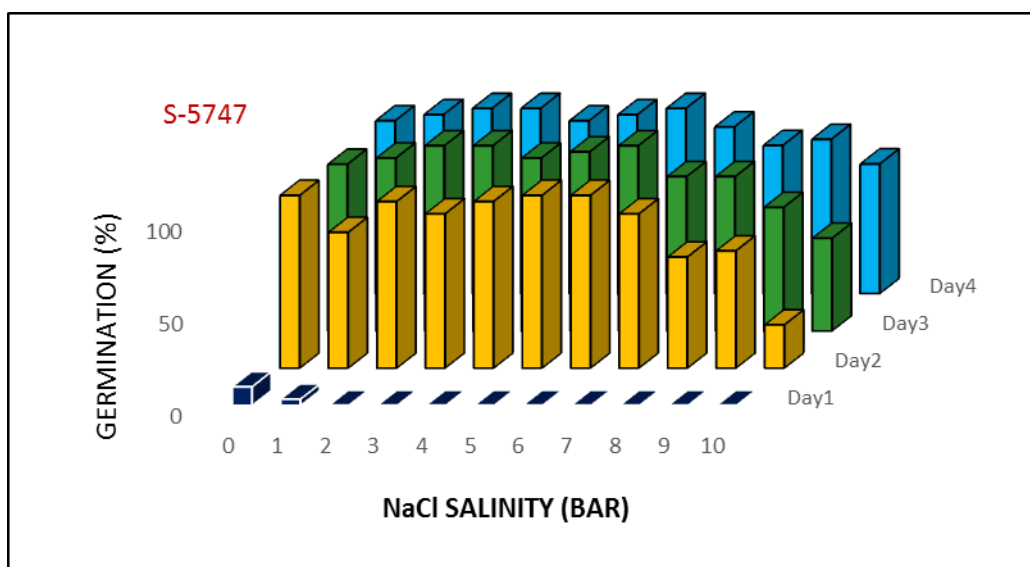
Substantial percentage of germination was observed on first day throughout the NaCl treatment (1 to 10 bar) in BR-2/1, BR-90 and BR-99. S-5733 could only germinate up till -7 bar. There was poor germination on first day in BR-2017, S-5742, S-5747, S-5759, S-5761, S-5765, S-5784, S-5785 and S-5798. Germplasms S-5825 and S-5797 could germinate up to 2 and 3 bar, respectively on first day of incubation. In all germplasms, the germination greatly improved with time (Fig. 4).

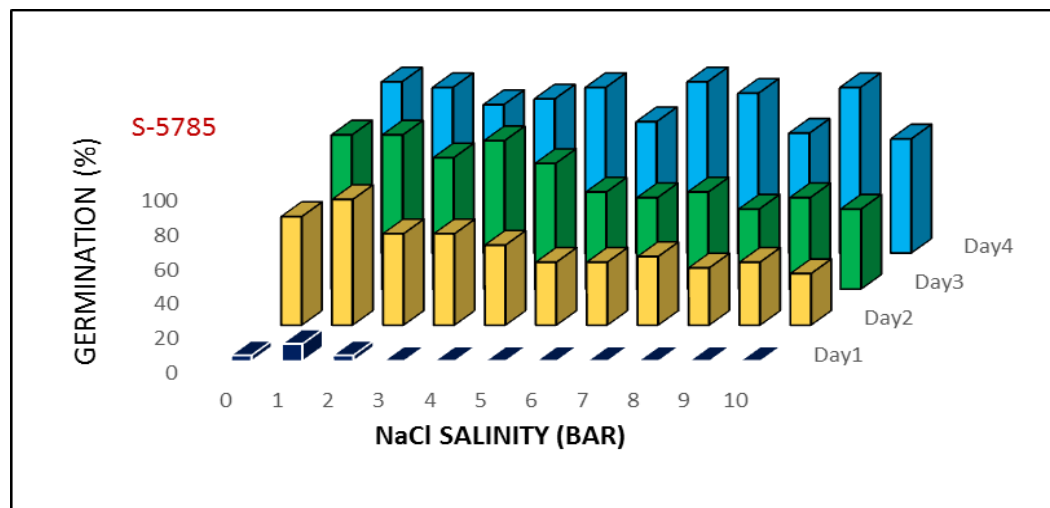
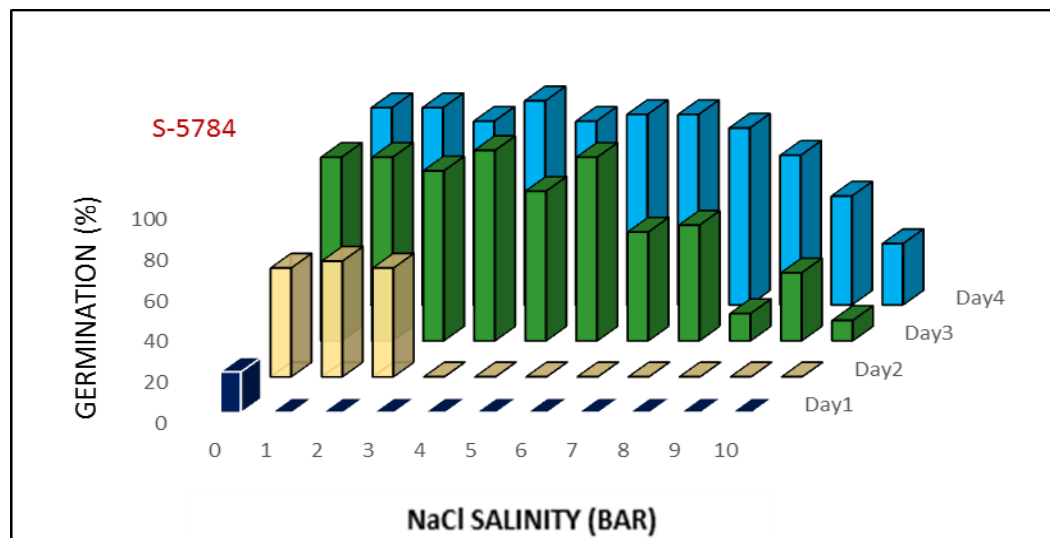
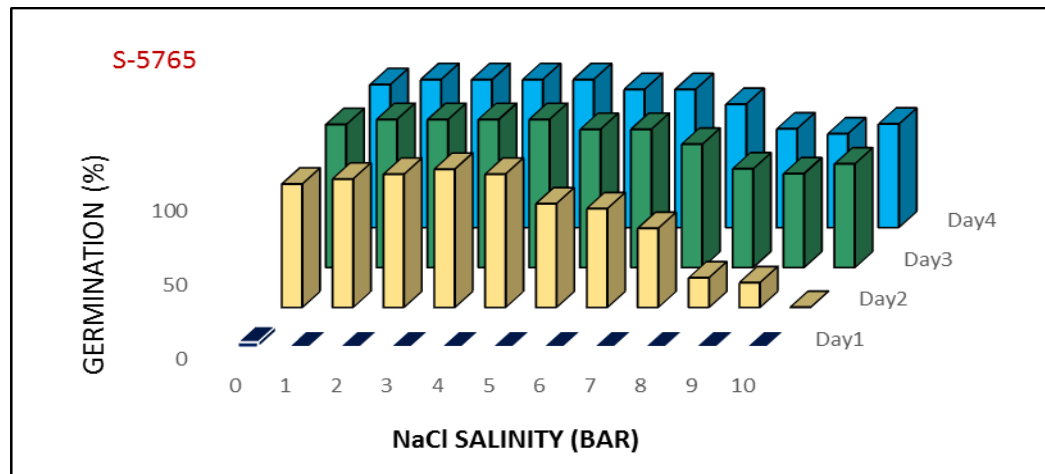
In certain germplasms such as BR- 99, BR- 2017, S-5747, S-5759, S- 5765 and S- 5798, germination on fourth day in 1 bar NaCl solution was somewhat higher than that in control. The promotion of germination (Nasr *et al.*, 2012) and stimulation of seedling growth (Patil and Karadge, 2012) under low NaCl salinity have previously been reported in literature. Slight stimulation of germination was also reported in *Rhynchosia minima* by Shaikat and Burhan (2000) in low NaCl salinity of 1 bar. Mulwani and Pollard (1939) had quite earlier found that small concentrations of NaCl stimulate germination.

Germplasms, BR-2/1, BR-90, BR-99 and S-5733 showed moderate germination on first day up to 4 bar concentration of Na₂SO₄. In higher concentration, there was meager germination. BR-2017, S-5759, S-5761, S-5765, S-5785 and S-5747 showed no or very little germination on first day except control. On first day of incubation, germplasms S-5825 and S-5797 could germinate up till 2 bar of Na₂SO₄ only. Like, NaCl, in later days of incubation, germination improved differentially in different germplasms. Na₂SO₄, however, appeared to be more deleterious to germination than NaCl (Fig. 5).









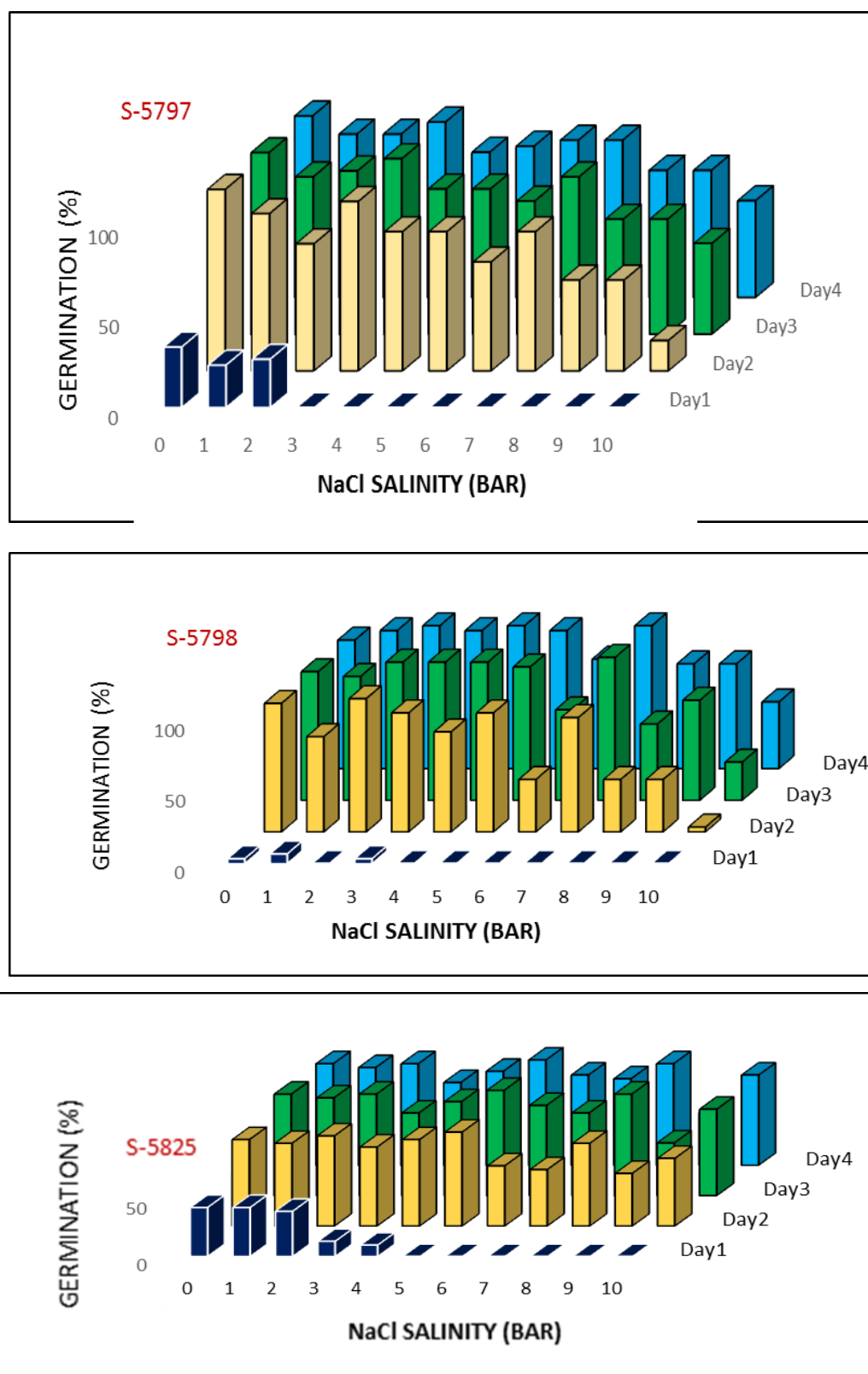
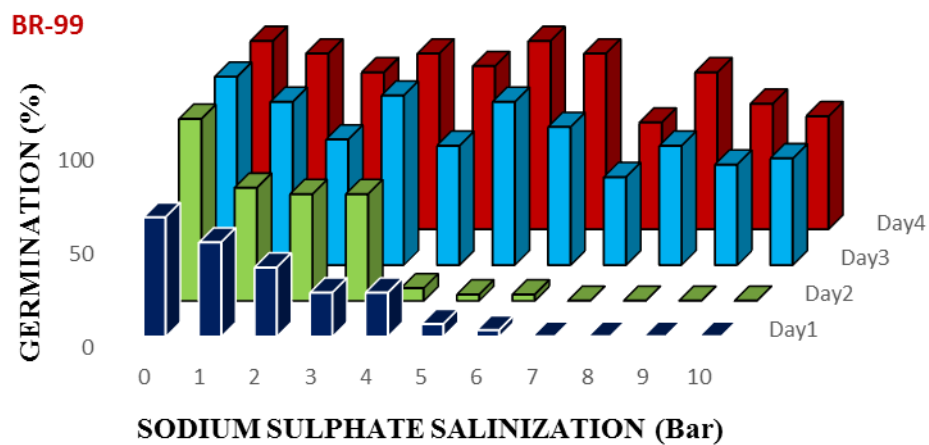
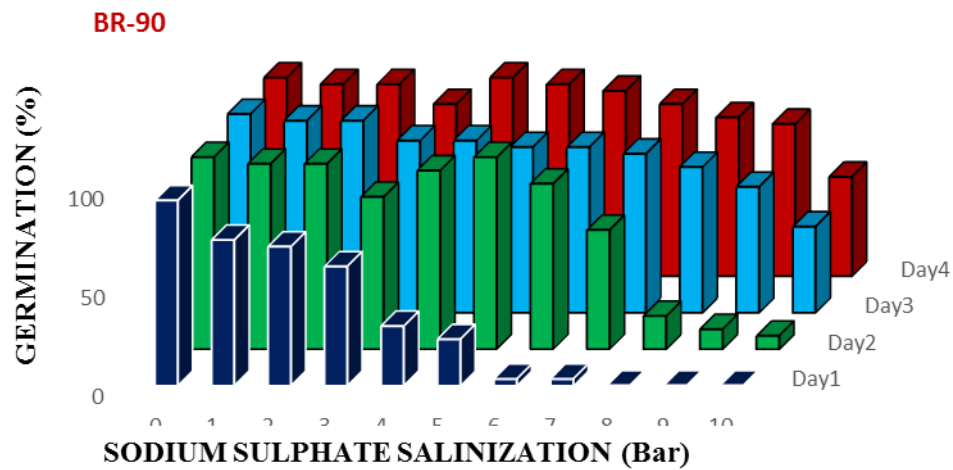
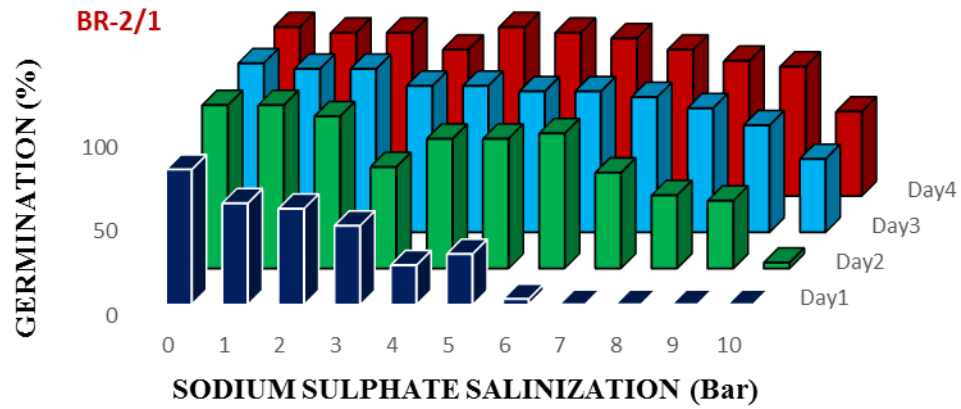
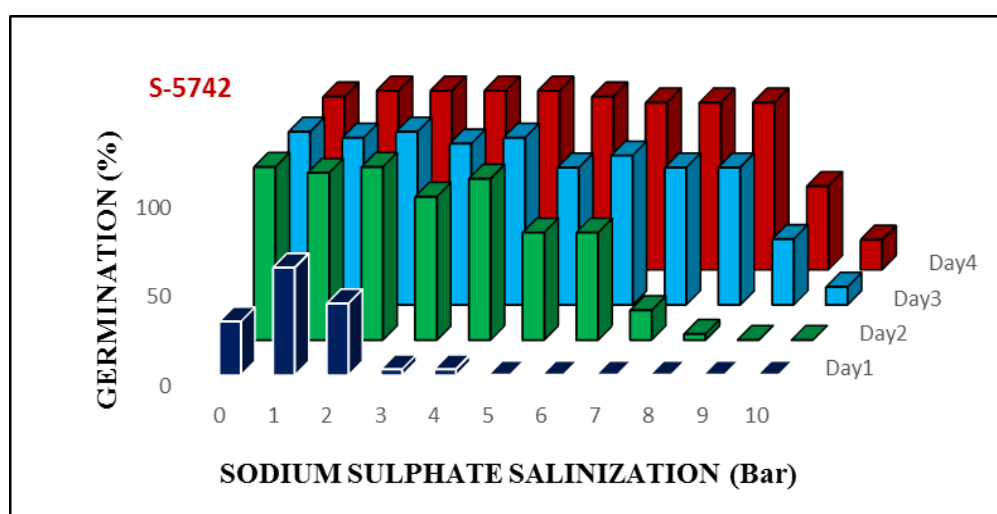
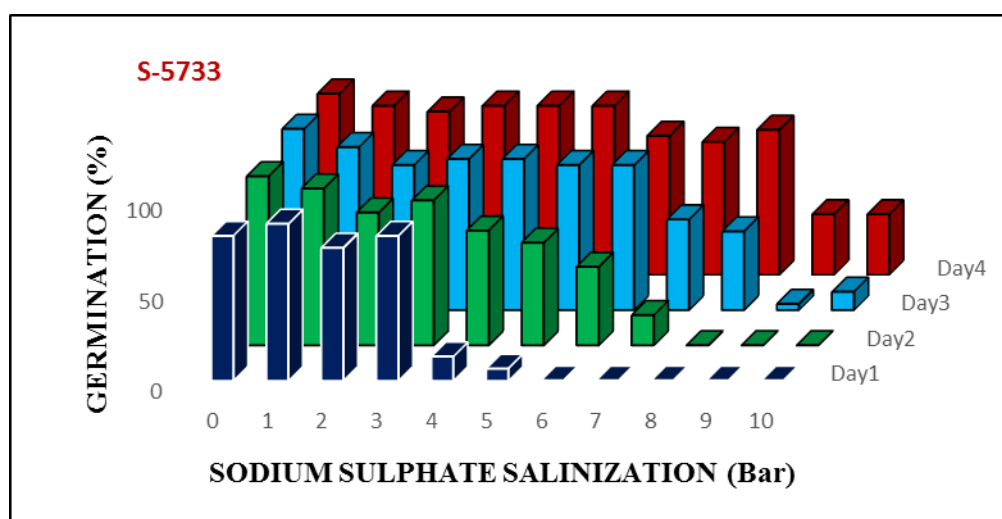
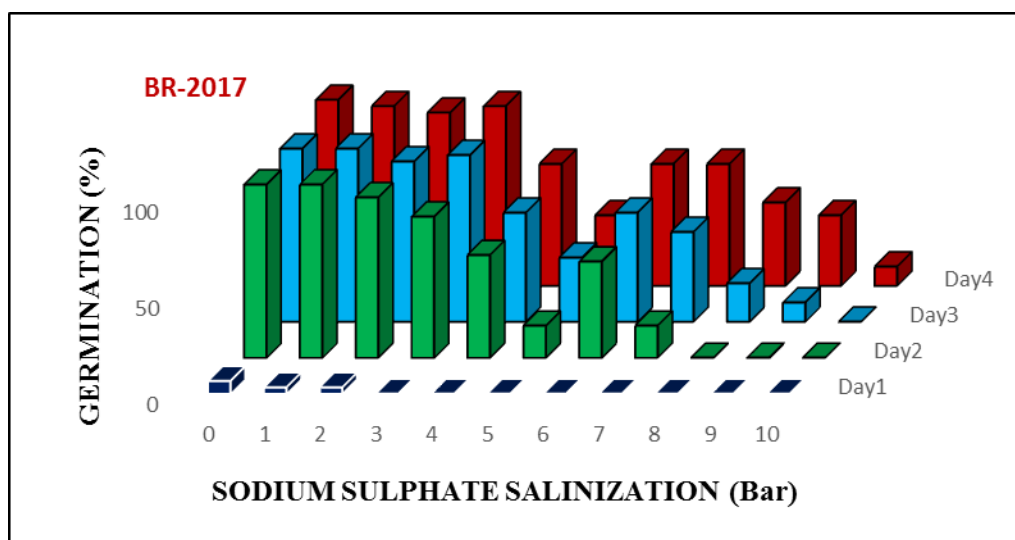
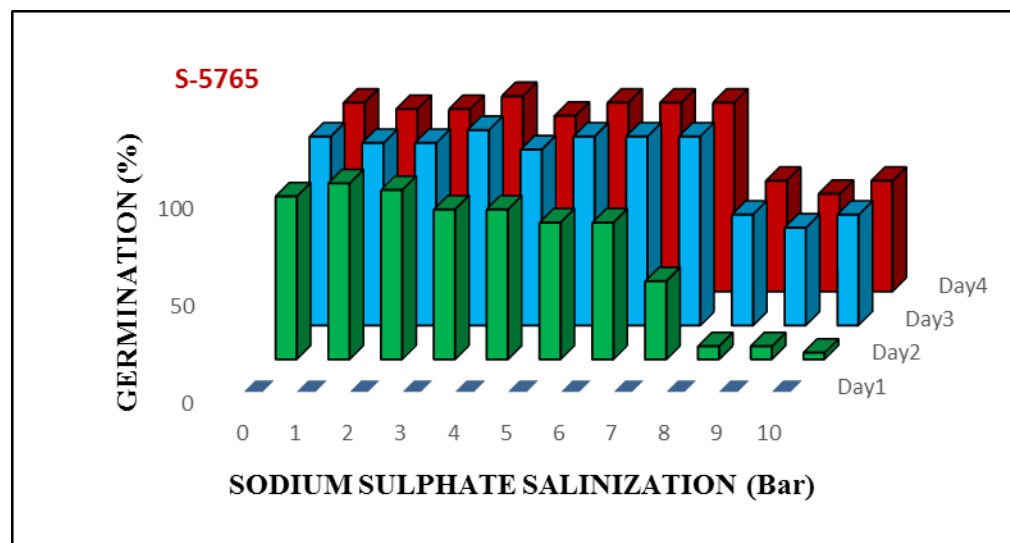
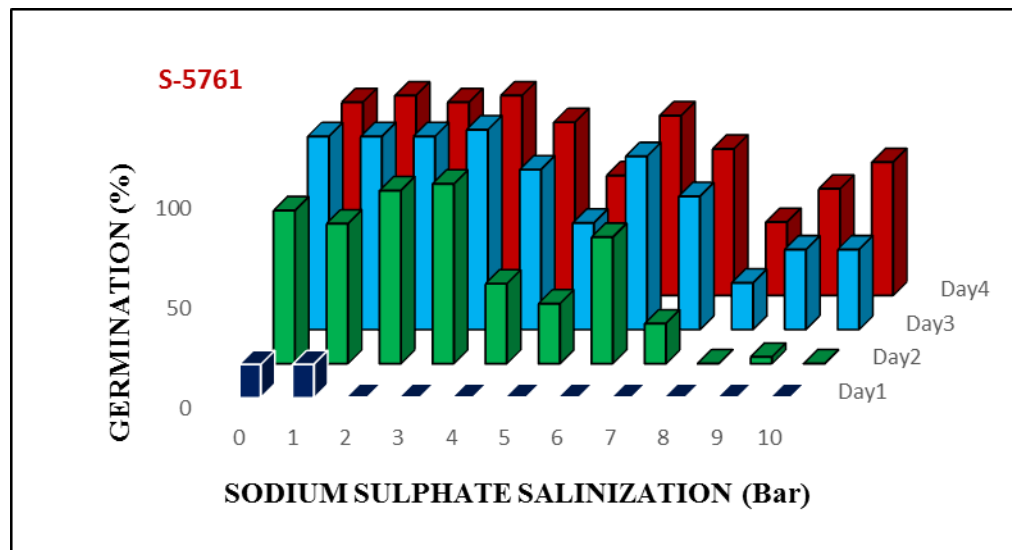
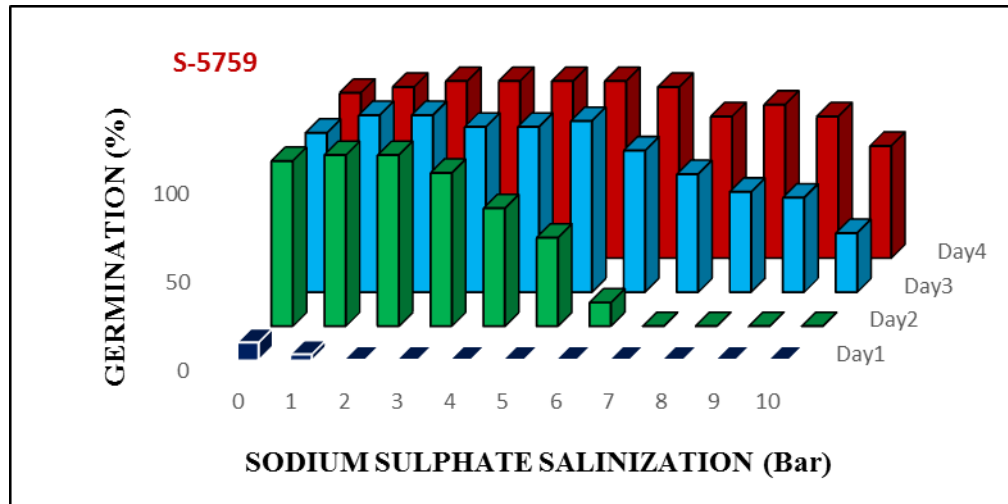
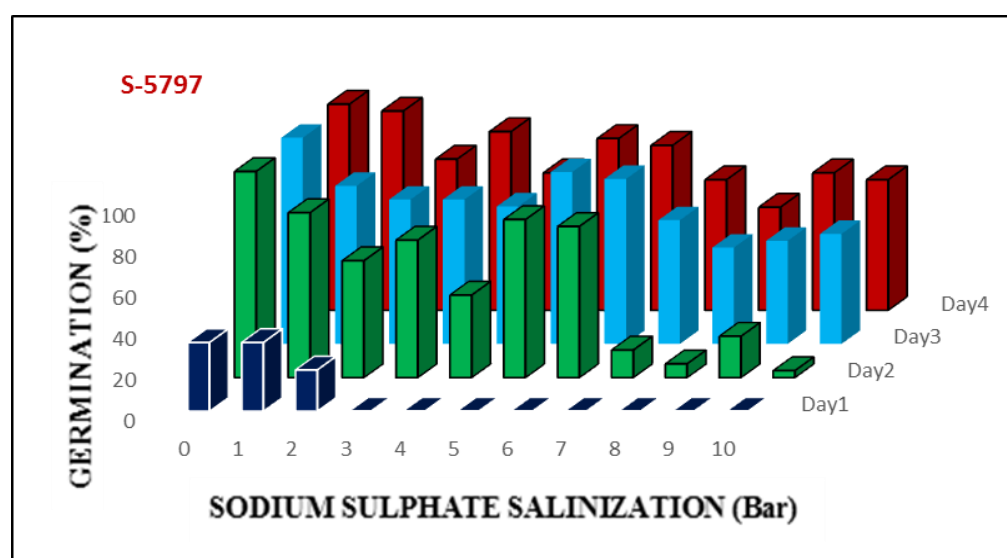
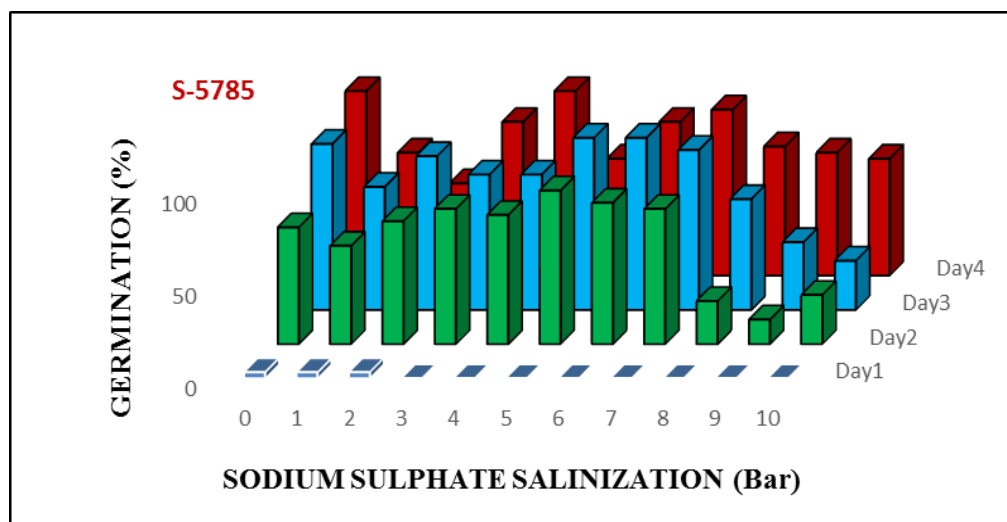
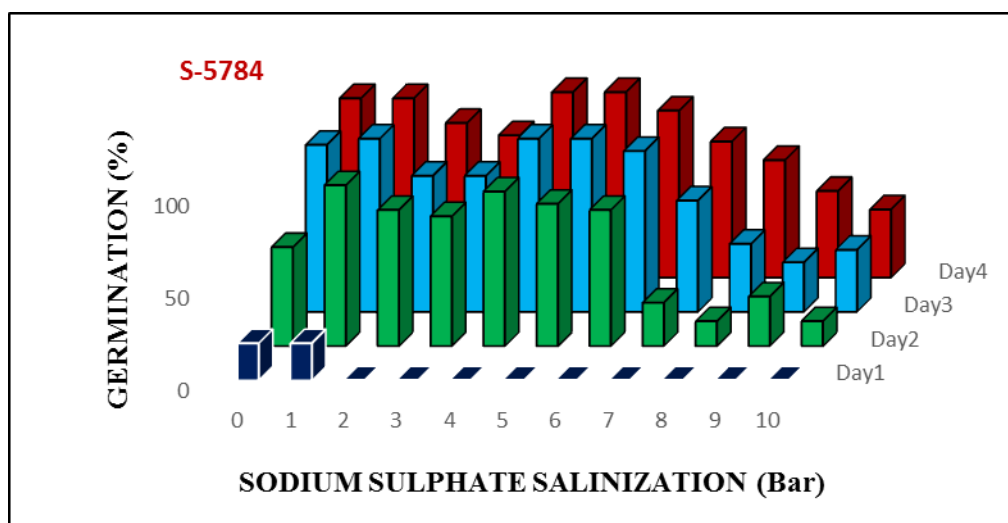


Fig. 4. Germination of 15 guar cultivars / lines under the influence of NaCl salinization and the incubation duration.









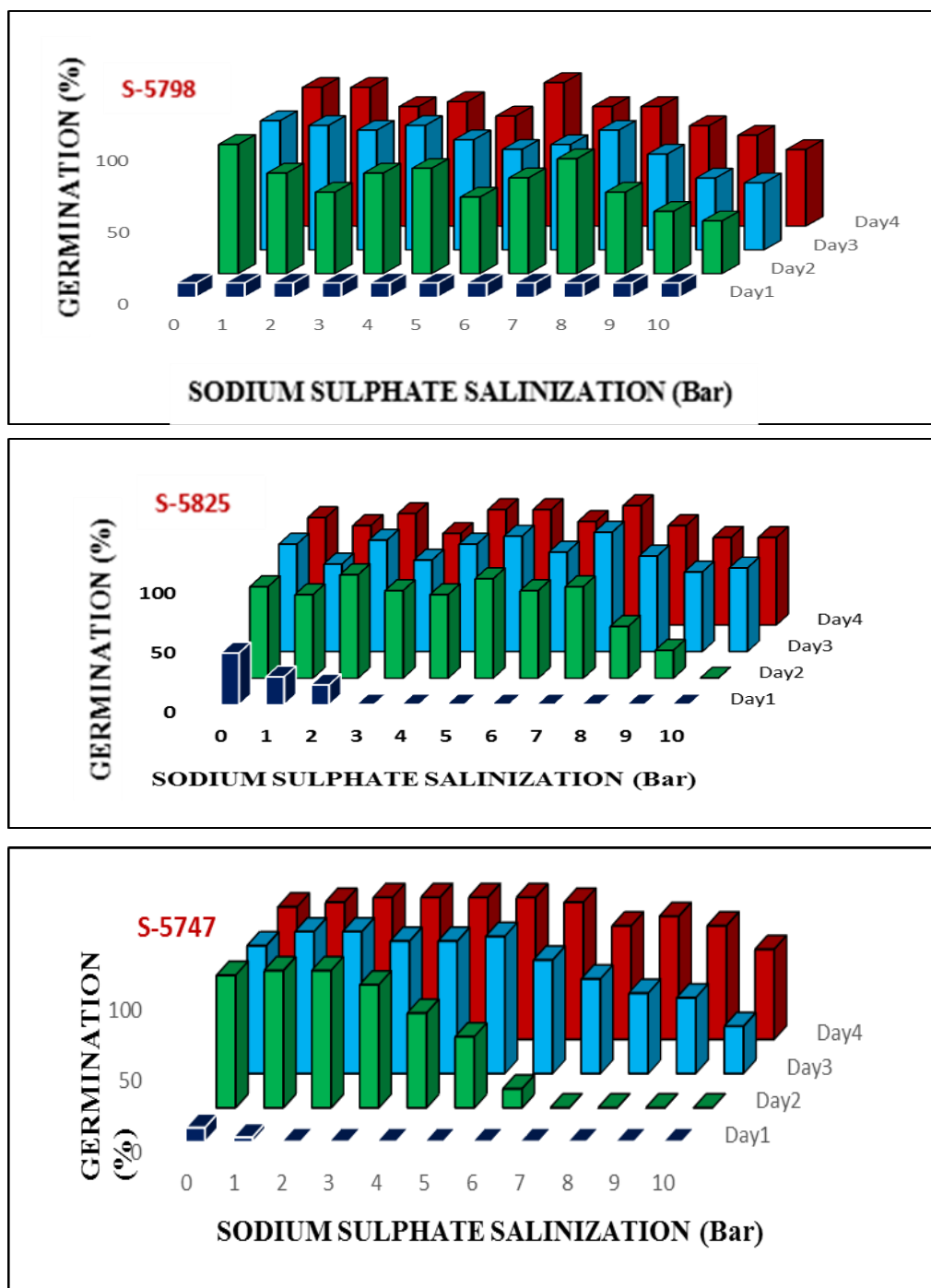


Fig. 5. Germination of 15 guar varieties / lines under sodium sulphate salinization for the given incubation period.

The 3-Way ANOVA of the component sources of variation in germination data is presented in Table 3 and 4. It is clear from the ANOVA that in both salts, the concentration of salts, nature of varieties and the incubation period appeared to influence germination highly significantly and these factors interacted with each other highly significantly. Germination declined in both salts with increasing concentration of the salts but increased with the incubation period. The varietal nature was important and inter-varietal differences of germination were significant. In this interactive system (Table 3) of three factors (germplasms, salt concentration and incubation period), NaCl concentration exhibited detrimental effects on germination more or less regularly. The maximum germination in this system was exhibited in control followed by in 1 and 2 bar. In higher concentration of salts the decline of

germination was progressively higher. Amongst the germplasms, the **maximal performance was shown by BR-90 (89.02 %) and BR-99 (88.64%)**. Germination was minimal in case of line S-5785 (50.53%). Germination was also direct function of number of days of incubation. Germination was maximum (89.64%) on the fourth day and minimum on the first day of incubation (19.3%).

More or less similar pattern of germination was exhibited under Na₂SO₄ (Table 4). With little irregularity, increase in Na₂SO₄ concentration caused decline in germination. Germination was highest in control (77.78%) followed by that in 2 bar Na₂SO₄ (74.22%). Germination was the lowest in 10 bar Na₂SO₄ (23.28%). The highest germination was exhibited by BR-90 (69.39%) followed by BR-2/1 (65.38%). The lowest germinability was shown by BR-2017 (39.77%). Germination was also the direct function of number of days of incubation. In Na₂SO₄ environment the highest germination was recorded on fourth day of incubation (81.13%) and the lowest at the first day (10.97%). The germination of individual germplasms as function of salinity (NaCl or Na₂SO₄) and number of days of incubation is given in Fig. 4 and Fig.5 showing differential degree of delay of germination in different germplasms. The delay in germination has been reported under salinity by many workers in different species (Francois *et al.*, 1986; Mondal *et al.*, 1988; Karim *et al.*, 1992; Khan *et al.*, 1984; Khan *et al.*, 1997; Chowdhury *et al.*, 2018). Emergence of guar cultivars Kinman and Esser remained unaffected by salt level up to 8.5 dS.m⁻¹ (c. 3 bar), greater levels of salt delayed germination / emergence but didn't significantly reduce the percent emerged (Francois *et al.*, 1989). Kumar and Bhardwaj (1981) have, however, reported that germination of mung bean was not affected even up to salinity of 24 dS.m⁻¹ (c. 10 bar NaCl). Abusuwar and Abbaker (2009) also reported delay in germination of guar against Seawater dilutions i.e., 31.2% germination on the first day which gradually increased to 76.3% after seven day of incubation. Decline of germination in guar under salinity is also reported by Vinisky and Ray (1988).

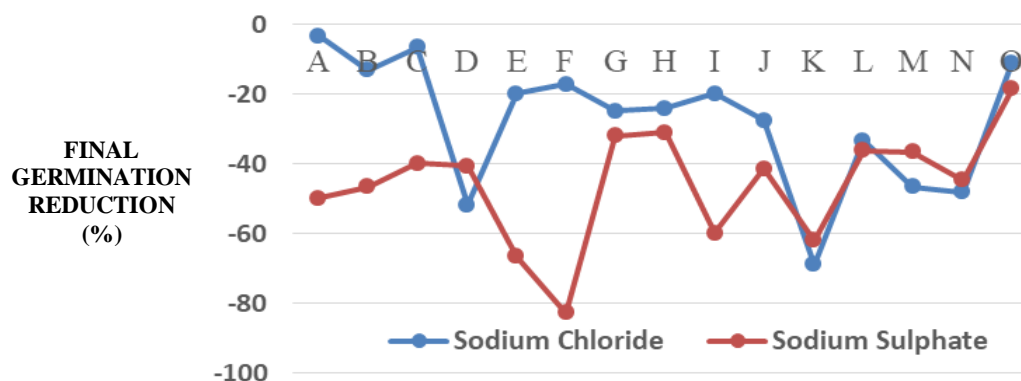


Fig. 6. *Per cent* reduction in Final germination over control under NaCl and Na₂SO₄ salinization. Varieties / Lines: A, BR-2/1, B, BR-90, C, BR-99, D, BR-2017, E, S-5733, F, S-5742, G, S-5747, H, S-5759, I, S-5761, J, S-5765, K, S-5784, L, S-5785, M, S-5797, N, S-5798 and O, S-5825.

The % germination reduction in various germplasms in 10 bar NaCl and Na₂SO₄ solutions over control is presented in Fig. 6. All germplasms exhibited decline in germination. Of course, it varied greatly amongst different germplasms. There was no or little reduction in germination of BR-2/1, BR-90, BR-99 and S-5825 in NaCl treatment and there appeared no significant difference amongst them as well. The germination was quite reduced in BR-2017, S-5797 and S-5798 (c. 50% in each) and in S-5784 (68.97%). Percent reduction in germination generally fluctuated around 20 – 30 % in S-5733, S-5742, S-5747, S-5759 and S-5761. Ramarajan *et al.* (2013) have reported decrease in germination of guar (variety 'Pusa Naubahar') at 150mM NaCl besides some adverse effects on morphological attributes as well. Al-Yemeny and Basahy (1997) reported 44% reduction in germination of guar under NaCl salinity level 16-20 mmhos /cm. Percent germination reduction was comparatively much higher in Na₂SO₄. Maximum reduction of 82.76% was exhibited by S-5742, c. 60 % by S-5761 and 62.07% by S-5784. Even the germplasms such as BR-2/1, BR-90, BR-99 and Br-2017 suffered substantially higher reduction (c. 45 – 50%) under Na₂SO₄ salinity. S-5825 suffered only by c. 20% reduction. The reduction amounted to 66.67% in S-5733. The data indicated that Na₂SO₄ was comparatively inhibitorier to guar germination than NaCl. That is to say that local in-hand germplasms are relatively more tolerant to NaCl at germination stage.

Fig. 7 and 8 presents the results of simple linear correlation and regression of FGP (Y-axis) against NaCl and Na₂SO₄ salinity using replicate data to accommodate all possible variation in germplasms performances. Against NaCl three germplasms, BR-2/1, BR-99 and S-5761 gave non-significant relationship while other related significantly negatively. The germplasms S-5785, S-5784, S-5765, S-5797, S-5798 and S-5825 better related to

NaCl concentration curvilinearly. Against Na_2SO_4 , all germplasms showed negative but differentially significant linear correlations but more closely curvilinearly. BR-99 showed significant curvilinear relationship not better than linear one. S-5733, S-5742, S-5761 and S-5765 under Na_2SO_4 showed to follow linear threshold response model of Maas and Hoffman (1977). From these scatter diagrams threshold salinities were approximated and are described later in this paper.

Germination velocity:

It is clear from the data presented above regarding germination of guar germplasms that germination varied considerably along the incubation period, obviously differentially in different germplasms. To quantify velocity of germination in different germplasms over the incubation period under varying salinity caused by NaCl and Na_2SO_4 solutions, Woodstock's (1976) Germination Velocity Index (GVI) was employed. The values of GVI (under NaCl and Na_2SO_4 environments are presented in Table 5a and 5b.

Table 5a. Values of germination velocity index of guar germplasms under NaCl salinity (Bar).

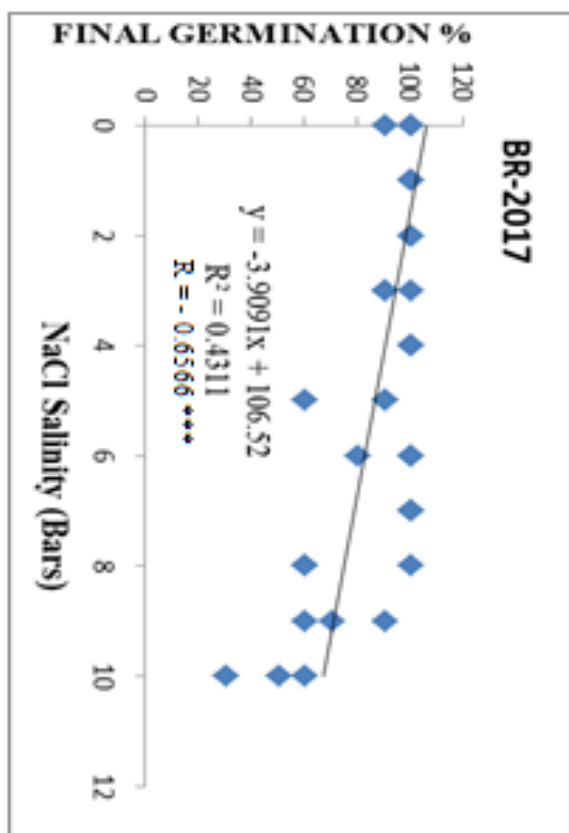
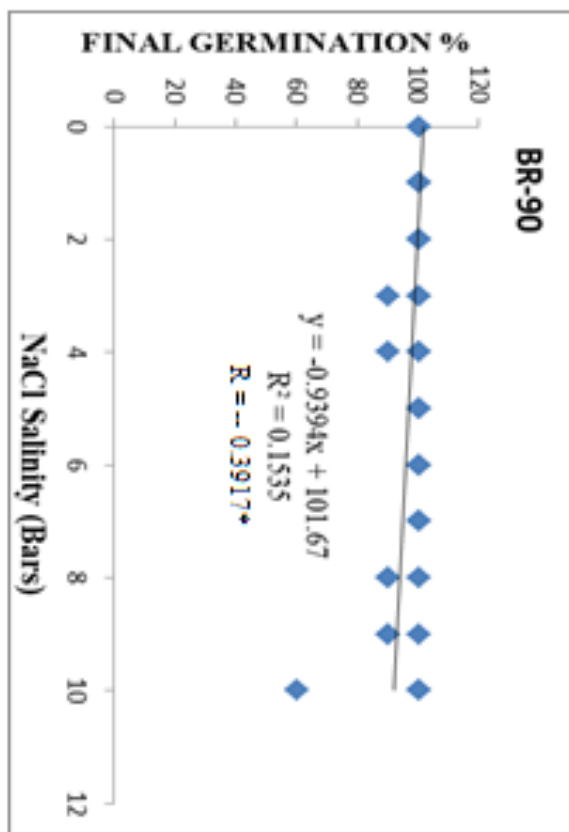
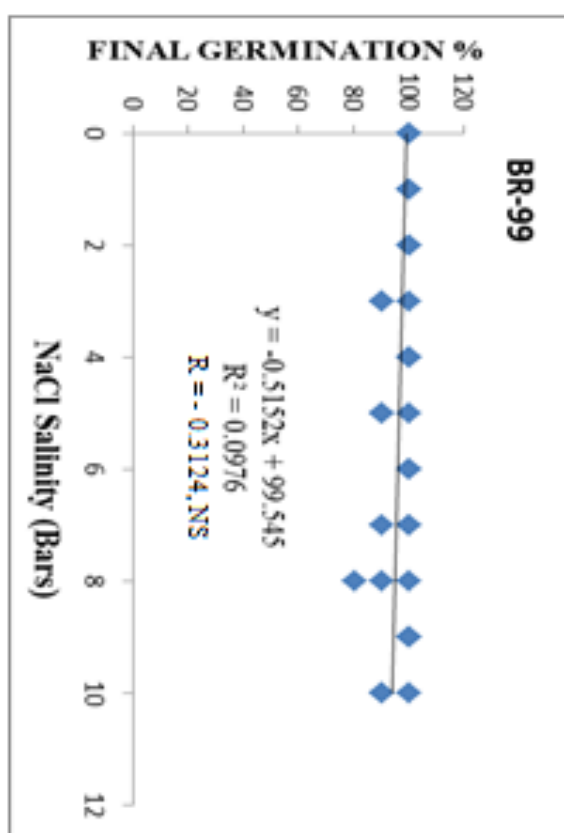
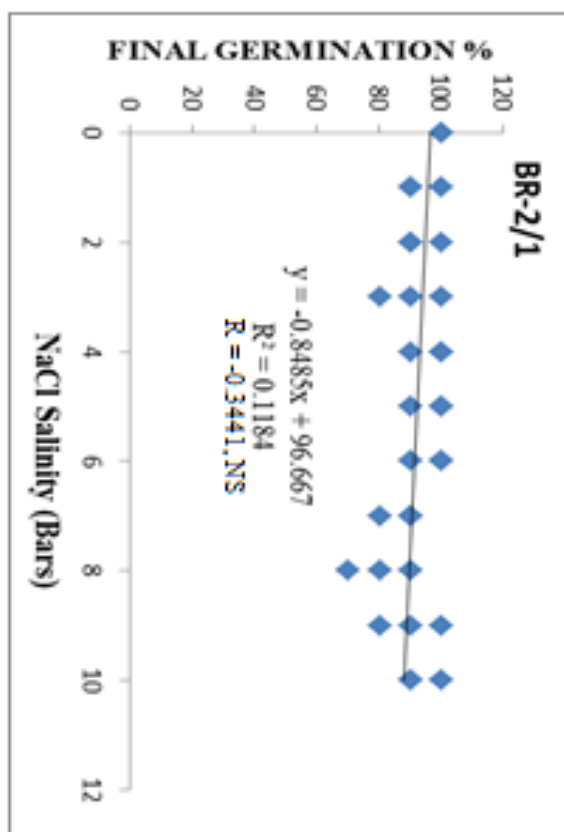
| Germplasm | NaCl Salinity (Bar) | | | | | | | | | | | Mean* | SE |
|-----------|---------------------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-------|------|
| | C | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | | |
| BR-2/1 | 26.83 | 27.83 | 25.91 | 24.25 | 23.67 | 21.08 | 23.75 | 16.75 | 15.91 | 20.08 | 16.08 | 22.09 | 1.31 |
| BR-90 | 28.75 | 28.10 | 28.5 | 23.92 | 26.5 | 23.10 | 25.83 | 24.17 | 22.25 | 22.23 | 21.83 | 25.01 | 0.80 |
| BR-99 | 24.33 | 26.6 | 26.83 | 25.10 | 26.0 | 25.0 | 23.0 | 25.5 | 26.25 | 24.33 | 25.85 | 25.35 | 0.34 |
| BR-2017 | 15.0 | 14.0 | 15.17 | 13.5 | 15.33 | 13.5 | 11.0 | 13.55 | 8.0 | 10.08 | 4.33 | 12.13 | 1.05 |
| S-5733 | 26.67 | 26.0 | 19.75 | 23.5 | 20.92 | 20.17 | 16.42 | 16.25 | 9.75 | 8.50 | 10.75 | 18.06 | 1.91 |
| S-5742 | 19.0 | 19.88 | 17.5 | 17.25 | 14.67 | 12.50 | 13.66 | 14.11 | 10.33 | 10.58 | 7.83 | 14.30 | 1.16 |
| S-5747 | 16.0 | 14.75 | 14.5 | 13.87 | 13.80 | 14.33 | 14.67 | 13.0 | 11.58 | 11.25 | 7.58 | 13.22 | 0.70 |
| S-5759 | 16.0 | 16.5 | 14.75 | 13.3 | 13.05 | 11.67 | 12.33 | 12.42 | 8.17 | 10.92 | 10.75 | 12.71 | 0.73 |
| S-5761 | 18.0 | 15.17 | 12.75 | 13.5 | 11.58 | 12.0 | 12.5 | 11.5 | 9.33 | 8.92 | 5.42 | 11.88 | 0.99 |
| S-5765 | 13.83 | 14.83 | 14.5 | 14.66 | 14.5 | 13.83 | 12.67 | 11.0 | 8.33 | 7.13 | 7.0 | 12.02 | 0.94 |
| S-5784 | 15.17 | 12.75 | 11.50 | 9.83 | 9.0 | 9.25 | 8.33 | 7.92 | 5.83 | 4.83 | 2.50 | 8.81 | 1.09 |
| S-5785 | 13.5 | 15.91 | 11.33 | 11.58 | 11.42 | 9.0 | 10.67 | 10.42 | 8.08 | 10.17 | 7.67 | 10.88 | 0.71 |
| S-5797 | 15.0 | 16.5 | 16.5 | 14.58 | 11.5 | 12.08 | 11.33 | 12.5 | 9.33 | 9.08 | 7.75 | 12.38 | 0.90 |
| S-5798 | 14.5 | 13.0 | 14.58 | 14.67 | 14.42 | 13.75 | 9.17 | 14.0 | 7.17 | 9.08 | 4.33 | 11.70 | 1.09 |
| S-5825 | 19 | 18.5 | 19.0 | 12.83 | 13.33 | 13.50 | 10.67 | 10.08 | 13.16 | 7.0 | 10.58 | 13.42 | 1.19 |

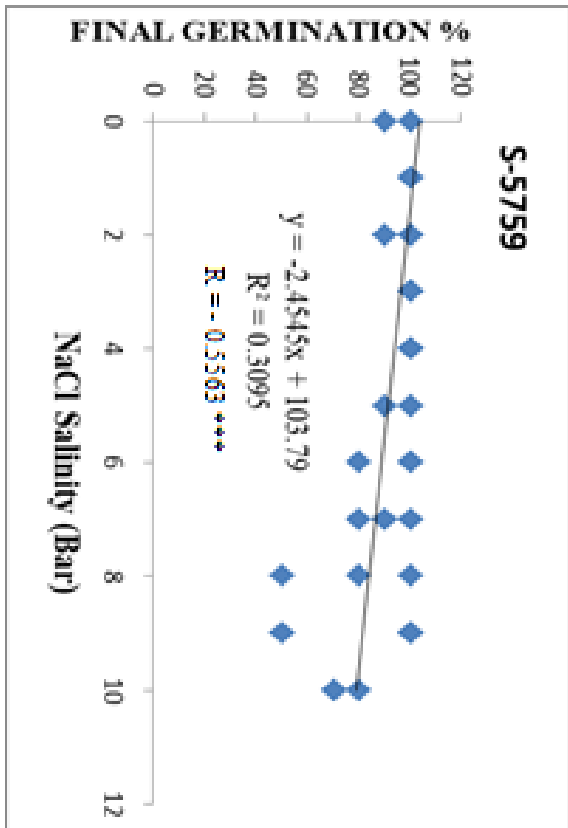
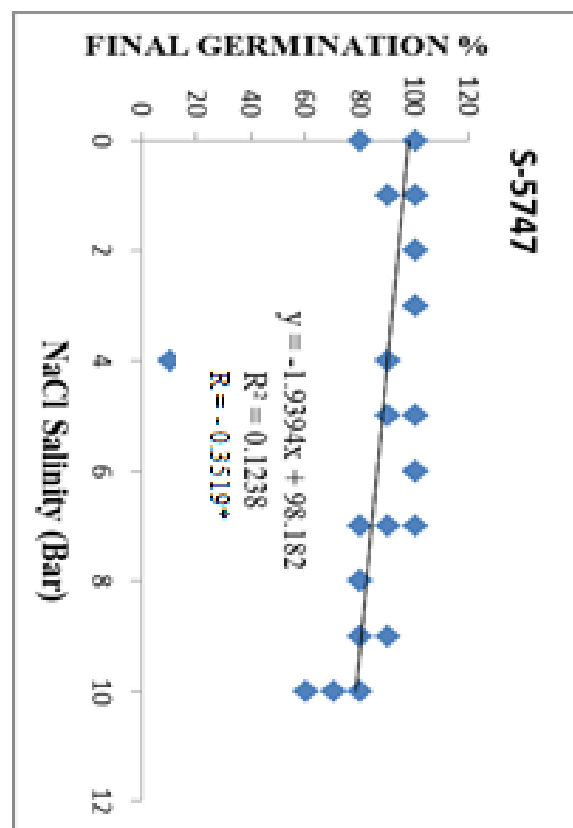
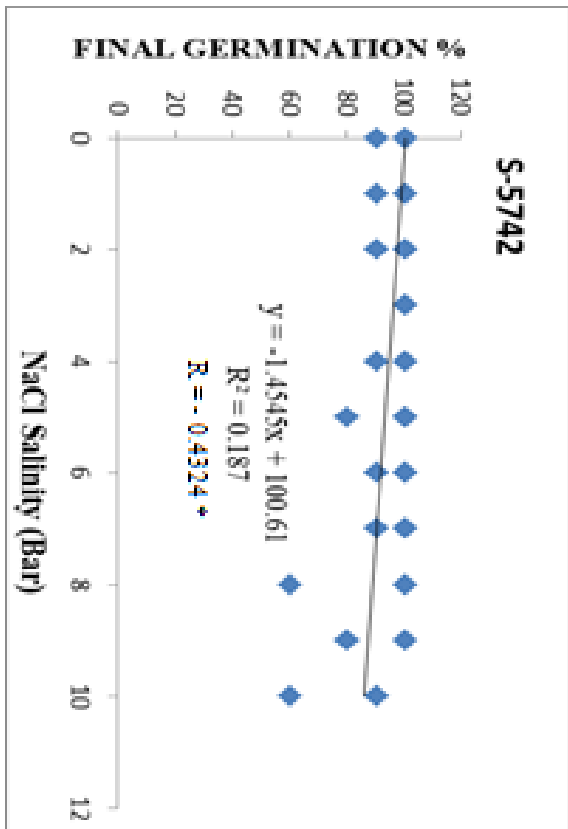
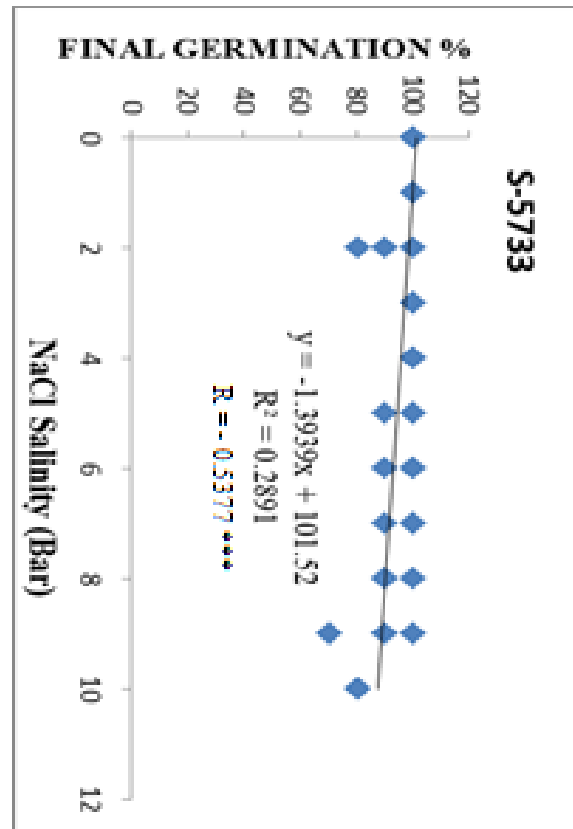
Table 5b. Values of germination velocity index of guar germplasm under Na_2SO_4 salinity (Bar).

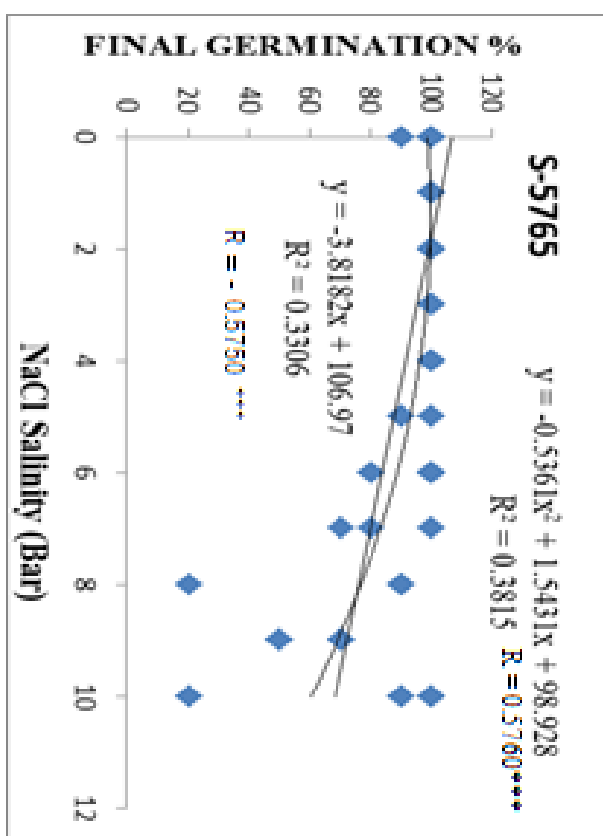
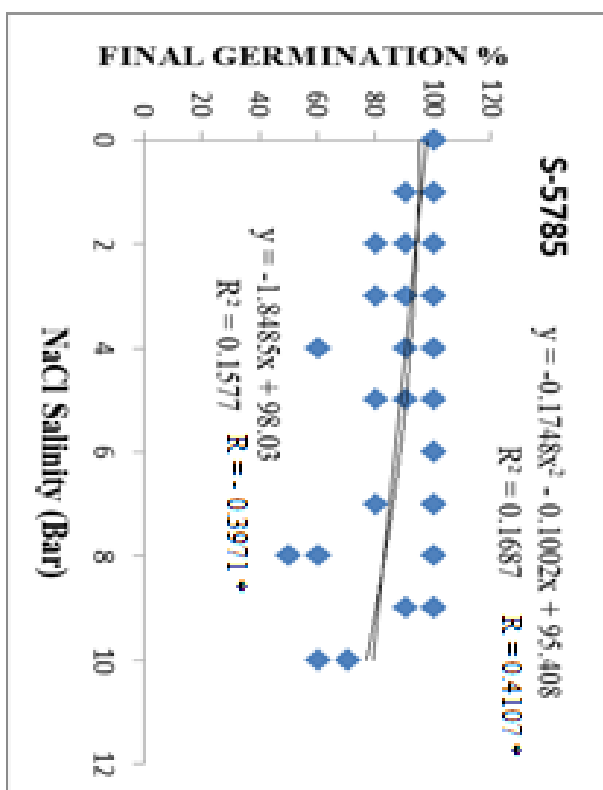
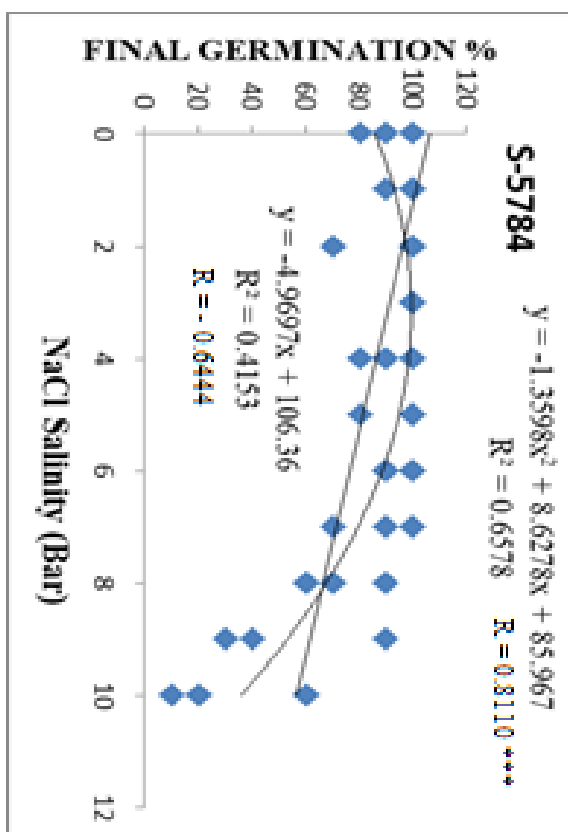
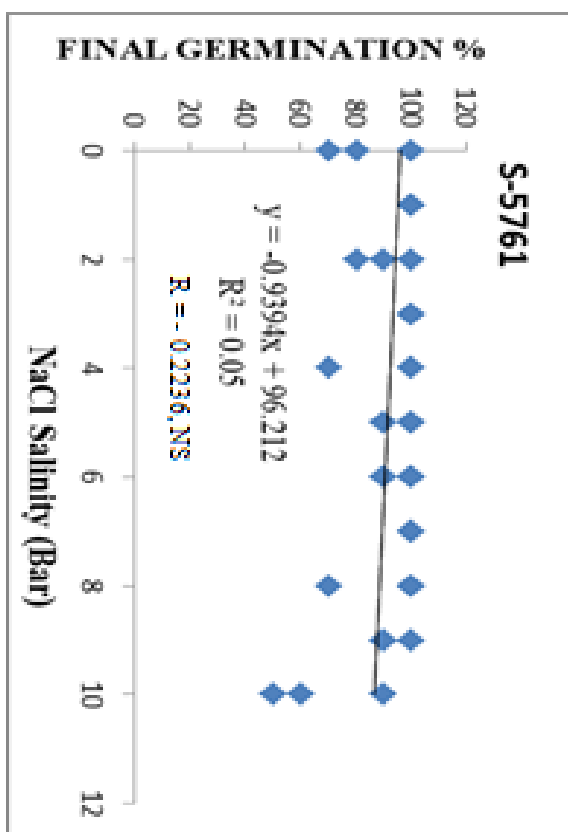
| Germplasm | Na_2SO_4 Salinity (Bar) | | | | | | | | | | | Mean* | SE |
|-----------|-----------------------------------------|-------|-------|-------|-------|-------|--------|-------|-------|-------|-------|-------|------|
| | C | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | | |
| BR-2/1 | 26.83 | 23.5 | 22.66 | 18.67 | 16.75 | 16.92 | 132.33 | 11.33 | 10.0 | 8.83 | 5.254 | 15.83 | 2.04 |
| BR-90 | 28.75 | 25.58 | 25.08 | 22.17 | 18.92 | 18.25 | 14.25 | 12.05 | 9.75 | 9.5 | 5.55 | 17.26 | 2.30 |
| BR-99 | 24.33 | 19.67 | 16.25 | 15.58 | 13.92 | 12.67 | 12.0 | 7.08 | 10.17 | 7.17 | 6.42 | 13.21 | 1.68 |
| BR-2017 | 15.0 | 14.25 | 13.25 | 13.17 | 8.88 | 4.42 | 8.67 | 7.16 | 4.75 | 3.0 | 0.75 | 8.48 | 1.49 |
| S-5733 | 26.67 | 26.58 | 23.41 | 25.08 | 16.16 | 14.16 | 11.17 | 7.92 | 7.08 | 2.58 | 2.75 | 14.87 | 2.83 |
| S-5742 | 19.0 | 23.50 | 20.75 | 14.25 | 15.25 | 12.16 | 11.83 | 9.75 | 11.33 | 7.17 | 1.50 | 13.32 | 1.89 |
| S-5747 | 16.0 | 14.8 | 14.6 | 14.10 | 13.17 | 12.42 | 10.33 | 7.67 | 7.92 | 7.33 | 5.58 | 11.26 | 1.09 |
| S-5759 | 16.0 | 15.92 | 14.0 | 14.5 | 10.5 | 7.08 | 12.08 | 8.17 | 3.33 | 4.67 | 6.0 | 10.20 | 1.39 |
| S-5761 | 18.0 | 20.5 | 11.42 | 12.42 | 11.25 | 11.75 | 9.92 | 6.83 | 5.38 | 4.08 | 2.50 | 10.39 | 1.68 |
| S-5765 | 13.83 | 13.83 | 13.66 | 13.83 | 13.5 | 14.17 | 14.17 | 11.67 | 6.0 | 5.033 | 5.83 | 11.44 | 1.13 |
| S-5784 | 15.17 | 16.92 | 11.0 | 12.08 | 14.0 | 13.67 | 12.58 | 8.17 | 6.33 | 5.50 | 4.25 | 10.88 | 1.27 |
| S-5785 | 13.5 | 10.0 | 12.67 | 12.25 | 13.0 | 13.5 | 13.08 | 12.55 | 9.41 | 6.58 | 6.75 | 11.21 | 0.79 |
| S-5797 | 15.0 | 18.25 | 12.83 | 11.58 | 8.67 | 12.17 | 11.67 | 6.92 | 5.25 | 7.25 | 6.25 | 10.53 | 1.22 |
| S-5798 | 14.5 | 12.92 | 11.17 | 13.17 | 11.0 | 11.92 | 11.67 | 12.3 | 9.75 | 8.17 | 7.0 | 11.23 | 0.66 |
| S-5825 | 19.0 | 14.67 | 17.17 | 11.30 | 11.20 | 13.83 | 11.25 | 13.83 | 10.42 | 8.33 | 7.25 | 12.57 | 1.07 |

C = control (0 bar); *, Mean over the range of treatments (0 to -10 bar) and the control.

All germplasms exhibited decline in GVI under increasing concentration (1 to 10 bar) of NaCl and Na_2SO_4 . In NaCl salinity (Table 5a), germplasms such as BR-90, BR-99 and BR-2/1 had the highest magnitude of mean GVI (25.35, 25.01 and 22.09, respectively) which was followed by S-5733 (18.06). The lowest mean GVI was shown by S-5784 (GVI: 8.81). Other germplasms had mean GVI varying amongst them around 10.88 to 13.22. The magnitude of BR-99 in all NaCl salinity treatments remained at par or above the GVI magnitude in control. This genotype exhibited c. 6.25% promotion of GVI even in 10 bar NaCl treatment. GVI of BR-90 was observed to be higher than that of BR-99 in lower salinities. In higher NaCl salinity BR-99 had little higher GVI resulting in almost equal GVI of the two varieties. BR-90 and BR-99 are the most rapidly germinating varieties amongst the genotypes tested (Table 5a). Under the influence of Na_2SO_4 salinity, magnitude of GVI declined in comparatively greater proportion than that under NaCl. BR-90 exhibited rapid germination (mean GVI = 17.26 ± 2.30) followed by BR-2/1 (15.83 ± 2.04) and S-5742 (13.32 ± 0.76). Mean GVI of BR-99 was almost equal to that of S-5742 in magnitude (13.21 ± 1.68). The lowest magnitude of GVI was exhibited by BR-2017 (8.48 ± 1.49) (Table 5b). The remaining germplasms had mean GVI varying from 10.20 to 12.57. Osmotic potential which retards the uptake of water necessary for mobilization of nutrients disturb the metabolism due to stress resulting in delay and inhibition of germination (Cicek and Cakirlar, 2002; Hassan *et al.*, 2018). Our germplasms appeared to be more sensitive to sulphate salinity.







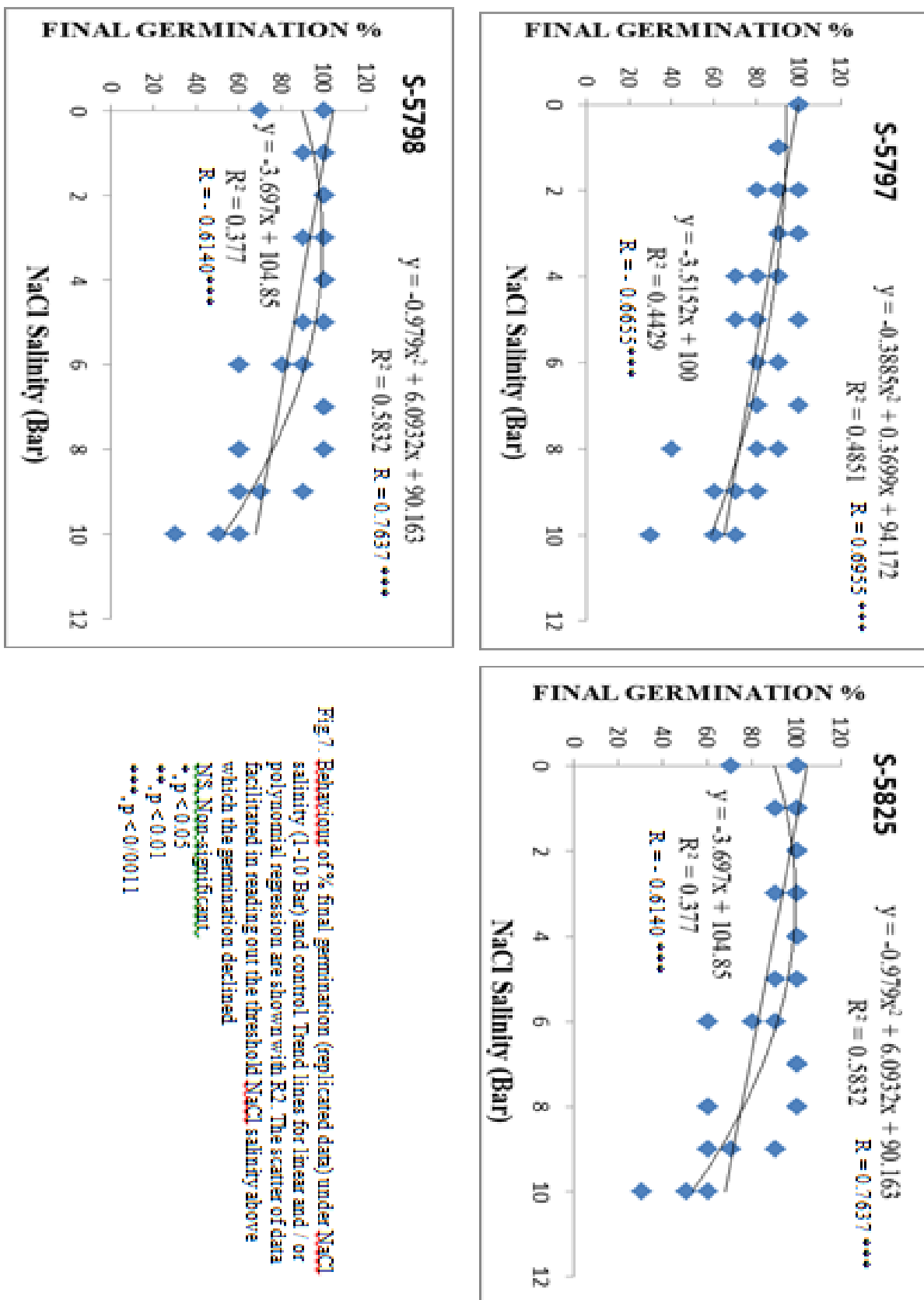


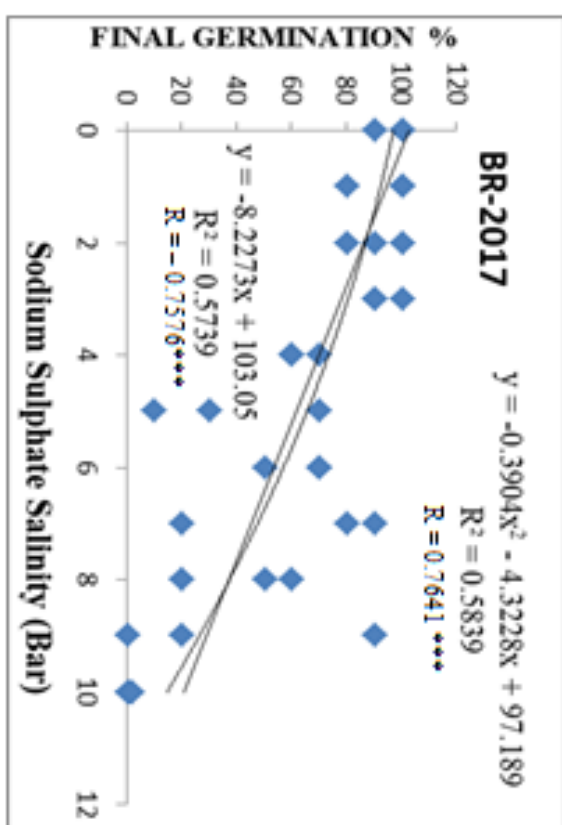
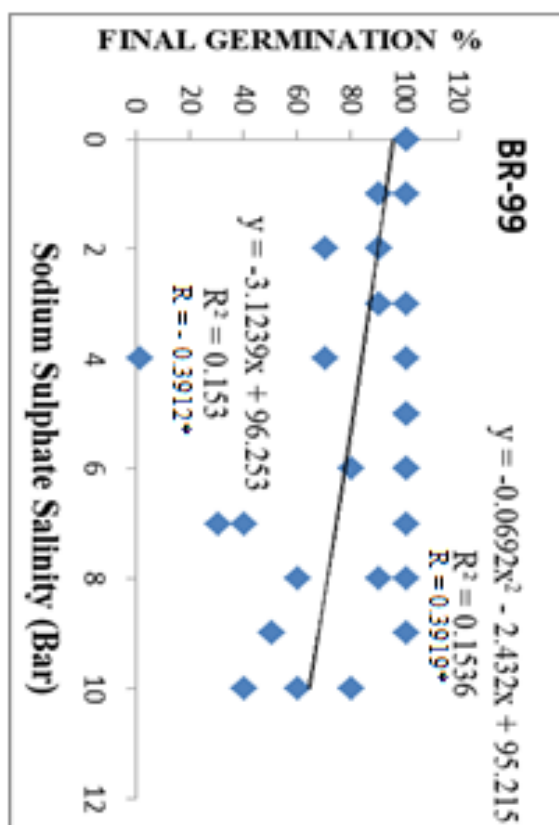
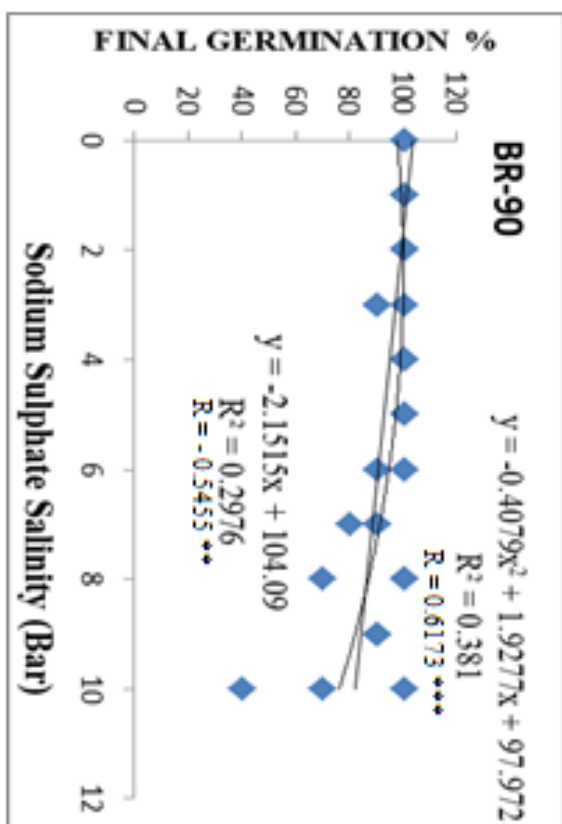
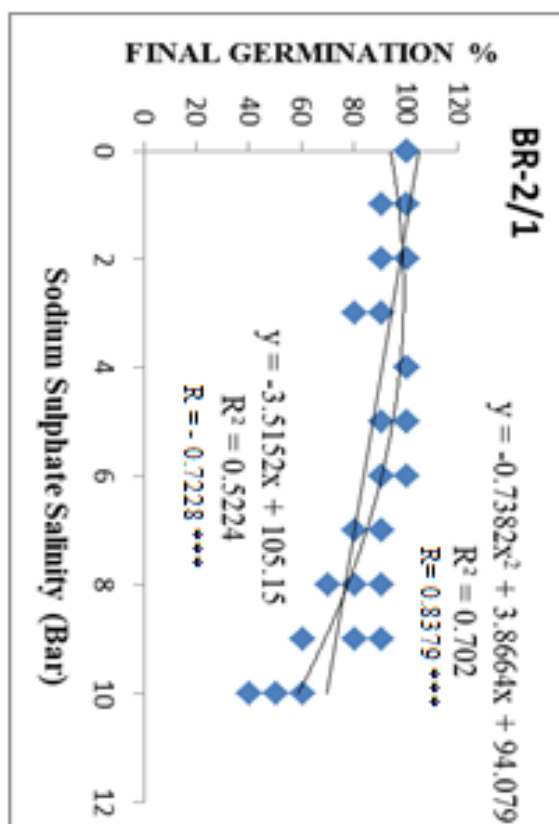
Fig.7. Behaviour of % final germination (replicated data) under NaCl salinity (1-10 Bar) and control. Trend lines for linear and / or polynomial regression are shown with R². The scatter of data facilitated in reading out the threshold NaCl salinity above which the germination declined.

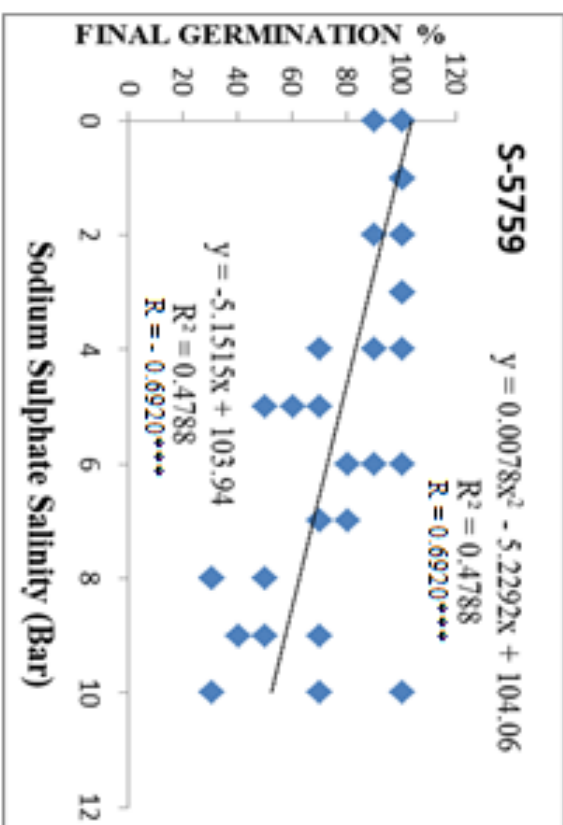
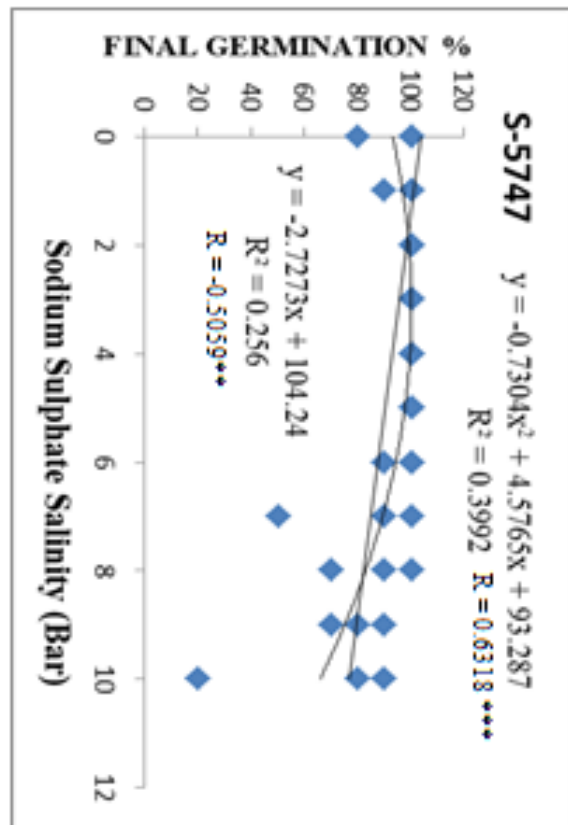
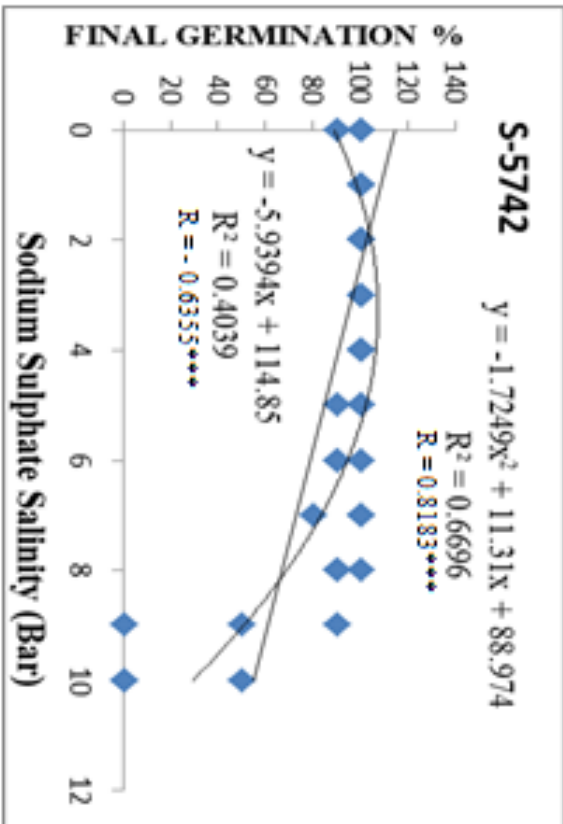
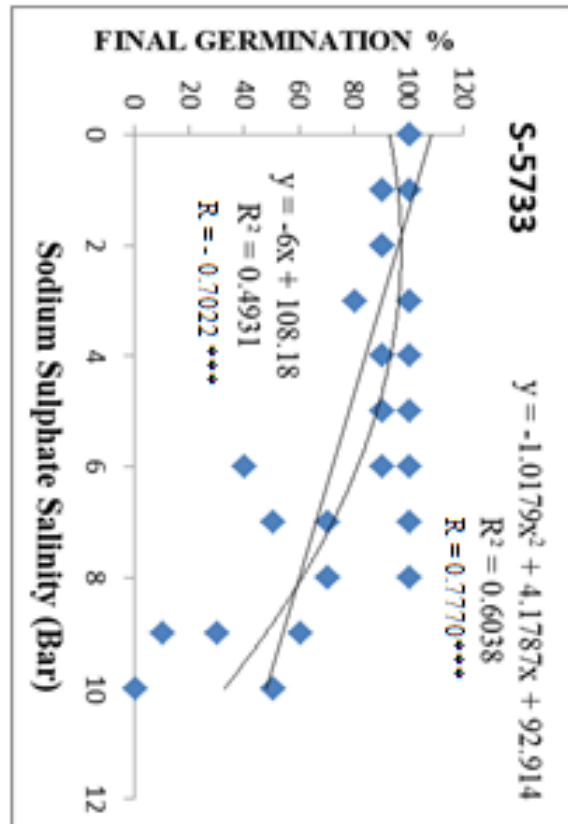
NS, Non-significant.

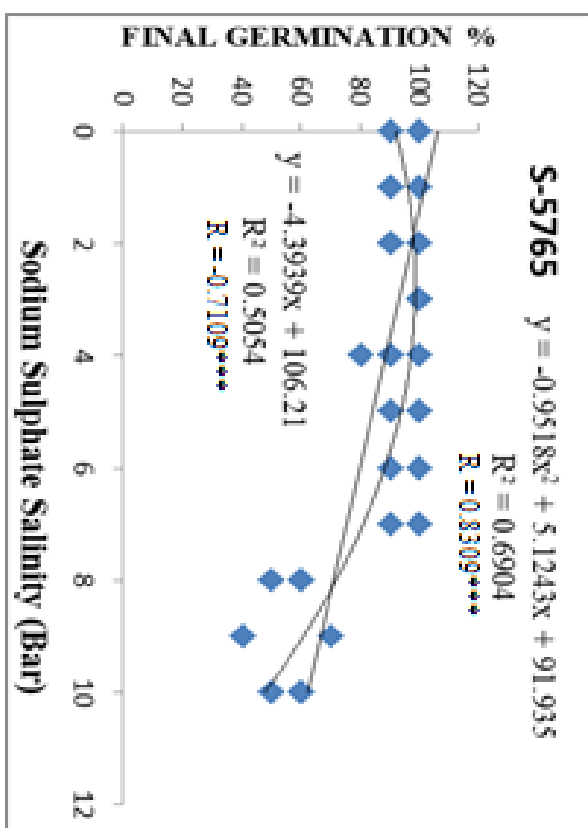
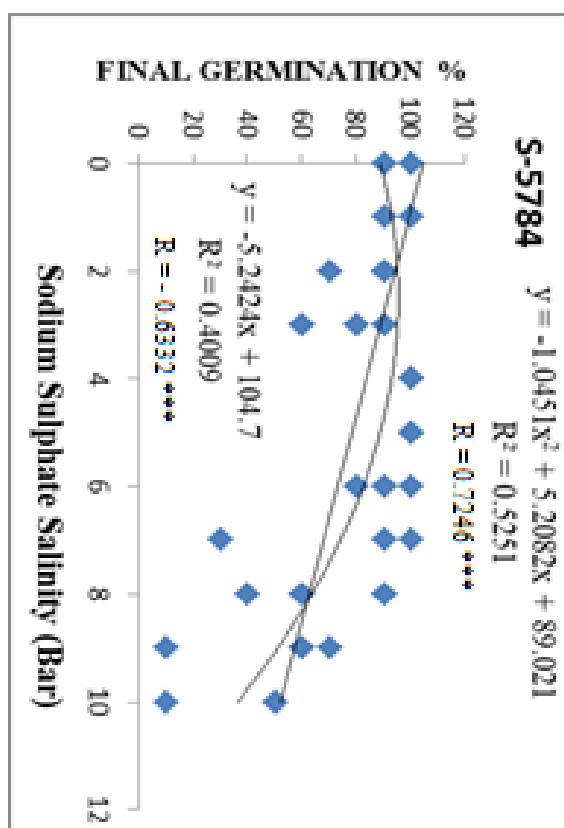
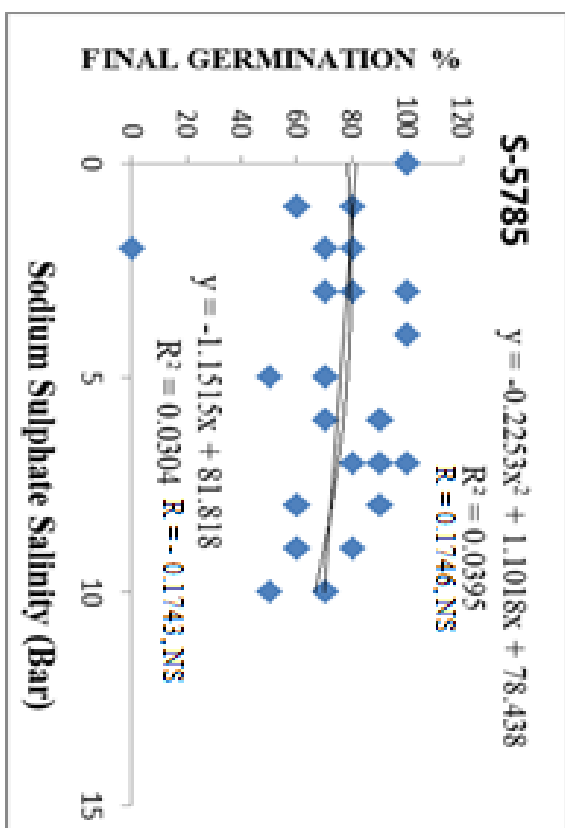
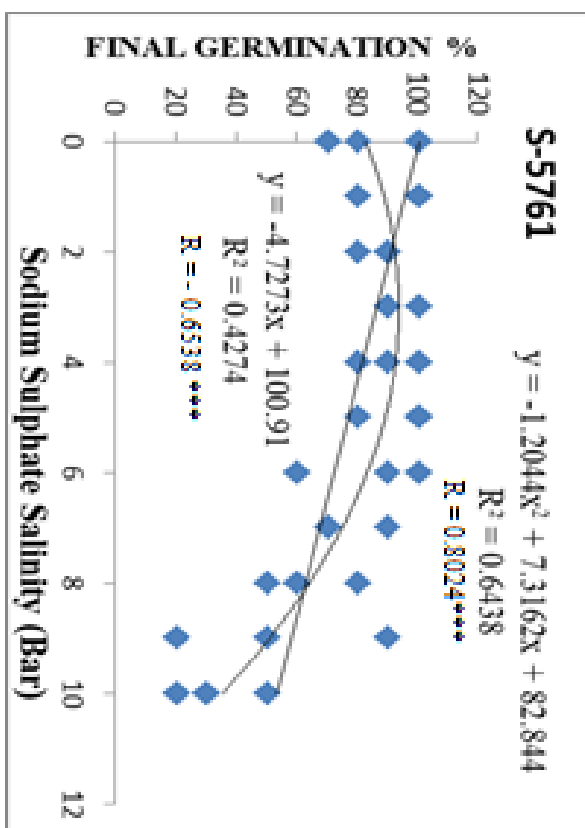
*, $p < 0.05$

**, $p < 0.01$

***, $p < 0.0011$







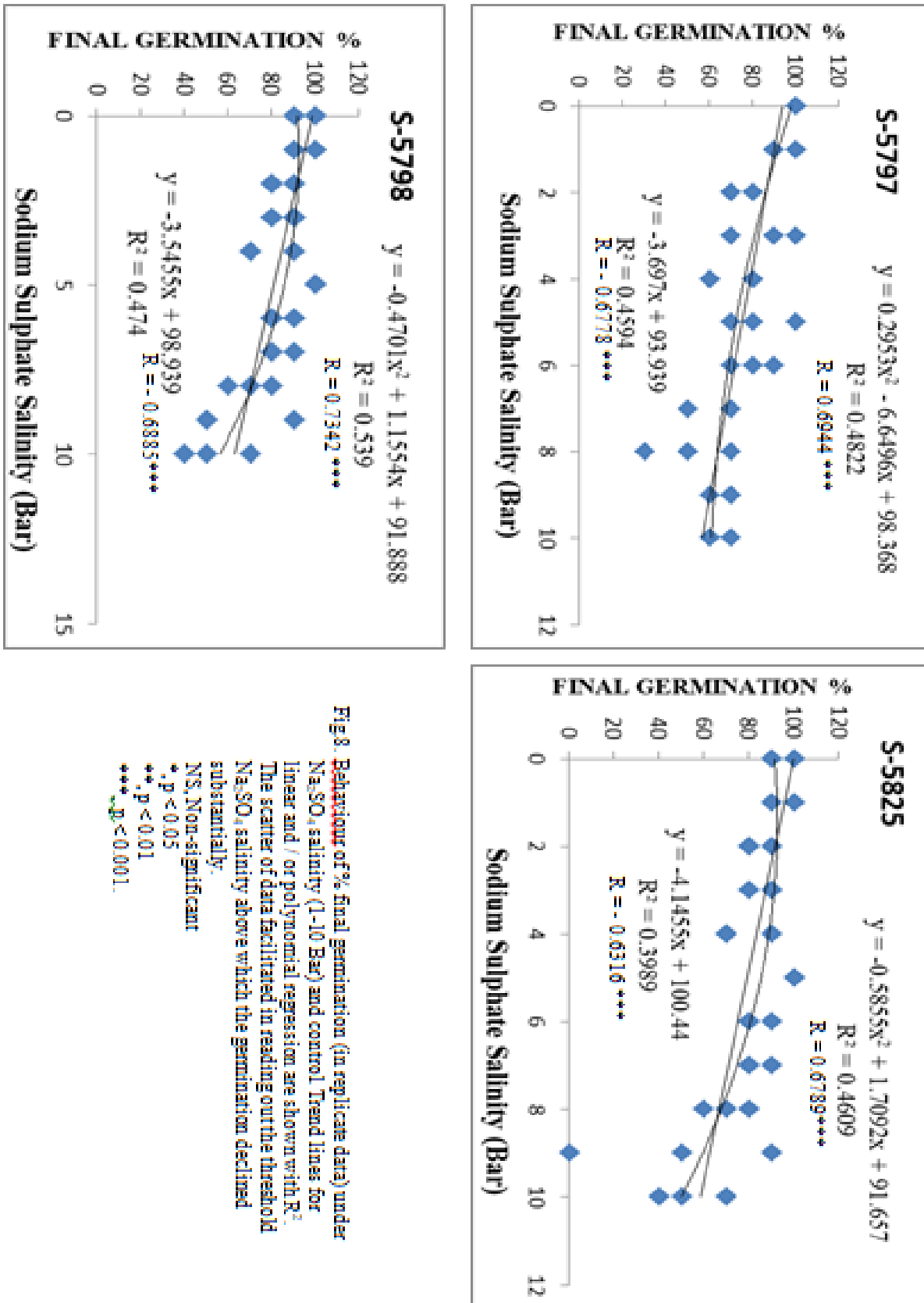


Fig 8. Behaviour of % final germination (in replicate data) under Na_2SO_4 salinity (1-10 Bar) and control. Trend lines for linear and / or polynomial regression are shown with R^2 . The scatter of data facilitated in reading out the threshold Na_2SO_4 salinity above which the germination declined substantially.

NS, Non-significant
 *, $p < 0.05$
 **, $p < 0.01$
 ***, $p < 0.001$.

Table 6. The relationships of Germination velocity index (GVI, Y) with NaCl salinization (X = bar) for 15 guar varieties / lines.

| Varieties / Lines | Best fit regression equations |
|-------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| BR-2/1 | $Y = 27.907 - 1.179 (\text{Bar}) \pm 1.969$; $t_a = 25.13$ ($p < 0.0001$), $t_b = -6.28$ ($p < 0.0001$) $R = 0.902$, $R^2 = 0.814$, $F = 39*.43$ ($p < 0.0001$) <i>Eq. 1</i> |
| BR-90 | $Y = 28.516 - 0.700 (\text{Bar}) \pm 1.321$; $t_a = 38.27$ ($p < 0.0001$), $t_b = -5.557$ ($p < 0.0001$) $R = 0.884$, $R^2 = 0.781$, $F = 30.882$ ($p < 0.0001$) <i>Eq. 2</i> |
| BR -99 | $Y = 25.591 - 0.049 (\text{bar}) \pm 1.192$; $t_a = 38.07$ ($p < 0.0001$), $t_b = -0.434$ ($p < 0.675$) $R = 0.141$, $R^2 = 0.020$, $F = 0.188$ (NS) <i>Eq. 3</i> |
| BR-2017 | $Y = 14.290 + 0.572 (\text{Bar}) - 0.143 (\text{Bar})^2 \pm 1.623$, $t_a = 11.56$ ($p < 0.0001$), $t_{b1} = 0.994$ ($p < 0.3498$), $t_{b2} = -2.59$ ($p < 0.032$); $R = 0.908$, $R^2 = 0.824$, $F = 18.84$ ($p < 0.001$) <i>Eq. 4</i> |
| S-5733 | $Y = 27.089 - 1.805 (\text{Bar}) \pm 2.20$; $t_a = 21.83$ ($p < 0.0001$), $t_b = -9.87$ ($p < 0.0001$) $R = 0.944$, $R^2 = 0.892$, $F = 74.11$ ($p < 0.0001$) <i>Eq. 5</i> |
| S- 5742 | $Y = 19.840 - 1.108 (\text{Bar}) \pm 1.177$; $t_a = 29.87$ ($p < 0.0001$), $t_b = -9.87$ ($p < 0.0001$) $R = 0.957$, $R^2 = 0.915$, $F = 97.38$ ($p < 0.0001$) <i>Eq. 6</i> |
| S-5747 | $Y = 16.200 - 0.598 (\text{Bar}) \pm 1.289$; $t_a = 22.28$ ($p < 0.0001$), $t_b = -4.86$ ($p < 0.0001$) $R = 0.851$, $R^2 = 0.724$, $F = 23.613$ ($p < 0.0001$) <i>Eq. 7</i> |
| S-5761 | $Y = 16.481 - 0.920 (\text{Bar}) \pm 1.338$; $t_a = 21.84$ ($p < 0.0001$), $t_b = -7.20$ ($p < 0.0001$) $R = 0.934$, $R^2 = 0.872$, $F = 52.06$ ($p < 0.0001$) <i>Eq. 8</i> |
| S - 5765 | $Y = 14.217 + 0.503 (\text{Bar}) - 0.135 (\text{Bar})^2 \pm 0.743$; $t_a = 25.52$ ($p < 0.0001$), $t_{b1} = 1.911$ ($p < 0.097$), $t_{b2} = -0.503$ ($p < 0.001$), $R = 0.977$, $R^2 = 0.944$, $F = 84.67$ ($p < 0.0001$) <i>Eq. 9</i> |
| S-5784 | $Y = 14.107 - 1.059 (\text{Bar}) \pm 0.856$; $t_a = 29.22$ ($p < 0.0001$), $t_b = -12.98$ ($p < 0.0001$) $R = 0.974$, $R^2 = 0.949$, $F = 168.56$ ($p < 0.0001$) <i>Eq. 10</i> |
| S-5785 | $Y = 13.838 - 0.590 (\text{bar}) \pm 1.385$, $t_a = 17.71$ ($p < 0.0001$), $t_b = -4.470$ ($p < 0.002$) $R = 0.830$, $R^2 = 0.689$, $F = 19.984$ ($p < 0.002$) <i>Eq. 11</i> |
| S-5797 | $Y = 16.549 - 0.834 (\text{Bar}) \pm 1.188$, $t_a = 24.694$ ($p < 0.0001$), $t_b = -7.37$ ($p < 0.0001$) $R = 0.926$, $R^2 = 0.858$, $F = 54.25$ ($p < 0.0001$) <i>Eq. 12</i> |
| S-5798 | $Y = 13.782 + 0.633 (\text{Bar}) - 0.150 (\text{Bar})^2 \pm 1.927$; $t_a = 9.39$ ($p < 0.0001$), $t_{b1} = 0.926$ ($p < 0.3810$), $t_{b2} = -2.28$ ($p < 0.050$), $R = 0.880$, $R^2 = 0.774$, $F = 13.73$ ($p < 0.003$) <i>Eq. 13</i> |
| S-5759 | $Y = 15.992 - 0.644 (\text{Bar}) \pm 1.212$, $t_a = 23.30$ ($p < 0.0001$), $t_b = -5.57$ ($p < 0.0001$) $R = 0.880$, $R^2 = 0.775$, $F = 31.01$ ($p < 0.0001$) <i>Eq. 14</i> |
| S-5825 | $Y = 18.931 - 0.484 (\text{Bar}) \pm 2.077$, $t_a = 15.87$ ($p < 0.0001$), $t_b = -5.224$ ($p < 0.0001$) $R = 0.867$, $R^2 = 0.752$, $F = 27.29$ ($p < 0.0001$) <i>Eq. 15</i> |

The relationship of GVI with NaCl and Na₂SO₄ salinization is presented in Table 6 and 7 in form of best fit regression equations and values of correlation coefficients 'r', F, and t.

Most of the germplasms exhibited linear inverse relationship but three germplasms viz. S-5765, S-5798 and Br-2017 exhibited curvilinear (polynomial) relationship. Germplasm BR-99 showed non-significant linear correlation as given above and exhibited promotion by 6.25% in GVI almost throughout the treatment range over control (Fig. 9).

The order of salinity tolerance of germplasms against NaCl, based on the decreasing magnitude of 'b' of linear regression between GVI and NaCl salinity (expressed as bar value) is as follows.

S-5733 ($b = -1.805$) < BR- 2/1 ($b = -1.179$) < S-5742 ($b = -1.108$) < S-5784 ($b = -1.059$) < S-5761 ($b = -0.920$) < S-5797 ($b = -0.834$) < BR- 90 ($b = -0.700$) < S-5759 ($b = -0.644$) < S-5747 ($b = -0.598$) < S-5785 ($b = -0.590$) < S-5825 ($b = -0.488$) (excluding BR-99 showing no correlation and Br-2017, S-5765 and S-5798 showing polynomial relationship).

That is to say that S-5733, BR-2/1, S5742 and S-5784 were more sensitive accessions as regards to the GVI decline. The sequential order of salinity tolerance against NaCl in terms of 50% reduction in germination velocity (GVI) as predicted on the basis of above equations (see Table 8) appeared to be as given below (50% reduction in

GVI for BR-2017, S-5765 and S-5798 was calculated on the basis of their curvilinear equations (bar values shown in parenthesis):

NaCl salinity:

S-5784 (6.66 bar) < S-5733 (7.50 bar) < S-5798 (8.55 bar) \approx S-5742 (8.95 bar) = S-5761 (8.95 bar) < BR-2017 (9.36 bar) < S-5797 (9.92 bar) < S-5765 (10.02 bar) < S-5785 (11.73bar) < S- BR-2/1 (11.83 bar) < S-5759 (12.42 bar) < S-5747 (13.55 bar) < S-5825 (19.56 bar) < BR-90 (20.36 bar) < BR-99 (exhibited promotion by 6.25% in 10 bar NaCl).

Clearly, BR-99 showed the best germination rate and was the most tolerant to NaCl salinity as regards to the germination velocity.

The sequential order of salinity tolerance of germplasms against Na₂SO₄, based on the decreasing magnitude of 'b' of linear regression between GVI and Na₂SO₄ salinity is given below. The order excludes two germplasms viz. S-5765 and S-5785 which exhibited curvilinear relationship:

Na₂SO₄ salinity:

S-5733 (-2.765) < BR-90 (-2.284) < BR-2/1 (- 2.024) < S-5742 (- 1.759) < BR-99 (- 1.606) < S-5761 (- 1.575) < S-5759 (-1.255) < S-5784 (-1.123) < BR-2017 (-1.140) < S-5797 (-1.1062) < S-5747 (-1.070) < S-5825 (-0.902) > **S-5798 (-0.562).**

Table 7. The relationships of Germination velocity index (GVI) over 0-10 bar of Na₂SO₄ salinization as obtained with linear regression between GVI (Y) and salinity in bar (X). .

| Varieties / Lines | Best fit regression equations |
|-------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| BR-2/1 | $Y = 25.945 - 2.024 (\text{Bar}) \pm 0.901$; $t_a = 51.08$ ($p < 0.0001$), $t_b = - 23.58$ ($p < 0.0001$) $R = 0.992$, $R^2 = 0.984$, $F = 555.77$ ($p < 0.0001$) Eq. 1 |
| BR-90 | $Y = 28.678 - 2.284 (\text{Bar}) \pm 0.846$; $t_a = 60.084$ ($p < 0.0001$), $t_b = - 28.31$ ($p < 0.0001$) $R = 0.994$, $R^2 = 0.989$, $F = 326.0882$ ($p < 0.0001$) Eq. 2 |
| BR -99 | $Y = 21.238 - 1.606 (\text{bar}) \pm 1.765$; $t_a = 21.33$ ($p < 0.0001$), $t_b = - 9.547$ ($p < 0.0001$) $R = 0.954$, $R^2 = 0.989$, $F = 91.14$ ($p < 0.0001$) Eq. 3 |
| BR-2017 | $Y = 15.481 - 1.140 (\text{Bar}) \pm 1.736$, $t_a = 15.81$ ($p, 0.0001$), $t_b1 = - 8.457$ ($p < 0.0001$) $R = 0.0.938$, $R^2 = 0.888$, $F = 71.51$ ($p < 0.001$) Eq. 4 |
| S-5733 | $Y = 28.686 - 2.765 (\text{Bar}) \pm 2.143$; $t_a = 25.17$ ($p < 0.0001$), $t_b = - 14.34$ ($p < 0.0001$) $R = 0.979$, $R^2 = 0.958$, $F = 91.43$ ($p < 0.0001$) Eq. 5 |
| S- 5742 | $Y = 22.112 - 1.759 (\text{Bar}) \pm 2.429$; $t_a = 16.13$ ($p < 0.0001$), $t_b = - 7.59$ ($p < 0.0001$) $R = 0.930$, $R^2 = 0.865$, $F = 57.67$ ($p < 0.0001$) Eq. 6 |
| S-5747 | $Y = 16.616 - 1.07 (\text{Bar}) \pm 0.806$; $t_a = 36.54$ ($p < 0.0001$), $t_b = -13.92$ ($p < 0.0001$) $R = 0.978$, $R^2 = 0.956$, $F = 193.89$ ($p < 0.0001$) Eq. 7 |
| S-5761 | $Y = 18.260 - 1.575 (\text{Bar}) \pm 1.922$; $t_a = 16.84$ ($p < 0.0001$), $t_b = - 8.59$ ($p < 0.0001$) $R = 0.944$, $R^2 = 0.891$, $F = 73.823$ ($p < 0.0001$) Eq. 8 |
| S – 5765 | $Y = 13.328 + 0.875 (\text{Bar}) - 0.179 (\text{Bar})^2 \pm 1.586$; $t_a = 11.028$ ($p, 0.0001$) $t_b1 = 1.555$ ($p < 0.158$), $t_b2 = - 3.304$ ($p, 0.011$), $R = 0.925$, $R^2 = 0.856$, $F = 23.75$ ($p < 0.0001$) Eq. 9 |
| S-5784 | $Y = 16.494 - 1.123 (\text{Bar}) \pm 2.081$; $t_a = 14.05$ ($p < 0.0001$), $t_b = - 5.660$ ($p < 0.0001$) $R = 0.884$, $R^2 = 0.781$, $F = 32.04$ ($p < 0.0001$) Eq. 10 |
| S-5785 | $Y = 11.376 + 1.087 (\text{Bar}) - 0.160 (\text{Bar})^2 \pm 1.466$, $t_a = 10.188$ ($p, 0.0001$), $t_b1 = 2.093$ ($p < 0.070$), $Tb2 = - 3.20$ ($p < 0.013$), $R = 0.865$, $R^2 = 0.748$, $F = 11.88$ ($p < 0.0001$) Eq. 11 |
| S-5797 | $Y = 15.840 - 1.062 (\text{Bar}) \pm 2.091$, $t_a = 13.427$ ($p, 0.0001$), $t_b = - 5.33$ ($p, 0.00001$) $R = 0.871$, $R^2 = 0.759$, $F = 28.36$ ($p < 0.0001$) Eq. 12 |
| S-5798 | $Y = 14.044 - 0.562 (\text{Bar}) \pm 1.242$; $t_a 20.05$ ($p < 0.0001$), $t_b = - 4.75$ ($p < 0.3810$) $R = 0.845$, $R^2 = 0.715$, $F = 22.59$ ($P < 0.001$) Eq. 13 |
| S-5759 | $Y = 16.481 - 1.255 (\text{Bar}) \pm 2.077$, $t_a = 14.07$ ($p < 0.0001$), $t_b = - 6.339$ ($p < 0.0001$) $R = 0.904$, $R^2 = 0.817$, $F = 4018$ ($p < 0.0001$) Eq. 14 |
| S-5825 | $Y = 17.080 - 0.902 (\text{Bar}) \pm 2.013$, $t_a = 15.0$ ($p < 0.0001$), $t_b = - 4.688$ ($p <, 0.0001$) $R = 0.842$, $R^2 = 0.709$, $F = 21.98$ ($p < 0.0001$) Eq. 15 |

The sequential order of salinity tolerance against Na₂SO₄ in terms of 50% reduction in germination velocity (GVI) as predicted from these equations (see Table 8) appeared to be as follows (50% reduction in GVI for S-5765 and S-5785 was calculated on the basis of their curvilinear equations (bar values corresponding to 50% reduction in parenthesis):

Na₂SO₄ salinity:

S-5733 (5.18) < S-5761 (5.79) < BR-2/1 (6.16) \approx BR-90 (6.27) \approx S-5742 (6.29) \approx S-5759 (6.57) \approx BR-99 (6.61) \approx BR-2017 (6.69) < S-5784 (7.34) \approx S-5797 (7.46) \approx S-5747 (7.76) < S-5765 (8.74) < S-5825 (9.47) < S-5785 (10.25) < S-5798 (12.49)

Clearly, line S-5798 was the most tolerant to Na₂SO₄ salinization as regards to GVI.

The reduction or promotion of GVI over control, under NaCl and Na₂SO₄ environment for various germplasms is presented in Fig. 9. GVI reduction for most germplasms was clearly higher in Na₂SO₄ as compared to that in NaCl. The % reduction in GVI averaged to 52.85 ± 3.86 (CV: 32.44%) (Excluding variety BR-99 which showed promotion) under NaCl. *Per cent* reduction of GVI of the germplasms tested averaged to 71.80 ± 3.86 (CV: 20.8%) in Na₂SO₄.

Seven germplasms under NaCl salinity viz. BR-2/1, BR-90, S-5759, S-5765, S-5785, S-5797 and S-5825 showed GVI reduction in lesser magnitude than the average GVI reduction of the germplasms. Germplasms, BR-2017, S-5733, S-5742, S-5761, S-5784 and S-5798 had larger decline than the average GVI. S-5747 had reduction nearly equal to the average GVI reduction.

Reduction of GVI under Na₂SO₄ environment was of higher order than in NaCl. Seven germplasms (S-5759, S-5747, S-5765, S-5785, S-5797 and S-5825) showed lesser than the average reduction where as germplasms, BR-2/1, BR-90, BR-99, BR-2017, S-5733, S-5742 and S-5761 showed larger reduction than the average reduction. Germplasm S-5784 exhibited GVI decline nearly equal to the quantum of average decline.

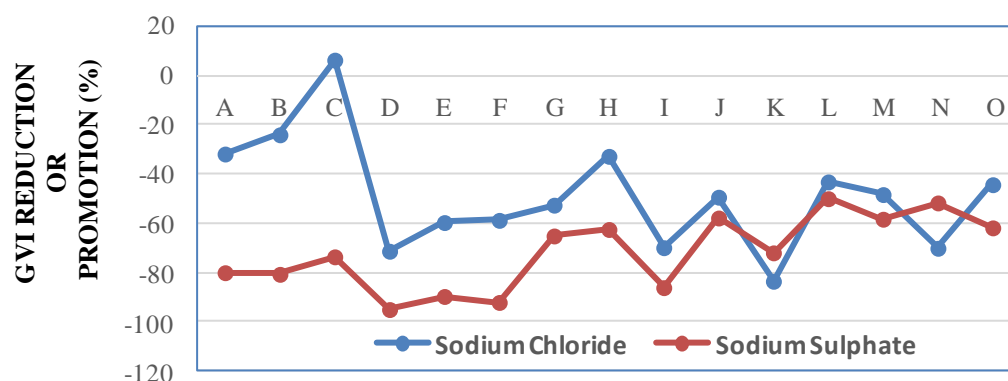


Fig. 9. *Per cent* reduction or promotion (%) of germination velocity index (GVI) in highest salt concentration of over control. Acronyms (A-O): A, BR-2/1; B, BR-90; C, BR-99; D, Br-2017; E, S-5733; F, S-5742; G, S-5747; H, S-5759; I, S-5761; J, S-5765; K, S-5784; L, S-5785; M, S-5797; N, S-5798 and O, S-5825.

Fifty *per cent* reduction in GVI of the germplasms in NaCl and Na₂SO₄ environment is presented in Table 8. Excluding BR-99 which showed promotion (6.25%) even in 10 bar NaCl the magnitude of NaCl salinity causing 50% reduction in GVI varied differentially amongst the germplasms – the lowest in S-5784 (6.66 bar) and the highest in BR-90 (20.36 bar) followed by S-5825 (19.56 bar). It is, however, apparent that NaCl salinity associated with 50% reduction was although differentially varying with germplasms but in all cases it was of quite high order. In terms of EC corresponding to 50 % reduction it ranged from 14.91 to 44.16 dS.m⁻¹.

In Na₂SO₄, fifty per cent decline in GVI occurred in comparatively lower salinities varying from 5.79 bar in S-5761 to 12.49 bar in S-5798. All varietal genotypes (BR-2/1, BR-90, Br-99 and BR-2017) showed relatively higher sensitivity to sulphate salinity which narrowly varied among these genotypes (6.16 to 6.69 bar i.e. 15-16 dS.m⁻¹). These genotypes were more resistant to NaCl salinity. As regards to GVI, lines S-5785 and S-5798 were comparatively more resistant to sulphate salinity than the four varietal genotypes.

Table 8. NaCl and Na₂SO₄ concentrations, in terms of bar values and EC (dS.m⁻¹) corresponding with 50% reduction in Germination velocity index.

| Germplasms | NaCl | Na ₂ SO ₄ |
|------------|----------------|---------------------------------|
| BR-2/1 | 11.83 (25.95)* | 6.16 (15.35)* |
| BR-90 | 20.36 (44.16) | 6.27 (15.60) |
| BR-99 | - | 6.61 (16.37) |
| BR-2017 | 9.36 (20.68) | 6.69 (16.55) |
| S-5733 | 7.50 (16.71) | 5.18 (13.12) |
| S-5742 | 8.95 (19.80) | 6.29 (15.64) |
| S-5747 | 13.55 (29.62) | 7.76 (18.03) |
| S-5761 | 8.95 (19.80) | 5.79 (14.50) |
| S-5765 | 10.02 (22.08) | 8.74 (21.21) |
| S-5784 | 6.66 (14.91) | 7.34 (18.03) |
| S-5785 | 11.73 (25.74) | 10.25 (14.65) |
| S-5797 | 9.92 (21.87) | 7.46 (18.30) |
| S-5798 | 8.55 (18.95) | 12.49 (29.74) |
| S-5759 | 12.42 (27.20) | 6.57 (16.28) |
| S-5825 | 19.56 (42.45) | 9.47 (22.87) |

EC values in parenthesis. The values for BR-2017, S-5785 and S-5798 in case of NaCl salinity and for S-5765 and S-5798 in case of Na₂SO₄ salinity were calculated on the basis of curvilinear equations.

Agglomeration of guar germplasms on the basis of germination and GVI:

The agglomeration of germplasms was done by cluster analysis of Ward (1963) on the basis of germination and GVI data of germplasms using Euclidean distances among the germplasms. The results are presented in form of cluster diagrams (Fig. 10-13).

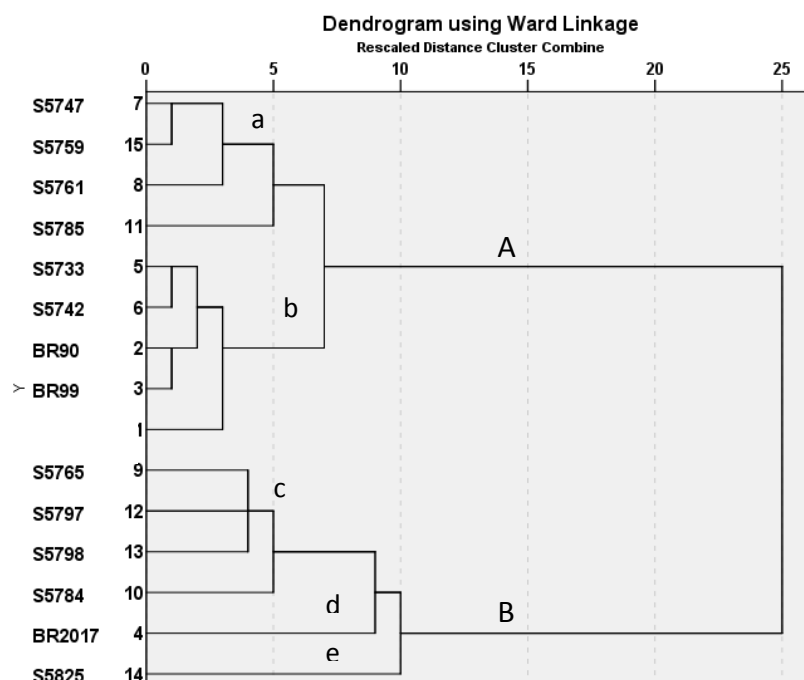


Fig. 10. Agglomerative clustering of guar germplasms on the basis of mean final % germination under NaCl salinization.

Germplasms agglomeration on the basis of germination under NaCl

Fig. 10 presents the cluster diagram of germplasms under NaCl. On the basis of 5% distance amongst the germplasm five discrete (all differentially salt tolerant) groups may be observed and on a distance of 10% only two groups (both salt tolerant) were observed as given below for their composition and their germination potentials.

Dissimilarity Distance: 5%

Group a: S-5747, S-5759, S-5761 and S-5785

(N = 44, mean final germination: 90.682 ± 1.5156 , 66.67-100, CV = 11.09%) – Rank II

Group b: S-5733, S-5742, BR-90, BR-99 and BR-2/1

(N = 55, mean final germination: 95.041 ± 0.78485 , 80-100, CV= 5.85%) – Rank I

Group c: S-5765, S-5797, S-5798 and S-5784

(N = 44, mean final germination: 84.69 ± 2.505 , 30-100, CV: 19.62%) – Rank III

Group d: BR-2017

(N = 11, final germination: 75.46 ± 4.8886 , 43.33 – 90.0, CV: 21.49) – Rank V

Group e: S-5825

(N = 11, final germination: 80.91 ± 3.9204 , 46.67 – 93.33, CV: 16.07%) – Rank IV

All groups of germplasms were considerably tolerant at germination phase but groups a and b predominated the germplasms for germinability.

Dissimilarity Distance: 10%

Group A: S-5747, S-5759, S-5761, S-5785, S-5733, S-5742, BR-90, BR-99 and BR-2/1

N = 99, mean final germination: 93.898 ± 0.8168 , 66.67 -100.0, CV: 8.13%) – Rank I

Group B: S-5765, S-5797, S-5798, S-5784, BR-2017 and S- 5825

(N = 66, mean final germination: 82.526 ± 3.920 , 30.-100, CV: 19.59%) --- Rank II

These two groups were significantly different from each other on the basis their germination behaviour as indicated by the t-test ($t = 4.92$, $p < 0.0001$). Group A had higher germination potential under NaCl salinity than the group B.

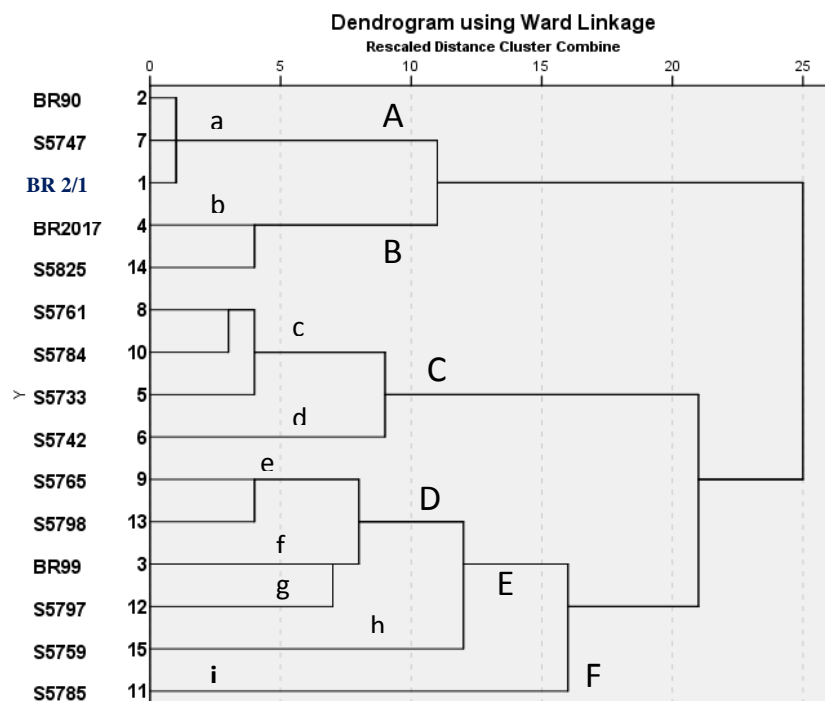


Fig. 11. Agglomerative clustering of guar germplasms on the basis of mean final germination under Na₂SO₄ salinization.

Germplasms agglomeration on the basis of germination under Na₂SO₄

Fig. 11 presents the cluster diagram of germplasms under Na₂SO₄. On the basis of 5% distance amongst the germplasm nine discrete and differentially tolerant groups may be observed and on a distance of 10% six groups were apparent as given below for their composition and their germination behaviour.

Dissimilarity Distance 5%

Group a: BR-90, S-5747 and BR-2/1

(N = 33, mean final germination: 90.00 ± 2.298 , 50-100, CV: 14.66% -- Rank I

Group b: BR-2017, S-5825

(N = 22, mean final germination: 83.79 ± 2.475 , 53.33-100, CV: 13.85%) – Rank III

Group c: S-5761, S-5784 and S-5733

(N = 33, mean final germination: 77.98 ± 3.661 , 33.33-100, CV: 26.97%) – Rank VII

Group d: S-5742

(N = 11, mean final germination: 85.15 ± 8.269 , 16.7-100, CV: 32.20%) – Rank II

Group e: S-5765 and S-5798

(N = 22, mean final germination: 82.72 ± 3.581 , 50.0-100, CV: 20.30%) – Rank V

Group f: BR-99

(N = 11, mean final germination: 83.34 ± 4.669 , 56.67-100, CV: 18.59%) – Rank IV

Group g: S-5797

(N = 11, mean final germination: 75.46 ± 4.633 , 50.0-100, CV: 20.37%) – Rank IX

Group h: S-5759

(N = 11, mean final germination: 78.18 ± 6.5198 , 36.0-100, CV: 27.66%) – Rank VI

Group i: S-5785

(N = 11, mean final germination: 76.06 ± 4.919 , 50-100, CV: 16.32%) – Rank VIII

Dissimilarity Distance 10%

Group A: BR-90, S-5747 and BR-2/1

(N = 33, mean final germination: 90.00 ± 2.298 , 50.0-100, CV: 14.69%) – Rank I

Group B: BR-2017 and S-5825

(N = 22, mean final germination: 83.78 ± 2.2747 , 53.33-100, CV: 13.85%) – Rank II

Group C: S-5761, S-5784, S-5733 and S-5742

(N = 44, mean final germination: 79.77 ± 36.417 , 16.67-100, CV: 28.42%) – Rank IV

Group D: S-5785, S-5798, BR-99 and S-5797

(N = 44, mean final germination = 81.06 ± 2.427 , 50-100, CV: 20.29%) – Rank III

Group E: S-5759

(N = 11, mean final germination: 78.18 ± 6.519 , 36.67-100, CV: 29.66%) – Rank V

Group F: S-5785

(N = 11, mean final germination: 76.06 ± 4.919 , 36.67-100, CV: 21.45%) – Rank VI

Germplasms agglomeration on the basis of mean GVI under NaCl

Fig. 12 presents the GVI- based cluster diagram of germplasms under NaCl. On the basis of 5% distance amongst the germplasm two discrete groups were observed as given below for their composition and their germination behaviour.

Dissimilarity Distance: 5%

Group A: BR-2017, S-5798, S-5797, S- 5759, S-5747, S-5761, S-5765, S-5785, S- 5742, S-5825 and S-5784

Mean GVI over the treatment range and the control = 12.13 ± 0.79176 , CV: 11.89% -- Rank II

Group B: BR-90, BR-99, BR-2/1 and S-5733

(Mean GVI over the treatment range and the control: 22.628 ± 1.502 , CV: 11.89% -- Rank I

The group B of germplasms composed of BR -90, BR -99, BR-2/1 and S-5733 was fast- germinating than germplasms of group A.

Germplasms agglomeration on the basis of mean GVI under Na₂SO₄

Fig. 13 presents the GVI-based cluster diagram of germplasms under Na₂SO₄. On the basis of 5% distance amongst the germplasm five discrete groups were observed and on the basis of 10% distance two groups were apparent as given below for their composition and their germination behaviour. The first ranking group was composed of BR-2/1, BR-99 and S-5733 followed by the second-ranking group composed of BR-99 and S-5742. The clustering was more dispersive under Na₂SO₄.

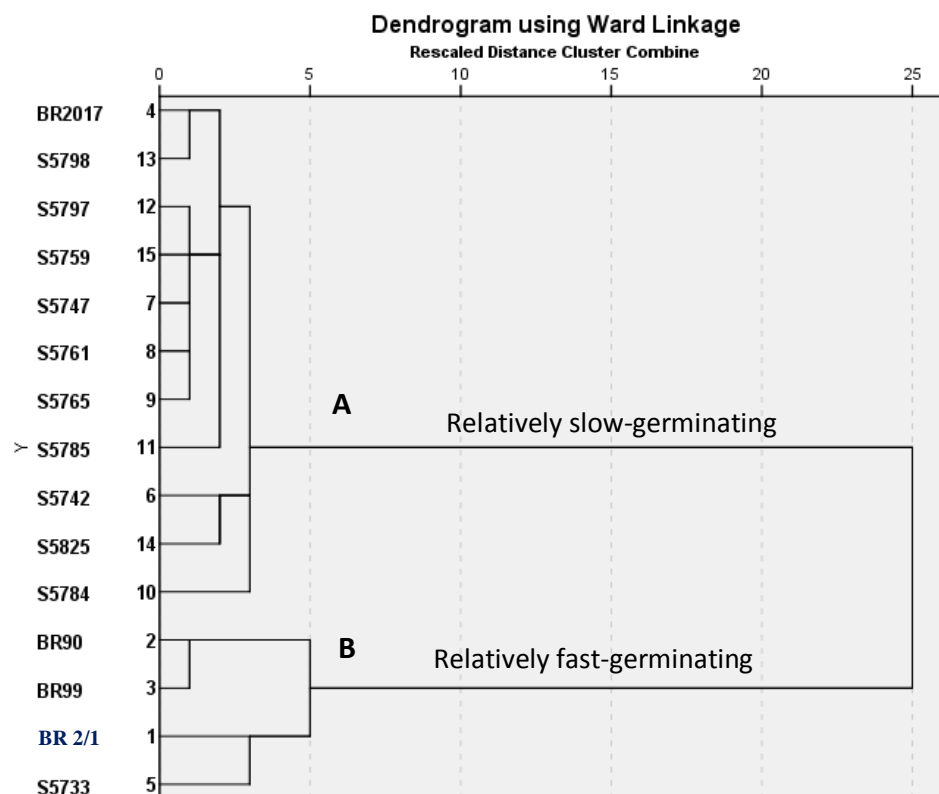


Fig. 12. Agglomerative clustering of guar germplasms based on GVI under NaCl salinization.

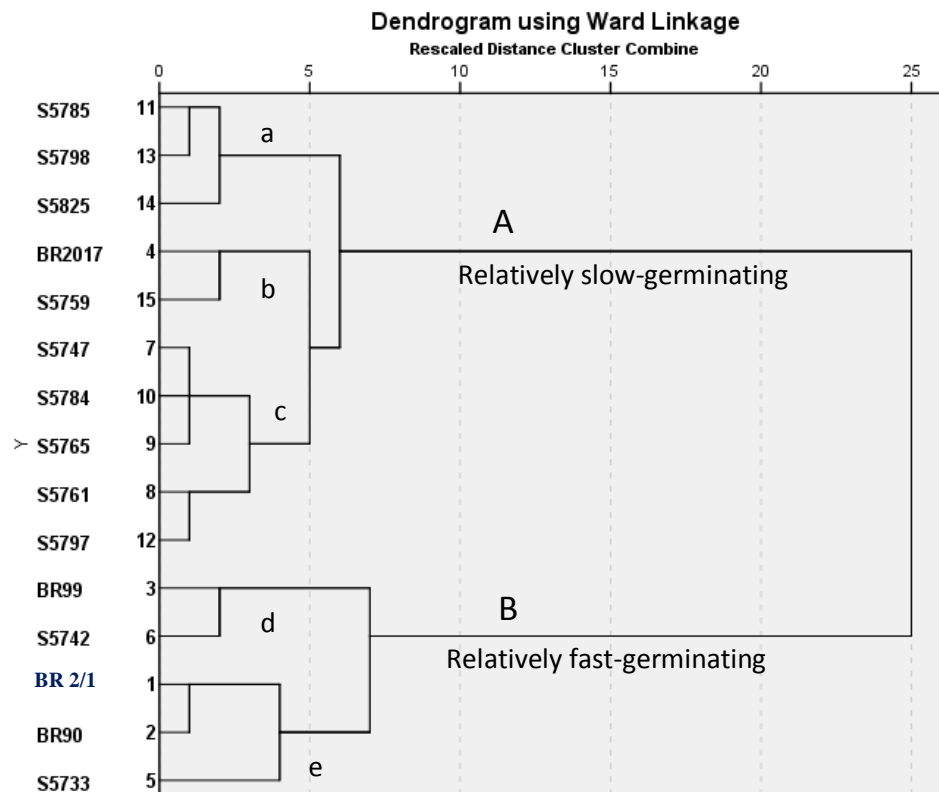


Fig. 13. Agglomerative clustering of guar germplasms based on GVI under Na₂SO₄ salinization.

Dissimilarity Distance: 5%**Group a: S-5785, S-5798 and S-5825**

Mean GVI over the treatment and control: 11.64 ± 0.45 , CV: 6.68% -- Rank III

Group b: BR-2017 and S-5759

Mean GVI over the treatment range and the control: 9.37 ± 0.86 , CV: 13.02% -- Rank V

Group c: S-5747, S-5784, S-5765, S-5761 and S-5797

Mean GVI over treatment range and the control: 10.90 ± 0.2032 , CV: 4.96 -- Rank IV

Group d: BR-99, S-5742,

Mean GVI over the treatment range and the control: 13.23 ± 0.0270 , CV: 0.30% -- Rank II

Group e: BR-2/1, BR-90 and S-5733

Mean GVI over the treatment range and the control: 15.99 ± 0.6944 , CV: 7.52% -- Rank I

Dissimilarity Distance: 10%**Group A: S-5785, S-5798, S-5825, BR-2017, S-5759, S-5742, S-5784, S-5765, S-5761 and S-5797**

Mean GVI over the treatment range and the control: 10.82 ± 0.3351 , CV: 9.79% -- Rank II

Group B: BR-99, S-5742, BR-2/1, BR-90 and S-5733

Mean GVI over the treatment range and the control: 14.84 ± 0.7607 , CV: 11.38% -- Rank I

Group B composed of BR-99, BR-2/1, S-5742 and S-5733 was comparatively more rapid in germination.

Seedling growth

Under significant germination delaying conditions under salinity as seen in the present work, seedling growth *in vitro* experiment of short period of time (four days) appears to be somewhat dubious due to disparity of age amongst the seedlings. However, since this age disparity is caused by differential response of the germplasms to salinity, seedling behaviour is often described in such studies. It is given below.

Seedling growth in the present work was assessed on three parameters – root length, shoot length and cumulative seedling dry weight after four days of incubation. Two-way ANOVA for these parameters is presented in Table 9-14. Analysis of variance for the component sources of variation of the root length under NaCl and Na₂SO₄ salinities is presented in Table 9 and 10. The both components sources significantly influenced germination under two types of salinities and the nature of varieties and their interactions were also significant.

During growth phase, salt stress is first perceived by the root. Both, NaCl and Na₂SO₄ regularly reduced the root length. The root length of various germplasms varied differentially. Root length was maximum in line in BR-99 and S-5765 and minimum in line S-5797. Root length varied between 2 to 3 cm in six germplasms (S-5825, BR-2/1, S-5742, BR-90, S-5747 and S-5759). Genotypes such as BR-2017, S-5761, S-5784, S-5785, S-5733 and S-5798 had root varying between 1 and 2 cm in length. Under Na₂SO₄ environment, the maximum root length was observed in S-5825 (2.44 cm) followed in S-5797 (0.56 cm). The root length of BR-99 was more affected under Na₂SO₄ salinity (Table 10). Generally, Na₂SO₄ salinity was more suppressive to root length than NaCl. The component sources of variation (salt concentration and nature of varieties) significantly affected the shoot growth and their interaction was also found to be significant. Under this interactive system, like NaCl salinity regularly declined shoot elongation in Na₂SO₄ (0 to 10 bar) but comparatively in larger magnitude than under NaCl i.e. Na₂SO₄ salinity was more detrimental to shoot growth than NaCl. Still BR-2/1 (2.81cm), BR-90 (2.51cm), S-5733 (2.28cm) S-5825 (2.19 cm) and BR-99 (2.09 cm) had root length larger or equal to 2 cm under sulphate salinity. Other germplasms showed root elongation between 1 and 1.7 cm (Table 11 and 12).

Analysis of variance for seedling growth under chloride and sulphate salinities is presented in Table 13 and 14, respectively. The salt concentration and varietal nature presented a highly interactive system as regard to the cumulative dry seedling mass under NaCl and Na₂SO₄ salinities. On four- day- interaction, seedling dry mass was observed to be the maximum in 7 bar NaCl followed by 5 bar salinity. In most germplasms seedling dry mass was higher in all treatments of NaCl as compared to control – only except that in 10 bar salinity (Table 13).

The sequential order of seedling mass productivity under two types of salinity (Table 13 and 14) indicated that seedling biomass was the highest in S-5784 (0.2115g) and the minimum in S-5761 (0.1216g). Under sulphate salinity, maximum seedling dry mass was represented by S-5733 (0.1933g) followed by S-5742 (0.1924g). Seedling mass was minimum in BR-99 (0.1199g). Like root and shoot lengths, seedling mass was more suppressed in sulphate salinity (Table 14). *Prosopis juliflora* (a known salt tolerant plant; Khan *et al.*, 1987) showed increase in shoot growth over control up to 8 bar NaCl and only up to 5 bar Na₂SO₄ concentration. In seedlings of this species, root length was larger up to 7 bar NaCl and was maximum at 4 bar (Khan *et al.*, 1984).

Table 9. Analysis of variance for root length data (cm) of 15 guar varieties tested against 11 concentrations of NaCl.

| Source | SS | df | MS | F | p |
|---------------------------|---------|-----|-------|-------|-------------|
| NaCl concentrations | 278.218 | 10 | 27.82 | 68.19 | 0.00001 *** |
| Varieties | 200.572 | 14 | 14.33 | 35.12 | 0.00001 *** |
| Concentration x varieties | 185.859 | 140 | 1.33 | 3.250 | 0.00001 ** |
| Error | 134.633 | 330 | 0.408 | - | - |
| Total | 799.282 | 494 | - | - | - |

| NaCl Concentrations LSD 0.0-5:0.268134 | | | | | Varieties or lines, LSD 0.05: 0.309329 | | | | |
|----------------------------------------------------------------------|---------|--------|----|-----|----------------------------------------|---------|-------|----|-----|
| Rank | TRT | Mean | n | NSR | Rank | TRT | Mean | n | NSR |
| 1 | Control | 3.344 | 45 | a | 1 | Br-99 | 3.200 | 33 | a |
| 2 | 1 bar | 2.856 | 45 | b | 2 | S-5765 | 3.119 | 33 | a |
| 3 | 2 bar | 2.669 | 45 | bc | 3 | S-5825 | 2.560 | 33 | b |
| 4 | 3bar | 2.651 | 45 | bc | 4 | BR-2/1 | 2.510 | 33 | b |
| 5 | 4 bar | 2.517 | 45 | cd | 5 | S-5742 | 2.440 | 33 | b |
| 6 | 5 bar | 2.273 | 45 | d | 6 | Br-90 | 2.396 | 33 | b |
| 7 | 6 bar | 1.923 | 45 | e | 7 | S-5747 | 2.361 | 33 | b |
| 8 | 7 bar | 1.779 | 45 | e | 8 | S-5759 | 2.297 | 33 | bc |
| 9 | 8 bar | 1.366 | 45 | f | 9 | BR-2017 | 1.988 | 33 | cd |
| 10 | 9 bar | 1.0584 | 45 | g | 10 | S-5761 | 1.864 | 33 | de |
| 11 | 10 bar | 0.885 | 45 | g | 11 | S-5785 | 1.842 | 33 | de |
| TRT, Treatments (salt concentrations or varieties; NSR, NS ranges | | | | | 12 | S-5733 | 1.571 | 33 | ef |
| | | | | | 13 | S-5784 | 1.435 | 33 | fg |
| | | | | | 14 | S-5798 | 1.210 | 33 | gh |
| | | | | | 15 | S-5797 | 0.936 | 33 | h |

Table 10. Analysis of variance of root length data (cm) of 15 guar varieties tested against 11 concentrations of Na₂SO₄.

| Source | SS | df | MS | F | p |
|------------------------------------------------|---------|-----|--------|---------|-------------|
| Na ₂ SO ₄ concentrations | 403.84 | 10 | 40.384 | 118.111 | 0.00001 *** |
| Varieties | 117.53 | 14 | 8.395 | 24.553 | 0.0000 *** |
| Concentration x varieties | 205.09 | 140 | 1.465 | 4.284 | 0.0000 *** |
| Error | 112.83 | 330 | 0.342 | - | - |
| Total | 839.289 | 494 | - | - | - |

| Na ₂ SO ₄ Concentrations LSD 0.0-5: 0.2424994 | | | | | Varieties or lines, LSD 0.05: 0.283178 | | | | |
|----------------------------------------------------------------------|---------|-------|----|-----|----------------------------------------|---------|-------|----|------|
| Rank | TRT | Mean | n | NSR | Rank | TRT | Mean | n | NSR |
| 1 | Control | 3.344 | 45 | a | 1 | S-5825 | 2.437 | 33 | a |
| 2 | 1 bar | 2.837 | 45 | b | 2 | S-5747 | 2.385 | 33 | a |
| 3 | 2 bar | 2.083 | 45 | c | 3 | S-5742 | 1.962 | 33 | b |
| 4 | 4 bar | 2.018 | 45 | c | 4 | S-5765 | 1.911 | 33 | bc |
| 5 | 3 bar | 1.662 | 45 | d | 5 | BR-2017 | 1.862 | 33 | bc |
| 6 | 5 bar | 1.516 | 45 | d | 6 | S-5785 | 1.687 | 33 | bcd |
| 7 | 6 bar | 1.167 | 45 | e | 7 | S-5759 | 1.637 | 33 | cde |
| 8 | 7 bar | 1.013 | 45 | e | 8 | BR-90 | 1.503 | 33 | def |
| 9 | 8 bar | 0.594 | 45 | f | 9 | BR-2/1 | 1.446 | 33 | defg |
| 10 | 9 bar | 0.552 | 45 | f | 10 | S-5733 | 1.356 | 33 | efg |
| 11 | 10 bar | 0.535 | 45 | f | 11 | S-5761 | 1.303 | 33 | fg |
| TRT, Treatments (salt concentrations or varieties; NSR, NS ranges | | | | | 12 | Br-99 | 1.212 | 33 | fgh |
| | | | | | 13 | S-5798 | 1.175 | 33 | gh |
| | | | | | 14 | S-5784 | 0.992 | 33 | h |
| | | | | | 15 | S-5797 | 0.561 | 33 | h |

The average seedling dry mass of various germplasms and within germplasms seedling-mass-variation in terms of Coefficient of Variation (CV) under the treatment range (0 to 10 bar) salinities is presented in Table 15. Maximum variation in seedling mass under NaCl was exhibited by S-5733 (40.68%) followed by S-5761 (39.83%), S-5765 (34.91%) and S-5797 (31.23%). The minimum variation was found in S-5798 (4.70%) followed by S-5747 (6.68%). Seedling mass variation was around 20% in S-5785, S-5784 and BR-90. Low order variation (below 15%) was exhibited in BR-2/1, BR-99, S-5742 and S-5825. Under sulphate salinity, variation of seedling mass was generally

of higher order- maximum in S-5761 (53.77%) followed by S-5784, BR-2017, S-5733, S-5784 and S-5798 (fluctuating between 30-35%). Variation was minimal in S-5765 (10.10%) followed by S-5747 and BR-2/1 (around 12%).

Table 11. Analysis of variance for shoot length data (cm) of 15 guar varieties tested against 11 concentrations of NaCl.

| Source | SS | df | MS | F | p |
|---------------------------|-----------|-----|---------|--------|-------------|
| NaCl concentrations | 372.322 | 10 | 37.23 | 124.88 | 0.00001 *** |
| Varieties | 397.398 | 14 | 28.39 | 95.206 | 0.00001 *** |
| Concentration x varieties | 301.9313 | 140 | 2.159 | 7.233 | 0.00001 *** |
| Error | 98.3891 | 330 | 0.29814 | - | - |
| Total | 1170.0471 | 494 | - | - | - |

| NaCl Concentrations LSD 0.0-5: 0.22645 | | | | | Varieties or lines, LSD 0.05: 0.264435 | | | | |
|----------------------------------------------------------------------|---------|-------|----|-----|----------------------------------------|---------|------|----|-----|
| Rank | TRT | Mean | n | NSR | Rank | TRT | Mean | n | NSR |
| 1 | Control | 3.765 | 45 | a | 1 | BR-2/1 | 4.30 | 33 | a |
| 2 | 1 bar | 3.271 | 45 | b | 2 | BR-90 | 4.05 | 33 | ab |
| 3 | 2bar | 3.243 | 45 | b | 3 | BR-99 | 3.88 | 33 | b |
| 4 | 3 bar | 3.207 | 45 | b | 4 | S-5733 | 2.72 | 33 | c |
| 5 | 4 bar | 2.574 | 45 | c | 5 | S-5765 | 2.24 | 33 | d |
| 6 | 5bar | 2.358 | 45 | c | 6 | S-5825 | 2.21 | 33 | d |
| 7 | 6 bar | 1.873 | 45 | d | 7 | S-5759 | 1.99 | 33 | de |
| 8 | 7 bar | 1.778 | 45 | d | 8 | S-5797 | 1.97 | 33 | def |
| 9 | 8 bar | 1.537 | 45 | e | 9 | S-5742 | 1.86 | 33 | ef |
| 10 | 9 bar | 1.372 | 45 | e | 10 | S-5784 | 1.84 | 33 | ef |
| 11 | 10 bar | 1.090 | 45 | f | 11 | S-5785 | 1.76 | 33 | ef |
| TRT, Treatments (salt concentrations or varieties; NSR, NS ranges | | | | | 12 | BR-2017 | 1.72 | 33 | ef |
| | | | | | 13 | S-5747 | 1.70 | 33 | ef |
| | | | | | 14 | S-5798 | 1.69 | 33 | ef |
| | | | | | 15 | S-5761 | 1.66 | 33 | f |

Table 12. Analysis of variance of shoot length data (cm) of 15 guar varieties tested against 11 concentrations of Na₂SO₄.

| Source | SS | df | MS | F | p |
|------------------------------------------------|---------|-----|---------|--------|-------------|
| Na ₂ SO ₄ concentrations | 382.81 | 10 | 38.28 | 159.15 | 0.00001 *** |
| Varieties | 122.54 | 14 | 8.753 | 36.39 | 0.0000 *** |
| Concentration x varieties | 282.91 | 140 | 2.020 | 8.40 | 0.0000 *** |
| Error | 79.399 | 330 | 0.24054 | - | - |
| Total | 867.642 | 494 | - | - | - |

| Na ₂ SO ₄ Concentrations LSD 0.0-5: 0.203399 | | | | | Varieties or lines, LSD 0.05: 0.237519 | | | | |
|----------------------------------------------------------------------|---------|-------|----|-----|----------------------------------------|---------|-------|----|-----|
| Rank | TRT | Mean | n | NSR | Rank | TRT | Mean | n | NSR |
| 1 | Control | 3.79 | 45 | a | 1 | BR-2/1 | 2.81 | 33 | a |
| 2 | 1 bar | 2.73 | 45 | b | 2 | BR-90 | 2.51 | 33 | b |
| 3 | 2 bar | 2.26 | 45 | c | 3 | S-5733 | 2.28 | 33 | bc |
| 4 | 3 bar | 2.04 | 45 | d | 4 | S-5825 | 2.19 | 33 | c |
| 5 | 4bar | 1.72 | 45 | e | 5 | BR-99 | 2.09 | 33 | c |
| 6 | 5 bar | 1.69 | 45 | e | 6 | S-5747 | 1.70 | 33 | d |
| 7 | 6 bar | 1.43 | 45 | f | 7 | S-5759 | 1.699 | 33 | d |
| 8 | 7 bar | 1.22 | 45 | g | 8 | S-5742 | 1.664 | 33 | de |
| 9 | 8 bar | 0.909 | 45 | h | 9 | S-5785 | 1.606 | 33 | de |
| 10 | 9 bar | 0.819 | 45 | h | 10 | S-5765 | 1.504 | 33 | de |
| 11 | 10 bar | 0.717 | 45 | h | 11 | BR-2017 | 1.443 | 33 | def |
| TRT, Treatments (salt concentrations or varieties; NSR, NS ranges | | | | | 12 | S-5798 | 1.399 | 33 | efg |
| | | | | | 13 | S-5784 | 1.210 | 33 | fgh |
| | | | | | 14 | S-5761 | 1.180 | 33 | gh |
| | | | | | 15 | S-5797 | 1.050 | 33 | h |

Table 13. Analysis of variance for collective seedling dry wt. (g) data of 15 guar varieties tested against 11 concentrations of NaCl.

| Source | SS | df | MS | F | p |
|---------------------------|----------|-----|-----------|--------|-------------|
| NaCl concentrations | 0.14784 | 10 | 0.0147839 | 5.268 | 0.00001 *** |
| Varieties | 0.387463 | 14 | 0.384626 | 9.7908 | 0.00001 *** |
| Concentration x varieties | 0.65837 | 140 | 0.004703 | 1.6759 | 0.00001 *** |
| Error | 0.92598 | 330 | 0.002806 | - | - |
| Total | 2.11681 | 494 | - | - | - |

| NaCl Concentrations LSD 0.0-5: 0.021968 | | | | | Varieties or lines, LSD 0.05: 0.025653 | | | | |
|----------------------------------------------------------------------|---------|--------|----|-----|----------------------------------------|---------|---------|----|-----|
| Rank | TRT | Mean | n | NSR | Rank | TRT | Mean | n | NSR |
| 1 | 7 bar | 0.1941 | 45 | a | 1 | S-5784 | 0.21150 | 33 | a |
| 2 | 5 bar | 0.1855 | 45 | ab | 2 | S-5765 | 0.20924 | 33 | a |
| 3 | 4 bar | 0.1788 | 45 | ab | 3 | S-5759 | 0.19739 | 33 | a |
| 4 | 3bar | 0.1747 | 45 | ab | 4 | S-5747 | 0.19045 | 33 | ab |
| 5 | 6 bar | 0.1680 | 45 | bc | 5 | S- 5733 | 0.18969 | 33 | ab |
| 6 | 1 bar | 0.1680 | 45 | bc | 6 | S-5742 | 0.18854 | 33 | ab |
| 7 | 2 bar | 0.1680 | 45 | bc | 7 | Br-2017 | 0.16363 | 33 | bc |
| 8 | 8bar | 0.1628 | 45 | cd | 8 | S-5798 | 0.15785 | 33 | cd |
| 9 | bar | 0.1499 | 45 | cd | 9 | S-5825 | 0.15627 | 33 | cd |
| 10 | Control | 0.1459 | 45 | cd | 10 | BR-2/1 | 0.15303 | 33 | cd |
| 11 | 10 bar | 0.1328 | 45 | d | 11 | BR-99 | 0.14848 | 33 | cde |
| TRT, Treatments (salt concentrations or varieties; NSR, NS ranges | | | | | 12 | S-5797 | 0.14833 | 33 | cde |
| | | | | | 13 | BR-90 | 0.13758 | 33 | cde |
| | | | | | 14 | S-5785 | 0.12894 | 33 | de |
| | | | | | 15 | S-5761 | 0.12164 | 33 | e |

Table 14. Analysis of variance of collective seedling dry wt. (g) data of 15 guar varieties tested against 11 concentrations of Na₂SO₄.

| Source | SS | df | MS | F | p |
|------------------------------------------------|----------|-----|-----------|--------|-------------|
| Na ₂ SO ₄ concentrations | 0.172829 | 10 | 0.017283 | 4.4093 | 0.00001 *** |
| Varieties | 0.284342 | 14 | 0.020310 | 5.1816 | 0.0000 *** |
| Concentration x varieties | 0.87435 | 140 | 0.006245 | 1.5933 | 0.0000 *** |
| Error | 1.293495 | 330 | 0.0039196 | - | - |
| Total | 2.6250 | 494 | - | - | - |

| Na ₂ SO ₄ Concentrations LSD 0.0-5: 0.025964 | | | | | Varieties or lines, LSD 0.05: 0.0303198 | | | | |
|----------------------------------------------------------------------|---------|--------|----|-----|-----------------------------------------|---------|--------|----|------|
| Rank | TRT | Mean | n | NSR | Rank | TRT | Mean | n | NSR |
| 1 | 5 bar | 0.1885 | 45 | a | 1 | S-5733 | 0.1933 | 33 | a |
| 2 | 4bar | 0.1780 | 45 | ab | 2 | S-5742 | 0.1924 | 33 | ab |
| 3 | 2 bar | 0.1679 | 45 | abc | 3 | S-5747 | 0.1874 | 33 | ab |
| 4 | 3 bar | 0.1663 | 45 | abc | 4 | S-5761 | 0.1757 | 33 | abc |
| 5 | 1 bar | 0.1659 | 45 | abc | 5 | S-5784 | 0.1744 | 33 | abc |
| 6 | 6 bar | 0.1624 | 45 | abc | 6 | S-5759 | 0.1742 | 33 | abc |
| 7 | 7bar | 0.1509 | 45 | bc | 7 | BR- 90 | 0.1655 | 33 | abcd |
| 8 | 8 bar | 0.1508 | 45 | bc | 8 | S-5825 | 0.1576 | 33 | bcde |
| 9 | Control | 0.1459 | 45 | c | 9 | BR-2017 | 0.1445 | 33 | cdef |
| 10 | 9 bar | 0.1393 | 45 | cd | 10 | S-5765 | 0.1436 | 33 | cdef |
| 11 | 10 bar | 0.1172 | 45 | d | 11 | S-5797 | 0.1416 | 33 | cdef |
| TRT, Treatments (salt concentrations or varieties; NSR, NS ranges | | | | | 12 | BR-2/1 | 0.1367 | 33 | def |
| | | | | | 13 | S-5798 | 0.1301 | 33 | ef |
| | | | | | 14 | S-5785 | 0.1257 | 33 | ef |
| | | | | | 15 | BR-99 | 0.1199 | 33 | f |

The variation of germination and seedling growth as observed here with germplasms, nature of salt and their concentrations are also reported to occur with genotypic nature of species and varieties and salt and its concentrations by Asana and Kale (1965) in wheat, Maliwal and Paliwal (1970) in Bajra, Kumar and Bhardwaj (1981) in mung and Bafeel (2014) in Sorghum.

Table15. Seedling dry weight (mg) of guar cultivars and lines over the treatments range (0 to 10 bar).

| Rank (NaCl Based) | Genotype | Seedling Dry Weight (mg) over treatments range (0-10 bar) – mean \pm SE, % CV) | | Rank (Na ₂ SO ₄ based) |
|-------------------------|----------|-------------------------------------------------------------------------------------|---------------------------------|----------------------------------------------------|
| | | NaCl | Na ₂ SO ₄ | |
| 10 | BR-2/1 | 153.09 \pm 6.26, 13.55 | 136.82 \pm 11.93, 12.01 | 12 |
| 13 | BR-90 | 137.45 \pm 8.78, 21.19 | 143.55 \pm 11.93, 27.55 | 10 |
| 11 | BR-99 | 148.55 \pm 6.28, 14.01 | 119.91 \pm 10.39, 28.74 | 15 |
| 7 | Br-2017 | 165.00 \pm 13.01, 26.15 | 145.64 \pm 14.75, 33.60 | 9 |
| 5 | S-5733 | 189.52 \pm 28.25, 40.68 | 193.27 \pm 19.98, 34.29 | 1 |
| 6 | S-5742 | 188.45 \pm 6.74, 11.86 | 192.45 \pm 11.63, 20.04 | 2 |
| 4 | S-5747 | 190.45 \pm 3.84, 6.68 | 187.27 \pm 7.11, 12.59 | 3 |
| 3 | S-5759 | 197.45 \pm 6.86, 11.53 | 174.18 \pm 12.68, 24.15 | 6 |
| 15 | S-5761 | 121.64 \pm 14.61, 39.83 | 175.73 \pm 20.49, 53.77 | 4 |
| 2 | S-5765 | 209.18 \pm 22.02, 34.91 | 165.55 \pm 5.04, 10.10 | 7 |
| 1 | S-5784 | 211.18 \pm 19.92, 21.86 | 175.27 \pm 18.20, 34.47 | 5 |
| 14 | S-5785 | 129.00 \pm 8.16, 20.98 | 125.73 \pm 9.22, 24.32 | 14 |
| 12 | S-5797 | 148.36 \pm 13.97, 31.23 | 141.64 \pm 10.04, 23.51 | 11 |
| 8 | S-5798 | 157.82 \pm 16.52, 4.70 | 130.09 \pm 13.06, 33.06 | 13 |
| 9 | S-5825 | 156.18 \pm 6.44, 13.68 | 157.73 \pm 20.28, 42.64 | 8 |

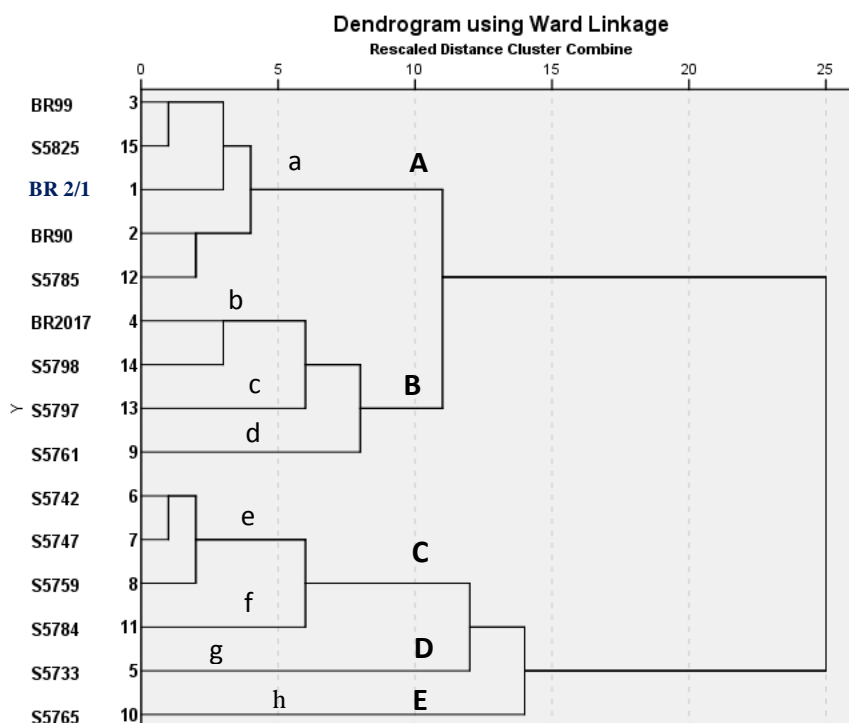


Fig. 14. Agglomerative clustering of guar germplasms on the basis of the seedling mass under NaCl salinity.

Our results on seedling growth are quite different than that reported for 25 guar germplasms (CG-1, HGS-25, HGS-18, CG-4, PLG-174, HFG-314, DSE-II, HGS -46, PLG-85, PLG-119, HGS-47, DSE - 16J, HFG-156, PNB, FS-277, HGS-43, HFG-119, Guara, HG-182, HGS-75, HFG-189, Suvidha, HGS-3, HG-17-1 and Hg-258) under mixed salt salinity by Datta and Dayal (1988). Under increasing mixed salinity while root elongation consistently increased, the shoot length decreased with salinity. Cotyledonary fresh weight was reported to increase markedly by 12 and 16dS.m⁻¹. Dry weights of shoot and root did not differ appreciably. Of cultivar studied, PLG-119 and HGS-18 were found relatively salt tolerant. Decline in root and shoot growth with salinity is, however, manifest in several studies (Cramer *et al.*, 1994, Mansour *et al.*, 2005, Chowdhury *et al.*, 2018). The presence of salt is reported to reduce vegetative and reproductive growth parameters of guar by Kumar *et al.* (1988). Garg *et al.* (1997) have

also reported reduction in dry matter production of guar at seedling stage under salinity. Kumar and Bhardwaj (1981) have reported genotypic differences in *Vigna radiata* seedling growth under salinity. Such reduction of growth may primarily be related to the limited supply of metabolic energy for the maintenance of normal growth processes (O'Leary, 1986). The amount of work to combat ionic and osmotic effects is increased and in consequence relatively less energy is available for growth (Nieman and Maas, 1978). The growth of even halophyte *Salicornia* is reported to be adversely affected by salinity (Hoffman and Sachert, 1967).

Relationship of cumulative seedling dry weight with NaCl and Na₂SO₄ salinities

NaCl salinity:

Table 16 portrays the relationship in terms of simple linear correlation coefficients of seedling dry weights of various germplasms to NaCl and Na₂SO₄ salinities (0 to 10 bars). Under NaCl, all germplasms exhibited non-significant relationships except S-5733 which exhibited significant positive correlation ($r = 0.755$, $p < 0.007$) and S-5798 which exhibited significant negative correlation (-0.828 , $p < 0.002$).

S-5733 – Seedling weight (mg) = $102.091 + 17.545 \text{ NaCl in bar} \pm 53.31$

$t = 3.40 \quad t = 3.45$

$p < 0.008 \quad p < 0.007$, $r = + 0.755$, $F = 11.92$ ($p < 0.007$)

S-5798 – Seedling weight (mg) = $226.230 - 13.682 \text{ NaCl in bar} \pm 32.34$

$t = 12.40 \quad t = - 4.44$

$p < 0.0001 \quad p < 0.002$, $r = - 0.828$, $F = 19.69$ ($p < 0.002$)

Table 16. Linear correlation Coefficient “r” between Seedling dry mass (mg) and NaCl and Na₂SO₄ salinities.

| Germplasms | NaCl | | Na ₂ SO ₄ | |
|------------|----------------|------------------|---------------------------------|------------------|
| | “r” | Significance, p | “r” | Significance, p |
| BR- 2/1 | 0.333 | 0.318, NS | 0.079 | 0.818, NS |
| BR-90 | 0.218 | 0.519, NS | 0.186 | 0.584, NS |
| BR-99 | 0.021 | 0.953, NS | 0.234 | 0.489, NS |
| BR-2017 | 0.491 | 0.126, NS | 0.312 | 0.351, NS |
| S-5733 | 0.755 | 0.007 *** | 0.851 | 0.001 *** |
| S-5742 | 0.274 | 0.415, NS | 0.281 | 0.402, NS |
| S-5747 | 0.275 | 0.414, NS | -0.692 | 0.018 ** |
| S-5759 | 0.248 | 0.463, NS | -0.761 | 0.007 *** |
| S-5761 | 0.368 | 0.265, NS | -0.345 | 0.298, NS |
| S-5765 | 0.498 | 0.119, NS | -0.139 | 0.684, NS |
| S-5784 | 0.204 | 0.548, NS | -0.403 | 0.218, NS |
| S-5785 | 0.203 | 0.550, NS | -0.274 | 0.415, NS |
| S- 5797 | 0.054 | 0.805, NS | -0.366 | 0.269, NS |
| S-5798 | - 0.828 | 0.002 *** | -0.869 | 0.001 *** |
| S-5825 | 0.319 | 0.339, NS | 0.331 | 0.320, NS |

Table 17. Promotion / reduction of cumulative seedling dry weight in -10 bar NaCl and Na₂SO₄ over control.

| Germplasms | Promotion / Reduction (%) | |
|------------|---------------------------|---------------------------------|
| | NaCl | Na ₂ SO ₄ |
| BR- 2/1 | +14.38 | -7.29 |
| BR-90 | -23.08 | -5.98 |
| BR-99 | +21.95 | -21.19 |
| BR-2017 | -62.15 | -49.72 |
| S-5733 | +163.33 | +192.22 |
| S-5742 | +21.74 | -22.46 |
| S-5747 | -9.18 | -30.61 |
| S-5759 | -8.24 | -30.77 |
| S-5761 | -57.30 | -20.22 |
| S-5765 | -17.68 | -12.71 |
| S-5784 | -39.39 | -61.04 |
| S-5785 | +51.16 | +12.79 |
| S- 5797 | -18.60 | -17.83 |
| S-5798 | -70.19 | -74.04 |
| S-5825 | +28.57 | +6.72 |

S-5742, S-5761 and S-5825 exhibited curvilinear relationships of cumulative seedling mass with NaCl salinity.

S-5742 - Seedling wt. (mg) = $148.336 + 22.44 \text{ bar} - 2.059 \text{ bar}^2 \pm 11.07$

$t = 17.58 \quad t = 5.72 \quad t = -5.05$

$P < 0.0001 \quad p < 0.0001 \quad p < 0.0001$; $R = 0.896$, $F = 16.36$ ($p < 0.0001$)

S-5761 - Seedling wt. (mg) = $99.210 + 27.509 \text{ bar} - 3.289 \text{ bar}^2 \pm 37.01$

$t = 3.51 \quad t = 2.09 \quad t = -2.59$

$p < 0.008 \quad p < 0.032 \quad p < 0.070$; $R = 0.729$, $F = 4.53$ ($p < 0.070$)

S-5825 - Seedling wt. (mg) = $121.119 + 18.581 \text{ bar} - 1.653 \text{ bar}^2 \pm 14.81$

$t = 10.74 \quad t = 3.54 \quad t = -3.27$

$p < 0.0001 \quad p < 0.008 \quad p < 0.011$; $R = 0.785$, $F = 6.404$ ($p < 0.022$)

Na₂SO₄ salinity:

Under Na₂SO₄ salinity, seedling weight of only four germplasms exhibited significant linear correlation with salinity, of which one (S-5733) showed positive correlation and three (S-5747, S-5759 and S-5798) showed negatively correlation. Br-90 and Br-5742 showed curvilinear relationship between seedling weight and sodium sulphate salinity. The best fit equation for these germplasms are as follows:

Linear Equations:

S-5733 - Seedling wt. (mg) = $108.273 + 17.00 \text{ bar} \pm 36.70$

$t = 5.23 \quad t = 4.86$

$p < 0.001 \quad p < 0.001$, $r = +0.851$, $F = 23.60$ ($p < 0.001$)

S-5747 - Seedling wt. (mg) = $211.864 - 4.9187 \text{ bar} \pm 17.936$

$t = 20.88 \quad t = -2.88$

$p < 0.0001 \quad p < 0.018$, $r = -0.692$, $F = 8.27$ ($p < 0.018$)

S-5759 - Seedling wt. (mg) = $222.409 - 9.645 \text{ bar} \pm 28.378$

$t = 13.38 \quad t = -3.51$

$p < 0.0001 \quad P < 0.007$, $r = -0.761$, $F = 12.36$ (0.007)

S-5798 - Seedling wt. (mg) = $186.864 - 11.355 \text{ bar} \pm 22.586$

$t = 14.67 \quad t = -5.27$

$p < 0.0001 \quad p < 0.001$, $r = -0.869$, $F = 27.80$ ($p < 0.001$)

Curvilinear Equations:

BR-90 - Seedling wt. = $86.645 + 32.824 \text{ bar} - 3.06 \text{ bar}^2 \pm 31.41$

$t = 3.82 \quad t = 3.12 \quad t = -3.02$

$p < 0.005 \quad p < 0.014 \quad p < 0.017$. $R = 0.740$, $F = 4.86$ ($p < 0.042$)

S-5742 - Seedling wt. (mg) = $156.41 + 31.609 \text{ bar} - 3.494 \text{ bar}^2 \pm 20.08$

$t = 10.23 \quad t = 4.45 \quad t = -5.10$

$p < 0.0001 \quad p < 0.002 \quad p < 0.001$, $R = 0.855$, $F = 14.45$ ($p < 0.002$)

Promotion / Reduction in seedling wt. In 10 bar salinities of NaCl and Na₂SO₄ over the respective controls

NaCl salinity:

Following sequence of germplasms describe their salt tolerance in terms of the promotion or reduction in seedling dry wt. In 10 bar NaCl and Na₂SO₄ salinities over the respective control. In NaCl, six germplasms showed promotion and three germplasms showed quite higher reduction in seedling mass.

S-5733 (+163.33%) > S-5785 (+51.16%) > S-5825 (+28.57%) > BR-99 (+21.95%) ≈ S-5742 (+21.74%) > BR-2/1 (+14.38%) > S-5759 (-8.24%) > S-5747 (-9.18%) > S-5765 (-17.68%) > S-5797 (-18.60%) > BR-90 (-23.08%) > S-5784 (-39.39%) > S-5761 (-57.30%) > BR-2017 (-62.15%) > S-5798 (-70.19%)

Na₂SO₄ salinity:

Under Na₂SO₄ salinity, three germplasms (S-5733, S-5785 and S-5825) showed promotion in seedling weight in 10 bar over control and three germplasms (BR-2017, S-5784 and S-5798) exhibited reduction near 50% or above 50% as given below. Three germplasms (S-5761, BR-99 and S-5742) were almost equally tolerant with reduction in seedling wt. Fluctuating around 20 – 22%.

S-5733 (+192.22%) > S-5785 (+12.79%) > S-5825 (+6.72%) > BR-90 (-5.98%) > BR-2/1 (-7.29%) > S-5765 (-12.71%) > S-5797 (-17.83%) > S-5761 (-20.22%) ≈ BR-99 (-21.19%) ≈ S-5742 (-22.46%) > S-5747 (-30.61%) ≈ S-5759 (-30.77%) > BR-2017 (-49.72%) > S-5784 (-61.04%) > S-5798 (-74.04%).

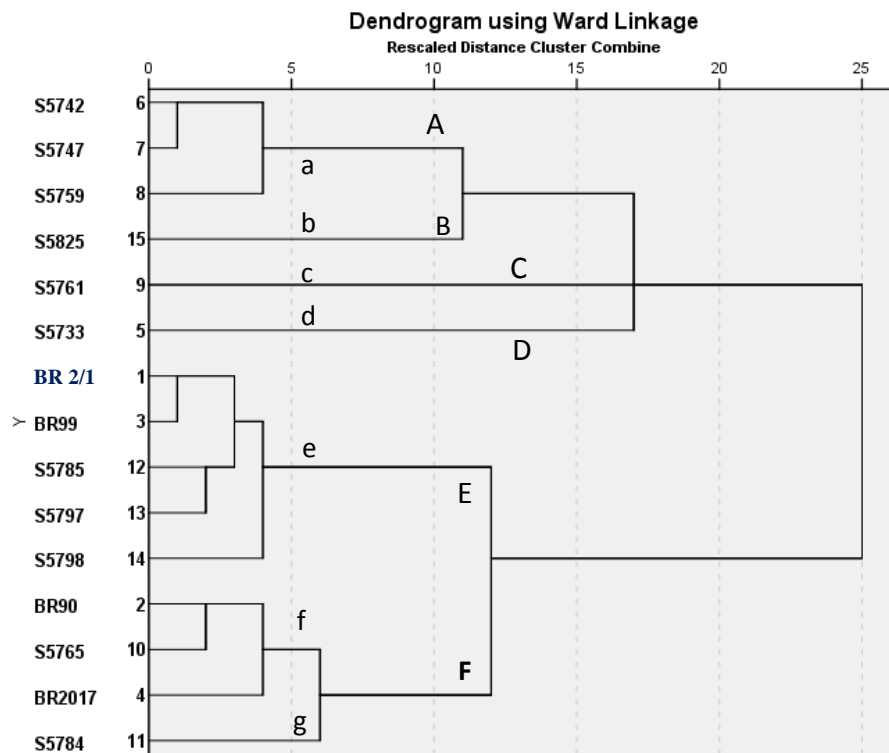


Fig.15. Agglomerative clustering of guar germplasms based on seedling dry weight under Na₂SO₄ salinization.

Germplasms agglomeration on the basis of cumulative seedling dry mass under NaCl

Fig. 14 presents the cluster diagram of germplasms based on seedling mass under NaCl. On the basis of 5% distance amongst the germplasm eight discrete groups were observed and on the basis of 10% distance five groups were discretely seen in the figure. They are described below for their composition and their seedling dry mass. Biomass production was much higher in group-f, S-5784 (211.18 mg) followed by group h, S-5765 (209.18mg).

Dissimilarity Distance: 5%

Group a: BR-99, S-5825, BR-2/1, BR-90, and S-5785

Mean seedling mass over the treatment range: $150.85 \pm 6.292\text{mg}$, CV: 9.32% -- Rank VI

Group b: BR-2017 and S-5798

Mean seedling mass over the treatment range: $161.41 \pm 3.590\text{mg}$, CV: 3.15% --Rank V

Group c: S-5797

Mean seedling mass over the treatment range: 148.36mg – Rank VII

Group d: S-5761

Mean seedling mass over the treatment range 121.46mg – Rank VIII

Group e: S-5742, A-5747 and S-5759

Mean seedling mass over the treatment range: $192.11 \pm 2.74\text{mg}$, CV: 2.47% -- Rank III

Group f: S-5784

Mean seedling mass over the treatment range: **211.18mg – Rank I**

Group g: S-5733

Mean seedling mass over the treatment range: 189.52mg – Rank IV

Group h: S-5765

Mean seedling mass over the treatment range: **209.18mg – Rank II**

BR-99 occupied group-a (BR-99, S-5825, BR- 2/1, BR- 90 and S-5785) was relatively low biomass producer. Group-f (S-5784) was the highest biomass accumulator followed by group h (S-5765). S- 5761 represented the lowest seedling biomass accumulation.

Dissimilarity Distance: 10%

Group A: BR-99, S-5825, BR-2/1, BR-90, and S-5785

Mean seedling mass over the treatment range: $150.85 \pm 6.292\text{mg}$, CV: 9.32% --Rank IV

Group B: BR-2017, S-5798, S-5797 and S-5761

Mean seedling mass over the treatment range: $143.28 \pm 7.75\text{ mg}$, CV: 10.82% --Rank V

Group C: S-5742, S-5747, S-5759 and S-5784

Mean seedling mass over the treatment range: $196.88 \pm 5.145\text{ mg}$, CV: 5.27% --Rank III

Group D: S-5733

Mean seedling mass over the treatment range: **198.52mg – Rank II**

Group E: S-5765

Mean seedling mass over the treatment range: **209.18mg – Rank I**

Germplasms agglomeration on the basis of mean seedling dry mass under Na_2SO_4

Fig. 15 presents the cluster diagram of germplasms based on seedling mass under Na_2SO_4 . On the basis of 5% distance amongst the germplasm seven discrete groups were observed and on the basis of 10% distance six groups were discretely observed. These groups are described below for their composition and their mean seedling dry mass.

Dissimilarity Distance: 5%

Group a: S-5742, S-5747 and S-5759

Mean seedling mass over the treatment range: **$184.63 \pm 5.44\text{ mg}$, CV: 5.10% -- Rank II**

Group b: S-5825

Mean seedling mass over the treatment range: 156.18mg – Rank IV

Group c: S-5761

Mean seedling mass over the treatment range: 121.64 mg – Rank VII

Group d: S-5733

Mean seedling mass over the treatment range: **189.52 mg –Rank I**

Group e: BR-2/1, BR-99, S-5785, S-5797 and S-5798

Mean seedling mass over the treatment range: $130.84 \pm 0.0663\text{ mg}$, CV: 6.60% -- Rank VI

Group f: BR-90, S-5765 and BR-2017

Mean seedling mass over the treatment range: $151.58 \pm 7.011\text{ mg}$, CV: 8.01% -- Rank V

Group g: S-5784

Mean seedling mass over the treatment range: 175.28 mg – Rank III

This cluster diagram was predominated by group-d (S-5733) (189.52mg) followed by group-a (S-5742, S-5747 and S-5759) ($184.63 \pm 5.44\text{mg}$)

Dissimilarity Distance: 10%

Group A: S-5742, S-5747 and S-5759

Mean seedling mass over the treatment range: **$184.63 \pm 5.436\text{ mg}$, CV: 5.10% -- Rank II**

Group B: S-5825

Mean seedling mass over the treatment range: 156.18mg – Rank IV

Group C: S-5761

Mean seedling mass over the treatment range: 121.64 mg – Rank IV

Group D: S-5733

Mean seedling mass over the treatment range: **189.52 mg – Rank I**

Group E: BR-2/1, BR-99, S-5785, S-5797 and S-5798

Mean seedling mass over the treatment range: $130.84 \pm 0.0663\text{mg}$, CV: 6.60% -- Rank V

Group F: BR-90, S-5765, BR-2017, S-5784

Mean seedling mass over the treatment range: $157.50 \pm 7.724\text{ mg}$, CV: 9.81% -- Rank III

Group-D (S-5733) (189.52mg) and group-A (S-5742, S-5747 and S-5759) ($184.63 \pm 5.44\text{mg}$) predominated the cluster diagram based on seedling biomass production.

Threshold salinities in relation to germination and Germination velocity Index

From the scatter diagrams presented in Fig. 7 and 8, threshold salinities of NaCl and Na_2SO_4 were approximated. The threshold salinity was defined here as the maximum salinity beyond which visible decline, greater or smaller, began to set in germination (and not the salinity corresponding with 50% reduction in a germination parameter). Such threshold salinities for various germplasms are presented in Table 17. In this respect BR-2017 was the most sensitive variety with threshold NaCl salinity c. 3 to 4 bar. In S-5797, threshold NaCl salinity was 4 to 5 bar and in S-5798 and S-5825 it was 5 bar NaCl. In six germplasms (S-5733, S-5742, S-5747, S-5784, S-5785 and S-5759, threshold salinity was c. 7 bar NaCl. The threshold salinity was quite high (9 bar) in S-5761 and BR-90. BR- 2/1, and BR-99 had Threshold salinity much above 10 bar NaCl. There is a generalization that growth reduction in many plants occurs at osmotic potentials of 2 to 4 bar NaCl (Wilcox, 1959).

In comparison, threshold salinity in Na₂SO₄ was quite low i.e., 3 bar in S-5797, S-5759 and Br-2017. In some other germplasms it was relatively higher, 5 bar in S-5798, S-5784, S-5761, S-5742, and S-5733 and 6 bar in BR-2/1, BR-90, BR-99 and S-5747. Two germplasms, S-5765 and S-5825 had maximum threshold salinity of 7 bar Na₂SO₄. It was obvious that performance of guar germplasms was comparatively somewhat lower in sulphate salinity than that in NaCl.

Table 17. Threshold salinities* for guar germplasms on the basis of germination and GVI under NaCl and Na₂SO₄ salinities. Threshold salinity of germination based on data behaviour in the scatter diagrams for the linear correlation and regression of final germination (for day 4) with salinity (see Fig.7 and 8), using replicate data (N = 33). Threshold salinity for GVI was approximated on the basis of data given in Table 5a and b.

| Genotype | GERMINATION | | GVI | |
|----------|---------------------------|---------------------------------|----------------------------------------------------------------------------------------|---------------------------------|
| | NaCl | Na ₂ SO ₄ | NaCl | Na ₂ SO ₄ |
| | Threshold salinity in bar | | | |
| BR-2/1 | >10 bar | 6 bar | 3 bar | 1 bar |
| BR-90 | c. 9 bar | 6 bar | 4 to 5 bar | 2 bar |
| BR-99 | > 10 bar | 6 bar | **Promotion = 6.25%; threshold above -10 bar i.e. EC > 22.2 dS.m ⁻¹ . | 1 bar |
| Br-2017 | 3 to 4 bar | 5 bar | 2 bar | 1 bar |
| S-5733 | 7 bar | 5 bar | 2 to 3 bar | 2 to 3 bar |
| S-5742 | 7 bar | 5 bar | 2 bar | 2 bar |
| S-5747 | 7 bar | 5 to 6 bar | 1 bar | 4 bar |
| S-5759 | 7 bar | 3 bar | 2 bar | 2 to 3 bar |
| S-5761 | 9 bar | 5 bar | 1 bar | 1 bar |
| S-5765 | 5 to 6 bar | 7 bar | 4 bar | 6 bar |
| S-5784 | 7 bar | 5 to 6 bar | 1 bar | 1 bar |
| S-5785 | 7 bar | 6 to 7 bar | 2 bar | 4 to 5 bar |
| S-5797 | 4 to 5 bar | 3 bar | 3 bar | 2 bar |
| S-5798 | 5 bar | 5 bar | 5 to bar | 1 bar |
| S-5825 | 5 bar | 7 bar | 2 bar | 1 bar |

*, Threshold salinity referred to the salinity above which decline in germination began. .
**, Promotion of GVI was recorded in even 10 bar NaCl-treatment over control in this germplasm (BR – 99).

Table 18. Approximated salinities (NaCl and Na₂SO₄ in bar associated with 50% reduction in final germination based on scatter of the replicated final germination data plotted against salinities (0 to 10 bar). See Figure 7 and 8).

| Germplasms | Approximated 50 % Reduction | |
|------------|---------------------------------|---------------------------------------|
| | NaCl (bar) | Na ₂ SO ₄ (bar) |
| BR-2/1 | Above 10 bar | 10 bar |
| BR-90 | Above 10 bar | Above 10 bar |
| BR-99 | Above 10 bar | c. 9 bar |
| BR-2017 | Variable, Above 10 bar | c. 5 bar, High behavioural dispersion |
| S-5733 | Above 10 bar | 9 bar, behavioural dispersion |
| S-5742 | Above 10 bar | 9 bar |
| S-5747 | Above 10 bar | 10 bar |
| S-5759 | 10 bar (Behavioural dispersion) | 8 bar |
| S-5761 | 10 bar | 8 to 9 bar |
| S-5765 | 8 to 9 bar | 8 bar |
| S-5784 | 9 bar | 8 bar, behavioural dispersion |
| S-5785 | 9 to 10 bar | 6 bar, behavioural dispersion |
| S-5797 | Above 10 bar | 7 to 8 bar |
| S-5798 | 10 bar | 9 to 10 bar |
| S-5825 | 10 bar | 10 bar |

It is obvious from Table 18 that in hand germplasms, on the basis of approximate levels of salinity causing 50 % reduction in final germination were comparatively more tolerant to NaCl than Na₂SO₄ salinity. All germplasms

exhibited moderate to high level of salt tolerance as regard to the final germination achieved after four days of incubation. Germination velocity is impeded, however quite early even in much lower salinities. The germplasms somehow cope up with salinity and restore germination in appreciable proportion reaching the fourth day of incubation.

Table 19. Values of Composite Performance Index (CPI) of various guar germplasms for NaCl and Na₂SO₄.

| Germplasms | Composite Performance Index | |
|------------|-----------------------------|---------------------------------|
| | NaCl | Na ₂ SO ₄ |
| BR-2/1 | 89.30 | 79.41 |
| BR-90 | 84.91 | 84.21 |
| BR-99 | 90.29 | 72.15 |
| BR-2017 | 84.20 | 78.34 |
| S-5733 | 100.80 | 95.44 |
| S-5742 | 98.71 | 96.97 |
| S-5747 | 98.29 | 96.38 |
| S-5759 | 100.56 | 87.52 |
| S-5761 | 75.00 | 87.79 |
| S-5765 | 103.03 | 87.08 |
| S-5784 | 100.70 | 88.21 |
| S-5785 | 76.12 | 71.00 |
| S-5797 | 80.06 | 75.87 |
| S-5798 | 85.29 | 74.18 |
| S-5825 | 83.50 | 85.66 |

CPI

Composite Performance Index = CPI = (Mean seedling dry mass (mg) after four days of incubation + Mean GVI over the treatment range (0 to 10 bar) + Mean final germination over the treatment range (0 to 10 bar)) /3.

Threshold salinities in case of GVI were approximated on the basis of data given in Table 5a and 5b. The decline in GVI was comparatively set in NaCl salinities (1 to 2 (- 3) in most of the germplasms except S-5798 (5 to 6 bar NaCl). BR-99 was, however, the only germplasms showing promotion in GVI over control even under 10 bar NaCl and threshold salinity was somewhere above 10 bar (Table 17). GVI began to decline in magnitude in lower salinities of 1 or 2 bar Na₂SO₄ except S-5733 (2 to 3 bar), S-5747 (4bar), S-5785 (4 to 5 bar) and S-5765 (6 bar). Francois and Kleiman (1990) have reported threshold ECe of guar to be 8.8 dS.m⁻¹ (around 3 bar). Inhibition of the seeds of all varieties tested (S1538, S2378, S 2395, S-2381, S-2376, S-196, S-287 and S-212) by Yadava *et al.* (1974), germinated up to 2.0 EC but only three varieties S 23765, S-196 and S- 287) showed 100 % germination at 4.0 EC. In 11 EC, germination ranged from 7.5 to 26.62% and more than 50% germination was observed by 6.0 EC. The germination in *Rhynchosia minima* was reported only up till 8 bar NaCl (Shaukat and Burhan, 2000).

Performance of germplasms on the basis of a composite index of performance

Since the performance of various germplasms varied with respect to germination, germination velocity and seedling growth, Composite Performance Index (CPI) was calculated as described above with this contention that higher is the CPI, higher the salt tolerance of a germplasm. On the basis of CPI (Table 19), there were three groups of germplasms evident in NaCl.

NaCl salinity:

1. Germplasms with CPI: 70-80 (S-5761, S-5785 and S-5797)
2. Germplasms with CPI: 81-90 (S-5825, BR-2017, BR-90, S-5798, BR- 2/1, and BR-99)
3. Germplasms with CPI: 98-103 (S-5747, S- 5742, S-5784, S- 5759, S-5733 and S-5765)

All germplasms were definitely but differentially tolerant to NaCl salinity. Group I was comparatively lesser tolerant, group II moderately tolerant and group III comparatively highly tolerant to salinity.

There were three groups of germplasms discernible in Na₂SO₄ salinity as follows (Table 18) as follows:

Na₂SO₄ salinity:

- I. Germplasms with CPI: 71-79 (S-5785, BR-99, S-5798, S-5797, BR-2017, BR- 2/1). All varietal germplasms except BR-90 entered the composition of this group.
- II. Germplasms with CPI: 84-88 (BR-90, S-5825, S-5765, S-5761, S-5784)
- III. Germplasms with CPI: 95-96 (S-5733, S-5747, S-5742)

Obviously, group III was relatively more tolerant.

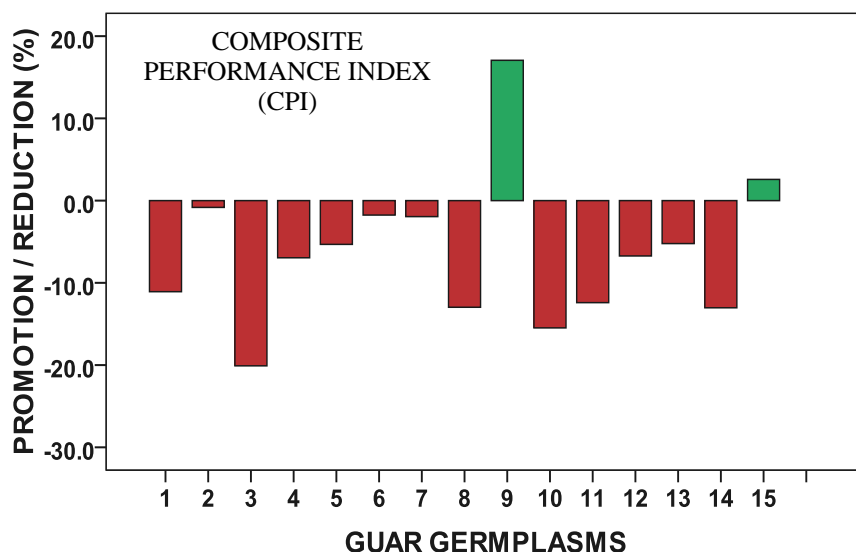


Fig. 16. *Per cent* promotion or reduction in composite performance index (CPI) under Na₂SO₄ salinity relative to the NaCl. **Germplasms acronyms:** 1, BR-2/1; 2, BR-90; 3, BR-99; 4, BR-2017; 5, S-5733; 6, S-5742; 7, S-5747; 8, S-5759; 9, S-5761; 10, S-5765; 11, S-5784; 12, S-5785; 13, S-5797; 14, S-5798; and 15, S-5825. Promotion / Reduction in CPI = $(CPI_{Na_2SO_4} - CPI_{NaCl} / CPI_{NaCl}) * 100$.

Comparison of germplasms performance in NaCl and Na₂SO₄ on the basis of CPI

Per cent promotion or reduction of CPI in Na₂SO₄ relative to NaCl is presented in Fig. 16. The germplasms under study tended to form five groups as regards to the CPI.

CPI reduction / promotion in Na₂SO₄ over NaCl:

The data of promotion/ reduction in the CPI behaviour of the germplasms in Na₂SO₄ relative to NaCl on the basis of three germination related parameters (Final mean germination over the treatment range, mean velocity of germination over the treatment range and the seedling dry mass at the last day of incubation in the treatment range (0 to 10 bar) is presented in Fig. 16. Following generalization may be made from this data.

- Germplasms showing CPI reduction: - 0.82 to -1.95% (BR- 90, S-5742 and S-5747). These germplasms were more or less equally tolerant to NaCl and Na₂SO₄ salinities.
- Germplasms with CPI reduction: -5.23 to - 6.95% (S-5797, S-5733, S-5785, and BR-2017)
- Germplasms with CPI reduction: -11.08 to - 15.48% (BR- 2/1, S-5784, S-5759, S- 5798 and S-5765) – more sensitive to Na₂SO₄ than above groups.
- Germplasms with CPI reduction: -20.99% (BR-99) – most sensitive to Na₂SO₄ amongst the tested germplasms.
- Germplasms with CPI promotion: + 2.58 to +17.06% (S-5825 and S-5761, respectively) – Quite tolerant to Na₂SO₄ salinity which was promotory to them over NaCl- particularly S-5761.

CPI reduction / promotion in NaCl over Na₂SO₄:

The results were obtained on the basis of CPI promotion / reduction in NaCl over Na₂SO₄. Following generalizations could be made:

- Germplasms showing CPI promotion: 0.83 to 1.94% (BR-90, S-5742 and S-5747) – More or less equally tolerant to NaCl and Na₂SO₄.
- Germplasms with CPI promotion: 4.9 to 7.48% (BR- 2/1, S-5747, S-5733, S- 5785 and BR-2017).
- Germplasms with CPI promotion: 14.17-18.32% (S-5784, S-5759, S-5798 and S-5765) - moderate promotion.
- Germplasm with CPI promotion: 25.14% (BR-99) – Maximum promotion in NaCl.
- Germplasms with CPI reduction: -2.52 to -14.31% (S-5825 and S-5761) – Less tolerant to NaCl than to Na₂SO₄ – particularly S-5761.

It follows from this data that BR-99 is although tolerant to NaCl, it is quite sensitive to Na₂SO₄ salinity at germination stage. AlShameri *et al.* (2017) have reported this germplasm to be less tolerant to drought than several

other accessions from Pakistan viz. BWP-5595, 24320, Chiniot 1, Chiniot 2, Kaloorkot 2 and BWP 5599. The accessions 24320, BWP 5595, Chiniot 1, Chiniot 2 and 22159 were also rated as salt tolerant by them. Accession BWP 5595 is reported to perform better than accession 24320 when subject to multiple stresses of heat, drought and salinity combined (AlShameri *et al.*, 2019).

It is apparent from the results given above regarding CPI reduction or promotion that germplasms were differentially tolerant to NaCl salinity and in any of the germplasms the reduction was not more than 21% in Na₂SO₄ salinity. Besides the varietal germplasms, especially BR-99 which is known for its salt tolerance (Rasheed *et al.*, 2015), some lines such as S-5747, S- 5742, S-5784, S- 5759, S-5733 and S-5765 are also of the interest particularly on the basis of the magnitude of their composite performance index. The germplasms in hand were also tolerant to Na₂SO₄ salinity but at somewhat reduced level. The germplasms, S-5733, S-5747, S-5742, were however, quite tolerant to Na₂SO₄. The germplasms, especially S-5761 and to some extent S-5825 showed promotion in their composite performance in Na₂SO₄ salinity than that in NaCl salinity. Germplasm S-5733, which is rated as an advance line and is currently under trial at Bahawalpur Agricultural Research Station, deserves special attention of researchers as regards to its salinity tolerance under field conditions. Several local guar germplasms that are also reported to be salt tolerant may be listed such as BR-99, 5597, 41671, Khushab white, Mardan white, Shianwali white (Rasheed *et al.*, 2015), accessions 281/ 3 from Balochistan (Ashraf *et al.* (2005), and Lines S- 5881, S 5932, S-6067 (Sudhar *et al.*, 2018b) are salt tolerant guar germplasms from Pakistan. Cultivar Kinman is a salt tolerant exotic germplasms (Andrade, 1985). Guar is reported to cope with salinity by accumulation of sugar and proline as osmoticum adjustment. Decline in protein and starch, however, takes place (Gulati and Dhingra (2014). The genotypes RGC-1002 and RGC-197 are reported by Soni *et al.* (2017) to respond to salt stress (*in vitro*) by modifying various biochemical pathways to prevent salt-induced oxidative damage i.e., by rapid accumulation of protective anti-oxidative enzymes (peroxidase and catalase) There is a need to investigate the physiological mechanism of salt tolerance of Pakistan germplasms of guar.

As regards to the sulphate sensitivity of some guar germplasms observed in the present studies as regards to their germination velocity or seedling growth, it may be mentioned that several plants have been reported to be more sensitive to SO₄²⁻ ion than Cl⁻ ion. Up to 4 bar NaCl (c. 9.77dS.m⁻¹); there appeared almost no effect on germination of *Achyranthes aspera* seeds beyond which both germination and rate of germination regularly declined. Na₂SO₄ inhibited its germination more drastically (Khan *et al.*, 1984). *Peristrophe bicalyculata* (now *P. paniculata*) and *Cassia holosericea* were more sensitive – their germination was greatly reduced at 3 bar concentration of NaCl and Na₂SO₄. In case of *Prosopis juliflora*, no effect on germination was recorded up to 8 bar NaCl. However, rate of germination was much impeded in Na₂SO₄. The shoot growth declined in all these species with salinity but more under the influence of Na₂SO₄ (Khan *et al.*, 1984). Strogonov (1964) reported white lupin, wheat, sorghum and cotton to be more sensitive to sulphate salinity.

Germination assays for salt tolerance do not predict plant growth and yield in saline soils (Teolis *et al.* (2009). It may be emphasized that salt tolerance at germination and other phases of life cycle of a plant are not necessarily correlated (Ayers, 1952; Khan, 1987; Khan and Ahmad, 1998). The results reported in literature are, however, controversial. Mayer and Poljakoff-Meyber (1975) observed that adult plants are more salt tolerant than the seeds. Ungar (1974) reported that seed germination in *Hordeum jubatum* was more resistant process than the later growth of seedlings. Azizov (1974) reported that seeds of *Limonium meyeri* cannot germinate in salinity above 1.5 % salt, yet the mature plant can grow even in the presence of 10% salt solution. The Ec_{iw} corresponding to 50% reduction in seed germination (10.03 dS.m⁻¹) in *Indigofera oblongifolia* was more or less comparable to that inducing 50% reduction in growth (12.05 dS.m⁻¹) (Khan and Ahmad, 1998). It was stressed by Angevine and Chabot (1979) that the germination of seeds and the subsequent early growth of seedlings represent the period of maximum vulnerability to changes in the environment and minimal potential for homeostatic response. Salinity studies should include, therefore, both germination and plant trials in field.

Three varieties of guar (PI 217923, PI 340246 and ARS Lewis) were studied by Liu (2003). Seed coat colour in guar ranges from black to dull white. The seeds of above accessions were sorted into light, dark gray and black and their germination was tested at 22 °C in dark. During germination seeds of black and dark gray colour had higher water absorption after 24 h and had higher germination rate than light coloured seeds of any accessions. After 72 h, black or dark gray seeds had higher germination percentage. Research with local germplasms is needed on this aspect also.

Seedling mortality

Seedling mortality in NaCl salinity

In our experimentation, no mortality except one seedling was recorded in NaCl salinity.

Seedling mortality in Na₂SO₄ salinity

There was substantial degree of seedling mortality in Na₂SO₄ salinity (Table 20). The germplasms tested behaved differentially in this respect. There was no mortality up to 3 bar Na₂SO₄ in any germplasms. Seedling mortality due to decay and burning began to take place from 4 bar (EC: 11.1 dS.m⁻¹) onwards in S-5742, S-5747, S-5759, S-5797 and S-5798 more or less regularly along the increase in salt level. Seedling mortality began from 5 bar (EC: 13.33dS.m⁻¹) in S-5733 and S-5785 and from 6 bar (EC: 15.9 dS.m⁻¹) in S-5765, S-5825 and BR-2017. The varietal germplasms, BR-2/1, BR-90 and BR-99 and the accessions S-5761 and S-5784 were relatively more tolerant and exhibited seedling mortality from 7 bar Na₂SO₄ (EC: 16.87 dS.m⁻¹) onwards. Obviously, S-5742, S-5747, S-5759, S-5797 and S-5798 were relatively lesser tolerant and varietal germplasms (BR-2/1, BR-90 and BR-99) were comparatively more tolerant to sulphate toxicity.

It is known that seedlings are particularly susceptible to harsh conditions. It is indeed the most vulnerable stage in the life of a plant (Stebbins, 1971; Fenner and Thompson, 2005). Na-salts, NaCl, Na₂SO₄, and Na₂CO₃ (Greenway and Munn, 1980) and NaF (Sabal *et al.*, 2006) are reported for ionic and osmotic effects on plants. Under salinity, growth is first reduced by a decrease in the soil water potential (osmotic phase) and later a specific effect of salt causes injury. The seedlings die because of a rapid increase in salt in the cell wall or cytoplasm when the vacuole can no longer sequester incoming salt (ionic phase) (Acosta-Motos *et al.*, 2017). According to Bernstein and Hayward (1958) and Strogonov (1946) deleterious effects of saline conditions on plant growth may be attributed to two main factors namely, increase in osmotic pressure of the rooting medium and specific ions. The crop plants cannot grow in the presence of high salt levels their growth is inhibited or completely prevented at 100-200 mM NaCl (Munns and Termaat (1986). There may be several factors which may cause seedling mortality. Green house studies of onion at diurnal temperature of 15 -25 °C induced hypocotyl and root mortality under high salinity. The salinity tolerance at seed germination stage was much higher than that of seedlings in comparison (Miyamoto, 1988). Seedling mortality under salinity is the phenomenon not only associated with glycophytes or salt tolerant plants, many halophytes are known to show seedling mortality in higher salinities. Only, 2 per cent of the seedlings of *Limonium emarginatum* could survive in 2 per cent salinity (Rodondo-Gomez *et al.*, 2006). In our *in vitro* experimental set up, seedling death of guar may probably be attributed to the ionic reasons due to Na₂SO₄.

Table 20. Seedling mortality of guar germplasm under Na₂SO₄ salinity (Bar).

| Germplasms | Seedling mortality (%) | | | | | | | | | | |
|------------|------------------------|---|---|---|----------|-----------|----------|-----------|-----------|-----------|-----------|
| | 0 | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 |
| BR-2/1 | - | - | - | - | - | - | - | 16.7±6.7 | 43.3±3.3 | 76.7±8.8 | 50.0±7.7 |
| BR-90 | - | - | - | - | - | - | - | 76.7±6.7 | 13.3±3.3 | 26.7±14.5 | 30.0±5.8 |
| BR-99 | - | - | - | - | - | - | - | 20.0±0.0 | 20.0±0.0 | 16.7±13.3 | 23.3±13.3 |
| BR-2017 | - | - | - | - | - | - | 6.7±3.3 | 36.7±20.3 | 30.0±15.3 | 70.0±23.3 | 30.0±11.6 |
| S-5733 | - | - | - | - | - | 13.3±3.3 | 13.3±6.7 | 46.7±3.3 | 33.3±6.7 | 23.3±6.7 | 66.7±16.7 |
| S-5742 | - | - | - | - | 6.7±3.3 | 10.0±0.0 | 10.0±0.0 | 20.0±0.0 | 33.3±3.3 | 43.3±3.3 | 50.0±0.0 |
| S-5747 | - | - | - | - | 23.3±3.3 | 20.0±10.0 | 13.3±6.7 | 36.7±8.8 | 26.7±3.3 | 30.0±0.0 | 33.3±6.7 |
| S-5759 | - | - | - | - | 46.7±3.3 | 60.0±5.8 | 43.3±3.3 | 43.3±3.3 | 86.7±8.8 | 40.0±5.8 | 46.7±3.3 |
| S-5761 | - | - | - | - | - | - | - | 23.3±3.3 | 33.3±3.3 | 26.7±8.8 | 23.3±3.3 |
| S-5765 | - | - | - | - | - | - | 13.3±3.3 | 20.0±0.0 | 26.7±3.3 | 33.3±3.3 | 36.7±6.7 |
| S-5784 | - | - | - | - | - | - | - | 53.3±13.3 | 20.0±0.0 | 26.7±3.3 | 20.0±0.0 |
| S-5785 | - | - | - | - | - | 10.0±0.0 | 20.0±5.8 | 23.3±3.3 | 23.3±3.3 | 26.7±3.3 | 30.0±0.0 |
| S-5797 | - | - | - | - | 20.0±0.0 | 16.7±3.3 | 16.7±3.3 | 20.0±0.0 | 23.3±3.3 | 23.3±3.3 | 33.3±3.3 |
| S-5798 | - | - | - | - | 20.0±0.0 | 13.3±3.3 | 26.7±3.3 | 50.0±5.8 | 56.7±8.8 | 26.7±3.3 | 33.3±3.3 |
| S-5825 | - | - | - | - | - | - | 13.3±3.3 | 13.3±3.3 | 16.7±3.3 | 13.3±3.3 | 23.3±3.3 |

C = control (0 bar); 1 to 10 represent the concentration on Na₂SO₄ in bar. -, denotes zero mortality of seedlings.

Salt tolerance of germplasms to NaCl salinity at germination phase relative to variety BR-99

Our studies indicated that BR-99 was the most tolerant variety to NaCl salinity amongst the tested germplasms. Considering BR-99 as an ideal local guar germplasm in NaCl salinity, other germplasms were graded for their tolerance relative to BR-99. For this purpose a parameter was generated through summation of FGP and GVI for each germplasms in extreme salinity of 10 bar NaCl. According to this parameter Variety BR-99 appeared to dominate over other germplasms. The degree of tolerance of each germplasm was then calculated relative to BR-99, as percent proportion of FGP + GVI of a germplasm to the FGP + GVI of BR-99.

Fig. 17 presents the salt tolerance of all germplasms relative to Variety BR-99. The results indicated that varietal germplasms BR-2/1 and BR-90 were nearly as tolerant as BR-99. Germplasms S-5733, S-5742, S-5747, S-5759, S-5761, S-5785, S-5785 and S-5825 were 60-75% tolerant compared to the tolerance of BR-99. S-5797

showed nearly 50% relative tolerance compared to BR-99. Varietal germplasm BR-2017 and S-5798 exhibited tolerance as low as 40% of BR-99. Germplasm S-5784 was much sensitive to NaCl in comparison to BR-99. Germplasms S-5733, S-5742, S-5747, S-5759, S-5761, S-5785, S-5785 and S-5825 may further be investigated through pot culture experimentation. Moreover, there is need to evaluate all in-hand germplasms for their responses to other salts like CaCl_2 , KCl, NaHCO_3 , etc. and the mixed salts salinity to get them fully screened for tolerance to various salts. Furthermore, it is necessary that these germplasms should be tested in green house and under the field conditions of salinity. In Pakistan, the salinity-affected areas are largely due to NaCl: the studied germplasms are potentially tolerant to NaCl salinity and deserve further large scale studies.

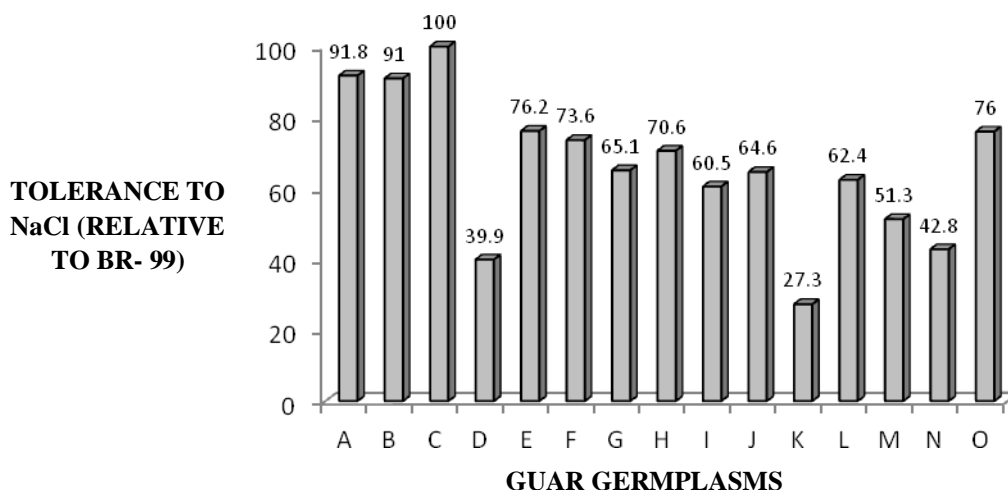


Fig.17. Showing degree of tolerance of germplasms to NaCl in comparison to BR-99 (the most salt tolerant local variety) based on an additive parameter of FGP and GVI in extreme salinity of 10 bar NaCl Data labels show the percent tolerance relative to the germplasm BR-99. Acronyms for Varieties / Lines: A, BR-2/1, B, BR-90, C, BR-99, D, BR-2017, E, S-5733, F, S-5742, G, S-5747, H, S-5759, I, S-5761, J, S-5765, K, S-5784, L, S-5785, M, S-5797, N, S-5798 and O, S-5825.

REFERENCES

- Abdou, M.A., A.A. El-Sayed, R.A.I. Taha, M.A. Abd-El-Sayed and W.S.E. Botras (2017). Effect of compost and some biostimulant treatments on guar plants. A. vegetative growth and seed yield. *Egypt Sci. J. Flowers and ornamentals Plants* 4(1): 143-157.
- Abusuwar, A.D. and J.A. Abbaker (2009). Effect of different concentrations of Red Seawater on germination and growth of some forage species. *Sudan J. Des. Res.* 1(1): 109-124.
- Acosta-Motos, J.R., M.F. Ortuño, A. Bernal-Vicente, P. Diaz-Vivancos, M.J. Sanches-Blanco and J.A. Hernandez (2017). Plant responses to salt stress: Adaptive Mechanisms. *Agronomy* 7(18): 1-38. Doi: 10.3390/agronomy7010018 (www.mdpi.com/journal/agronomy).
- Aflaki, F., M. Sadghi, A. Pazuki and M. Pessarakali (2017). Investigation of seed germination indices for early selection of salinity tolerant genotypes: A case study in wheat. *Emirates J. Food & Agriculture* 29(3): 222-226.
- Afria, B.S., N.S. Nathawat and M.L. Yadav (1998). Effect of cycocel and saline irrigation on physiological attributes, yield and its components of varieties of guar (*Cyamopsis tetragonoloba* L. Taub. *Ind. J. Pl. Physiol.* 3(1) NS: 46-48.
- Al-Yemeny, M.N. and A.Y. Basahy (1997). Salinity effect on the growth of *Cyamopsis tetragonoloba* L. *Geobios* 24(2-3): 93-98.
- Ali, Z., M. Ashraf, F. Al-Qurainy, S. Khan and N.A. Akram (2015). Field screening of guar (*Cyamopsis tetragonoloba* (L.) Taub.) Accessions for enhanced forage production on hot dry lands. *Pak. J. Bot.* 47(4):1429-1437.
- Al-Mudaris, M.A. (1998). *Notes on various parameters recording the speed of seed germination.* Derropenlandwirt, Beitage zur tropischen Land wirtschaft und veterinarmedizin, 99 Jahrgang, Oktober 98, S.147-154.
- AlShameri, A., F. Al-Qurainy, S. Khan, M. Nadeem, Abdel-Rhman Gaafar, A. Alameri, M. Tarroum, S. Alansi and M. Ashraf (2019). Morpho-physiological responses of guar [*Cyamopsis tetragonoloba* (L.) Taub.] to multiple stresses of drought, heat and salinity. *Pak. J. Bot.* 51 (3): 817-822. [(Doi: 10.30848/PJB2019-3(5). uploaded 28/3/2019)].
- AlShameri, A., F. Al-Qurainy, S. Khan, M. Nadeem, A. Ghaffar, M. Tarroum, A. Alameri, S. Alansi and M. Ashraf (2017). Appraisal of guar [*Cyamopsis tetragonoloba* (L.) Taub.] Accessions for forage purpose under the

- typical Saudi Arabian environmental conditions encompassing high temperature, salinity and drought. *Pak. J. Bot.* 49(4): 1405-1413.
- Amin, A., B.A. Arain, T. M. Jahangir, M.S. Abbasi and F. Amin (2018) Accumulation and distribution of lead (Pb) in plant tissues of guar (*Cyamopsis tetragonoloba* L.) and sesame (*Sesamum indicum* L.): profitable phytoremediation with biofuel crops, *Geology, Ecology, and Landscapes*, 2:1, 51-60, DOI: 10.1080/24749508.2018.1452464
- An Association of Official Seed Analysis (AOSA) (1990). Rules for testing seeds. (Revised). *J. Seed Technology* 12: 1-122.
- Andrade, M.I. (1985). *Physiology of salt tolerance in guar (Cyamopsis tetragonoloba (L.) Taub.* M.Sc. Thesis, The University of Arizona, USA, xi + 131 pp.
- Arora, R.N. and S.K. Pahuja (2008). Mutagenesis in guar (*Cyamopsis tetragonoloba* (L.) Taub.). *Plant Mutation Reports*, 2(1): 7-9.
- Asana, R.D. and V.R. Kale (1965) A study on salt tolerance of four wheat varieties. *Indian J. Plant Physiology*, 8: 5-22.
- Ashraf, M.Y., K. Akhtar, G. Sarwar and M. Ashraf (2005). Role of rooting system in salt tolerance potential of different guar accessions. *Agron. Sustain. Dev.* 25: 253-259.
- Ashraf, M.Y., K. Akhtari, G. Sarwar and M. Ashraf (2002). Evaluation of arid and semi-arid ecotypes of guar (*Cyamopsis tetragonoloba*. *J. Arid Environ.* 52: 476-482.
- Ayers, A.D. (1952). Seed germination as affected by soil moisture and salinity. *Agron. J.* 44: 82-84.
- Azizov, A.A. (1974). Effect of salt concentration on germination of Meyer Sea lavender seeds. *Uzb. Biol. Zh.* 18: 22-24.
- Bafeel, S.O. (2014). Physiological parameters of salt tolerance during germination and seedling growth of *Sorghum bicolor* cultivars of the same subtropical origin. *Saudi Journal of Biological Sciences*. King Saud University. 300-304. ([http://dx. Doi.org/10.1016/j-sjbs.2014.05.005](http://dx.doi.org/10.1016/j.sjbs.2014.05.005))
- Bernstein, L. and H.E. Hayward (1958). Physiology of salt tolerance. *Ann. Rev. Pl. Physiol.* 9: 25-48.
- Bina, F. and A. Bostani (2017). Effect of salinity (NaCl) stress on germination and early seedling of three medicinal plants. *Adv. Life Sciences* 4(3): 77-83.
- Carrow, R.N. and R.R. Duncan (2011). *Best management Practices For Saline and sodic Tuft Grass Soils. Assessment and Reclamation*. CRC Press- 496 Pp.
- Chaudhary, S.P.S. , N.P. Singh, R.V. Singh D.D. Saini, and O.P. Khedar (2007a). Promising guar variety RGC – 1031 (guar Kranti) for Rajasthan State. *J. Arid Legumes*, 4(1): 18-21.
- Chaudhary, S.P.S., N.P. Singh, R.V. Singh, D.D. Saini and R.V. Singh (2007b). Performance of promising guar genotypes RGC-1033 and R.G.C-1038. *J. Arid Legumes*, 4(1): 22-25.
- Chowdhury, F.M.T., M.A. Halim, F. Hossain and N. Akhtar (2018). Effect of sodium chloride on germination and seedling growth of sunflower (*Helianthus annuus* L). *Jahangirnagar Univ. J. Biol. Sci.*, 7(1): 35-44.
- Chunmei, Hu, Ji Junjie, L. Guohua and W. Xiufeng (2002). Water-absorbing trends and germinating characteristics of guar seeds. *J. Shandong Univ.*, 33 (3): 281-285.
- Cicek, N. and H. Cakirlar (2002). The effect on some physiological parameters in two maize cultivars. *Bulgarian J. Pl. Physiol.* 28(1-2): 66-74
- Cramer, G.R., E. Epstein and A. Läuchli (1988). Kinetic of root elongation of maize response to short term exposure to NaCl and elevated calcium concentration. *J. Exp. Bot.*, 39: 1513-1522.
- Datta, K.S and J. Dayal (1988). Effect of salinity on germination and early growth of guar (*Cyamopsis tetragonoloba* L.). *Indian J. Plant Physiol.*, 31 (4): 359-363.
- De Villiers, A.J., M.W. Van Rooyen, G.K. Theron and H.A. van de Venter (1994). Germination of three Namaqualand pioneer species, as influenced by salinity, temperature and light. *Seed Sci. Technol.*, 22: 427-433.
- Deepak, P., S. Singh, R.P. Gupta, J. Singh and G. Yadav (2003). High-frequency multiple shoot regeneration from cotyledonary nodes of guar (*Cyamopsis tetragonoloba* L. Taub). *In Vitro Cell. Dev. Biol. – Plant.* 39: 384-387.
- Deepika and H.R. Dhingra (2014). Effect of salinity stress on morpho-physiological, biochemical and yield characters of cluster bean (*Cyamopsis tetragonoloba* (L.) Taub. *Ind. J. Pl. Physiol.*, 19(4): 393-398.
- Dzyubenko, N.I., E.A. Dzyubenko, E.K. Potokina and S.V. Bulyntsev (2017). Cluster beans (*Cyamopsis tetragonoloba* (L. Taub. – Properties, use, plant genetic resources and expected introduction in Russia (Review). *Agricultural Biology*, 52 (6): 1116-1128. SEL'SKOKHOZYAISTNENNYA BIOLOGYA).
- ElSheikh, E.A.E. and K.A. Ibrahim (1999). The effect of Bradyrhizobium inoculation on yield and seed quality of guar (*Cyamopsis tetragonoloba* L.) *Food Chem.* 65: 183-187.
- Fenner, M. and K. Thompson (2005). *The Ecology of Seeds*. Cambridge Univ. Press, 260 Pp.
- Francois, L.E. and R. Kleiman (1990). Salinity effects on vegetative growth, seed yield and fatty acid composition of Crambe. *Agron. J.* 82: 1110-1114.

- Francois, L.E., E.V. Maas, T.J. Donovan and V.L. Youngs (1986). Effect of salinity on grain yield and quality vegetative growth and germination of semi-dwarf drum wheat. *Agron. J.* 78:1053-1058.
- Francois, L.E., T.J. Donovan and E.V. Maas (1989). Salinity effects on emergence, vegetative growth and seed yield of guar. *Crop Sci.* 82(3): 587-592.
- Ganal El-Awad El-Daw (1998). *A study of guar seed and guar gum properties (Cyamopsis tetragonoloba)*. M.Sc. Thesis, University of Khartoum, Sudan.
- Garg, B.K., S. Kathju, S.P. Vyas and A.N. Lahiri (1997). Sensitivity of cluster bean to salt stress at various growth stages. *Ind. J. Pl. Physiology* 2(1): 49-53.
- Greenway, H. and R. Munn (1980). Mechanisms of salt tolerance in non-halophytes. *Ann. Rev. Pl. Physiol.* 31: 149-190.
- Gresta, F.A., Cristando, C. Trostle, U. Anastasi, P. Guarnaccia, S. Catara and A. Onofri (2018). Germination of guar (*Cyamopsis tetragonoloba* (L.) Taub. Genotypes with reduced temperature requirements. *Aust. J. Crop Sci.* 12 (6): 954-960.
- Gul, H., R. Ahmad and M. Humayoun (2015). Impact of exogenously applied ascorbic acid on growth of some biochemical constituents and ionic composition of guar (*Cyamopsis tetragonoloba*) subjected to salinity stress. *Pakhtunkhwa J. Life Sci.* 3(1-2): 22-40.
- Gulati, D. and H.R. Dhingra (2014). *Salt tolerance of guar and its improvement by "in vitro" mutagenesis: Improvement of salt tolerance through mutagenesis*. Lambert Acad. Publishing. 156 Pp.
- Hassan, N., M.K. Hasan, M.O. Shaddam, M.S. Islam, C. Bartçular and A. El Sabagh (2018). Responses of maize varieties to salt stress in relation to germination and seedling growth. *Int. Letters of Natural Sciences* 69: 1-11.
- Hoffman, P. and H. Sachert (1967). Effect of urea on the development of *Salicornia brachystachya*: A contribution to halophyte problem. *Ber. Deut. Bot. Ges.* 80: 437-446.
- Iqbal, M.A. (2015). Cluster bean (*Cyamopsis tetragonoloba* L.) germination and seedling growth as influenced by seed invigoration techniques. *Am. Eurasian J. Agric. & Environment Sci.* 15(2): 197-204.
- Ismail, A.M.A. (1990). Germination ecophysiology in populations of *Zygophyllum qatarensis* Hadidi from contrasting habitats. *J. Arid Environ.* 18: 185-194.
- Jat, R., N. Chaurasia, A.K. Chaurasia, A. K. Prashant, K Rai, M. Kumar and V.K. Mishra (2015). Effect of priming on seed quality parameters of cluster bean (*Cyamopsis tetragonoloba* L. Taub.) seed. *Agrica* 4: 102-107.
- Joshi, A. and S. Datta (2017). Effect of sodium perchlorate treatment on germination of guar. *The Pharma Innovation J.* 6(10): 324-327.
- Karim, M.A.N., Utsunomiya and S. Shigenaga (1992). Effect of sodium chloride on germination and growth of hexaploid triticale at early seedling stage. *Jpn. J. Crop Sci.* 61: 279-284.
- Khafagy, M.A., M.M. Dorowish, S.M. Salama and E.A.M. Abo-El-Kheer (2010). Effect of presoaking guar seeds in some plant vitamins or phytohormones on germination and seedling growth in the presence of NaCl. *J. Plant Production* 1(3): 367-382.
- Khalid, M.A.UR-Rehman, L.H. Akhtar, R. Minhas, Z.A. Bukhari, M. Zubair and M.A. Iqbal (2017). Screening of guar accessions [*Cyamopsis tetragonoloba* (L.) Taub.] for higher yield potential under irrigated conditions. *Afr. J. Agric. Res.* 12(37): 2788-2794.
- Khan, D. (1987). *Phytosociological Survey of Pakistan Coast with Special Reference to Pasture and Forest Development through Biosaline Technique*. Ph.D. Thesis. Department of Botany, University of Karachi, Pakistan.
- Khan, D. and R. Ahmad (1998). Effect of saline water irrigation on germination, growth and mineral distribution in *Indigofera oblongifolia* Forssk. *Hamdard Medicus* XLI (4): 81-93.
- Khan, D., I. Jahan, L.H. Akhtar, M.J. Zaki and R. Minhas (2018). Seed mass variation in seed lots of fifteen germplasms of guar [*Cyamopsis tetragonoloba* (L.) Taub.]. *Int. J. Biol. Biotech.* 15(4): 711-720.
- Khan, D., R. Ahmad and S. Ismail (1987). Germination, growth and ion regulation in *Prosopis juliflora* (Swartz.) DC. under saline conditions. *Pak. J. Bot.* 49: 131-138.
- Khan, D., R. Ahmad, S. Ismail and H. Zaheer (1989). Effects of saline water irrigation on growth and mineral distribution in guar (*Cyamopsis tetragonoloba* (L.) Taub. *Pak. J. Bot.* 21(2): 290-301.
- Khan, D., S.S. Shaikat and M. Faheemuddin (1984). Germination studies of certain desert plants. *Pak. J. Bot.* 16(2): 231-254.
- Khan, M.S.A., A. Hamid, A.B.M. Salaududdin, A. Quasem and M.A. Karim (1997). Effect of sodium chloride on growth, photosynthesis and mineral accumulation of different types of rice (*Oryza sativa* L.) *J. Agron. Crop Sci.* 179: 149- 161.
- Khanzada, B., M.Y. Ashraf, S.A. Ala, S.M. Alam, M.U. Shirazi and S.A. Ansari (2001). Water relations in different guar (*Cyamopsis tetragonoloba* (L.) Taub.) Genotypes under water stress. *Pak. J. Bot.* 33: 279-287.
- Kumar, A.B., B. Bahadur and B.K. Sharma (1988). Effect of salinity on germination, seedling growth and some quantitative characters of *Cyamopsis tetragonoloba* (L.) Taub. *New Bot.* 15: 23-27.

- Kumar, K. T., P. Kumar, B. Patel and K. Pushpalettha (1990). Effect of chemical effluent on germination and growth of guar (*Cyamopsis tetragonoloba* L. Taub.). *J. Adv. Plant Sci.* 31(1): 34-42.
- Kumar, S. and P. Bhardwaj (1981). Studies on genotypic differences in the early seedling growth of various crop plants under salinity condition. I. Mung (*Vigna radiata* Wilczek). *Indian J. Plant Physiology* 24(2): 123-127.
- Kumar, S., A.R. Modi, M.J. Parekh, H.R. Mehta, R. Sharma, R.S. Fought, D. Yadav, N.R. Yadav and G.B. Patil (2017). Role of conventional and biotechnological approaches for genetic improvement of cluster bean. *Industrial Crop and Products* 97: 639-648.
- Lahiri, A.N., B.K. Garg, S.P. Vyas, S. Kathju and P.C. Mali (1996). Genotypic differences to salinity in cluster bean. *Arid Soil Res. Rehab.* 10: 333-345.
- Liu, W. (2003). *Evaluation of guar (Cyamopsis tetragonoloba (L.) Taub. For gum and agronomic trait quality*. Ph. D. Thesis, Texas Tech. University, USA.
- Maas, E.V. and G. J. Hoffman (1977). Crop salt tolerance- Current assessment. *J. Irrigation & Drainage. Div., ASAE* 103 (IR2): 115-134.
- Maliwal, G.L. and K.V. Paliwal (1970). Salt tolerance of some Bajra varieties at germination stage. *J. Ind. Soil. Sci.* 18: 209-214.
- Mansour, M.M.F., K.H.A. Salma, F.Z.M. Ali and A.F. Abou Hadid (2005). Cell and plant responses to NaCl in Zea mays cultivars differing salt tolerance. *Gen Appl. Plant Physiol.* 31 (1-2): 29-41.
- Mayer, A.M. and A. Poljakoff-Meyber (1975). *The Germination of Seeds*. Pergamon Press, N.Y.
- Mehta, B.V. and K.S. Desai (1958). Effect of soil salinity on germination of some seeds. *J. Soil Water Conserv., India* 6: 169-171.
- Meiri, A., J. Kamburoff and A. Poljakoff-Mayber (1971). Responses of bean plants to sodium chloride and sodium sulphate salinization. *Ann. Bot.* 35 (4): 837-847.
- Miyamoto, S. (1988). Salt effects on germination, emergence and seedling mortality of onion. *Agron. J.* 81(2): 202-207.
- Mondal, T.K., A. R. Bal and S. Dal (1988). Effect of salinity on germination and seedling growth of different rice (*Oryza sativa* L.) cultivars. *J. Indian Soc. Coastal Agric. Res.* 6: 91-97.
- Morris, J.B. (2010). Morphological and reproductive characterization of guar (*Cyamopsis tetragonoloba*) genetic resources generated in Georgia, USA. *Genet. Resour. Crop Evol.* 57:985-993.
- Muftahizade, H., Y. Hamidoghli, M.H. Assareh and M.J. Dakheli (2017). Effect of sowing date and irrigation regimes on yield components, protein and galactomannan content of guar (*Cyamopsis tetragonoloba* L.) in Iran climate. *Aust. J. Crop Sci.* 11(11): 1481-1487.
- Mulwani, B.T. and A.G. Pollard (1939). Effects of alkali salts on germination of seeds. *Agric. Livestock of India.* 9: 548-555.
- Munns, R. and A. Termaat (1986). A whole plant response to salinity. *Aust. J. Pl. Physiology*, 13: 143-160.
- Myers, B.A. and W.C. Morgon (1989). Germination of the salt tolerant grass, *Diplachne fusca*. II. Salinity responses. *Aust. J. Bot.* 37: 225-237.
- Nasr, S.M.H., A. Parsakhoo, H. Naghavi and S.K.S. Koohi (2012). Effect of salt stress on germination and seedling growth of *Prosopis juliflora* (Sw). *New Forests.* 43(1): (newforest.43.10.1007/s11056-011-9265-9).
- Nieman, R.H. and E.V. Maas (1978). 6th Intl. Biophys. Cong. Abst. Pp. 121 (cited in *Salinity Tolerances in Plants Strategies for Crop Improvement* (R.C. Staples & G.H. Toenniesen, Eds.). 1984. John Wiley & Sons, N.Y.
- O'Leary, J.W. (1986). A critical analysis of the used of Atriplex species as crop plants for irrigation with highly water. In: *Prospects For Biosaline Researches* (R. Ahmad and A. S. Pietro, Eds. US- Pakistan Biosaline Workshop Proceedings, University of Karachi (1985), Pakistan.
- Omer, EA, Fattah A., Razin M., S.S. Ahmed (1993). Effects of cutting, phosphorus and potassium on guar (*Cyamopsis tetragonoloba*) in newly claimed soil in Egyptian. *Plant Food Hum. Nutr.* 47:277-284.
- Pathak, R., Singh S.K., Singh M., Henry A. (2010). *Molecular assessment* of genetic diversity in cluster bean (*Cyamopsis tetragonoloba*) genotypes. *J. Gent.* 89:243-246.
- Patil, A.V. and B.A. Karadge (2012). Effect of NaCl salinity on the growth and mineral nutrition of one month old *Prosopis juliflora* (Sw.) DC. Seedlings. *Pharmacog. J.* 4(31): 63-66.
- Prakash, M., S. Pallavamallan, G. Sathiyarayan and S. Rameshkumar (2019). Effect of seed pelleting with botanicals on germination and seedling growth of cluster bean under induced salinity. Article ID = LR-4060. (Doi: 10.8805/LR-4060)
- Rai, P.S., Dharmatti P.R., T.R. Shashidha, R.V. Patil and R.R. Patil (2012). Genetic variability studies in cluster bean (*Cyamopsis tetragonoloba* (L.) Taub). *Karnataka J. Agric. Sci.* 25:108-111.
- Ramarajan, S., B.A. Rayan S, J.L. Henry, and S. Gandhi A. (2013). Seed germination and biochemical changes in *Cyamopsis tetragonoloba* (L.) Taub. (Var. Pusa Naubahar) under salt stress. *Research & Reviews: Journal of Crop sci. & Tech.* PRJoCST 4-9. (www.stmjournals.com)

- Ranal, MA, D.G. Santana, W.R. Ferreira and C. Mendes-Rodrigues. (2009). Calculating germination measurements and organizing spreadsheets. *Braz. J. Bot.*, 32, 849-55.
[<http://dx.doi.org/10.1590/S0100-84042009000400022>]
- Rangasamy, P. (2010). Osmotic and ionic effects of various electrolytes on growth of wheat. *Aust. J. Soil Res.* 48: 120 -124.
- Rao, N.K. and M. Shahid (2011). Potential of cowpea (*Vigna unguiculata* (L.) Walp.) and guar (*Cyamopsis tetragonoloba* (L.) Taub.) as alternative forage legume for the United Arab Emirates. *Emir. J. Food Agric.* 23(2): 147-156.
- Rasheed, M.J.Z., K. Ahmad, M. Ashraf, F. Al-Qurainy, S. Khan and Habib-Ur-Rahman Athar (2015). Screening of diverse local germplasm of guar [*Cyamopsis teteragonoloba* (L.) Taub.] for salt tolerance: A possible approach to utilize salt –affected soil. *Pak. J. Bot.* 47(5): 1721-1726.
- Raychaudhuri, S.P. (1952). Desert soil and desert farming. *Bull. Nat. Inst., India*, 1:266-268.
- Rodondo-Gomez, S., E.M. Naranjo, O. Garzón, J.M. Costillo, T. Luque and M.E. Figuerosa (2006). Effects of salinity on germination and seedling establishment of endangered *Limonium emarginatum* (Willd.) O. Kuntze. *J. Coastal Res.* 24 (1A): 201-205.
- Rozema, J. (1975). The influence of salinity, inundation and temperature on the germination of some halophytes and non-halophytes. *Oecologia Plantarum.* 10: 341-353.
- Sambangi, P. and P.U. Rani (2016). Role of physical and biochemical parameters of different cluster bean varieties on the guar gum content and aphid infestation. *Int. J. Current Res. Biosciences and Plant Biology* 3 (4): 83-90.
- Scott, S.J., R.A. Jones and W.A. Williams (1984). Review of data analysis methods for seed germination. *Crop Sci.* 24: 1192 – 1199.
- Senn, H.A. (1938). Chromosome number relationship in the Leguminosae. *Bibliog. Genetics* 12: 175-336.
- Sharma, P. (2010). *Guar Industry Vision 2020: Single Vision Strategy*. NIAM Res. Rep. Anurag Bhatnagar, IAS, Jaipur.
- Shaukat, S.S. and N. Burhan (2000). Fecundity, seed characteristics and factors regulating germination of *Rhynchosia minima* (L.) D.C. *Pak. J. Bot.* 32(1): 211-226.
- Shehata, A.M., M.A.H. Abdou, A.A. El-Sayed, F.A. Attia and R.A. Taha (2017). Organic fertilization and natural substances treatments affect chemical constituents of guar plants. *Hortic. Biotech. Res.* 3: 26-36.
- Singh, R. (2014). *Improved Cultivation Practices for Cluster Bean in Kharif and Summer Season*. CAZRI (ICAR), Jodhpur, India.
- Singla, S., K. Grover, S.V. Angadi, S.H. Begna, B. Schutte and D.V. Leeuwen (2016). Growth and yield of guar (*Cyamopsis tetragonoloba* L.) genotypes under different planting dates in the semi-arid Southern high plains. *Am. J. Plant Sci.* 7: 1246-1258.
- Soni, V., R. Sen, P. Kumar and V. Bhardwaj (2017). Effect of NaCl induced salt stress on biochemical parameters of *Cyamopsis tetragonoloba* (L.) Taub. *Int. J. Food, Agriculture and Veterinary Sciences* (2): 77-80.
- Sortino, O. and F. Gresta (2007). Growth and yield performance of five guar cultivars in a Mediterranean environment. *Ital J. Agron. / Riv. Agron.* 4: 359-364.
- Stebbins, G.L. (1971). Adaptive radiation in reproductive characteristics in Angiosperms. II: Seeds and Seedlings. *Ann. Rev. Ecol. & Systematics* 2: 237-260.
- Strogonov, B.P. (1946). On the adaptation of the cotton plant to high salinity. *Acad. Sci. URSS Compt. Reind.* 54: 453-456.
- Strogonov, B.P. (1964). *Physiological Basis of Salt Tolerance of Plants*. Academic Sci. USSR. Davey & Co., New York.
- Sudhar, J.D., I. Rajpar, G. Keshavamurthy, Ganjegunte and Zia-ul-Hasan (2018b). Salinity tolerance of guar or cluster bean (*Cyamopsis tetragonoloba* L.) Genotypes. *Poster Graduate students Competition. Ph.D. students.* Feb. 4, 2018, Jacksonville, FL., USA.
- Sudhar, J.D., I. Rajpar, G.K. Ganjegunte and Zia-ul-Hasan (2018a). Comparative study of early growth stage of 25 guar (*Cyamopsis tetragonoloba* L.) genotypes under elevated salinity. *Industrial Crops Products.* 123:164-172.
- Sultan, M., M.N., Yousaf, M.A. Rabbani, Z. K. Shinwari and M.S. Masood (2012). Phenotypic divergence in guar (*Cyamopsis tetragonoloba* L.) landrace genotypes of Pakistan. *Pak. J. Bot.* 44:203-210.
- Teolis, I. W. Liu and E. B. Peffley (2009). Salinity effects on germination and plant growth of guar. *Crop Sci.* 49(2): 637-642.
- Thiam, M., A. Champion, D. Diouf and Mame Ouréye SY (2013). NaCl effects on in vitro germination and growth of some Senegalese cow pea (*Vigna unguiculata* (L.) Walp.) Cultivars. *ISRN Biotechnology* vol. 2013, Article ID 382417, 11 pages. ([http:// dx.doi.org/10.5402/2013/382417](http://dx.doi.org/10.5402/2013/382417)). Hindawi Pub, Corp.
- Thorner, J.J. (1909). Vitality of seeds under water. *Arizona Agric. Exp. Stun. Bull.* 6: 438-439.
- Ungar, I.A. (1974). The effect of salinity and temperature on seed germination and growth of *Hordeum jubatum*. *Can. J. Bot.* 52: 1357-1362.

- Vinisky, T. and D.T. Ray (1988). Germination of guar seeds under salt and temperature stress. *J. Am. Soc. For Hortic. Sci.* 113 (3): 437-440.
- Ward, J.H. (1963). Hierarchical grouping to optimize an objective function. *J. Am. Statistical Assoc.*, 58(301): 236-244.
- Weixin, L. (2003). *Evaluation of Guar (Cyamopsis tetragonoloba (L.) Taub.) For Gum and Agronomic Trait Quality*. Ph. D. Thesis, Texas Tech. Univ. Xi + 82 PP.
- Whistler, R. I. and T. Hymowitz (1979). *Guar: agronomy, production, industrial use, and nutrition*. Purdue University Press, West Lafayette, Indiana, USA. 765:494-4773.
- Wilcox, L.V. (1959). Effects of industrial wastes on water for irrigation use. *Am. Soc. Testing Materials Spec. Tech. Publ.* 273: 58-64.
- Woodstock, L. W. (1976). *Progress Report on the Seed vigour Testing Handbook*. Association of Official Seed Analysis of America (AOSA). *Newsletter*, 59 (2): 1-78.
- Yadava, R.B.R., K.L. Mehra, K.I. Magoon, P.R. Sreenath, and M.S. Yadav (1974). Varietal differences in salt tolerance during seed germination of guar. *Ind. J. Plant Physiology* 18: 16-19.
- Zafar, S.A., S. Shokat, H.G.M. Ahmed, A. Khan, M.Z. Ali and R.M. Atif (2015). Assessment of salinity tolerance in rice using seedling based morpho-physiological indices. *Adv. Life Sci.* 2(4): 142-149.
- Zheng, G.-H., Z.-H Gu, and B.-M. Xu (1980). A physiological study of germination of guar (*Cyamopsis tetragonoloba* (L.) Taub.) seeds. *Acta Phytophysiologia Sinica* 6(2): 115-126.

(Accepted for publication April, 2019)