AJAB

Original Article

Allelopathic effect of *Lantana camara* and *Chromolaena odorata* leaf extracts on plant germination

Arjay Julio, Wynsel Carven Tandoc, Hans Daniel Tipace, Yannah Franzine Vendivil, Zyrene Yanesa, Maria Violeta R. Tare, Elmar Jon Lactaoen, Ken Joseph Clemente Science, Technology, Engineering, and Mathematics Strand Strand, Senior High School, University of Santo Tomas, Sampaloc, Manila, 1015, Philippines

Received: April 23, 2018 Accepted: January 23, 2019 Published: June 30, 2019	Abstract Allelopathy is a biological process where plants affect, often inhibitory, the growth and germination of other species within their space. This study aimed to demonstrate and compare the allelopathic effect of <i>Lantana camara</i> and <i>Chromolaena odorata</i> leaf extracts on plant germination, with <i>Vigna radiata</i> as the test plant. Leaf extracts were assayed at 10%, 25%, 50%, 75%, and 100% concentrations, and the corresponding allelopathic effects were compared to that of control. Findings indicated that <i>L. camara</i> and <i>C. odorata</i> leaf extracts inhibited <i>V. radiata</i> seedling growth and germination in increasing concentrations, with <i>C. odorata</i> leaf extract exhibiting greater inhibitory effect. The estimated marginal mean lengths (in cm) of root, hypocotyl, and epicotyl of <i>V. radiata</i> are 0.917, 5.937, and 3.263 under the control; 0.195, 0.813, and 0.499 under <i>L. camara</i> ; and 0.101, 0.217, and 0.051 under <i>C. odorata</i> , respectively. Phytochemical analysis showed presence of several allelochemicals in both leaf extracts. These compounds were suspected to be the primary drivers of the observed allelopathic effect. It is suggested that the quantitative phytochemical analysis and herbicidal properties of <i>L. camara</i> and <i>C. odorata</i> be studied further. Keywords : <i>Lantana camara</i> , <i>Chromolaena odorata</i> , Allelopathy, Invasive Alien Species, Bioassay
*Corresponding author email: keclemente@ust.edu.ph	How to cite this: Julio A, Tandoc WC, Tipace HD, Vendivil YF, Yanesa Z, Tare MV, Lactaoen EJ and Clemente KJ, 2019. Allelopathic effect of <i>Lantana camara</i> and <i>Chromolaena odorata</i> leaf extracts on plant germination. Asian J. Agric. Biol. 7(2):190-196.

This is an Open Access article distributed under the terms of the Creative Commons Attribution 3.0 License. (<u>https://creativecommons.org/licenses/by/3.0</u>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Introduction

Allelopathy is a biological process by which plants influence the growth of their neighbors through releasing allelochemicals (Sodaeizadeh and Hosseini, 2012). At low allelochemical concentrations, plants commonly exhibit stimulatory allelopathic effect on its neighbors, whereas high concentrations can suppress the growth and germination of nearby plants (El-Kenany and El-Darier, 2013). Allelopathy is primarily exhibited by invasive alien species (IAS). IAS commonly invade accidentally or deliberately places that are not part of their natural habitats, causing imbalance to the ecosystem and depletion in crop production (Callaway and Aschehoug, 2000). However, despite the inhibitory effect of IAS, their allelochemicals were potentially found to be utilized as natural herbicides against other IAS, as an

🐑 Asian J Agric & Biol. 2019;7(2):190-196. 👘 190

alternative to the commonly used synthetic herbicides that pose soil pollution and health risks to the consumers (Jabran and Farooq, 2013).

Two of the most common IAS in the Philippines are Lantana camara and Chromolaena odorata. L. camara is a perennial and invasive shrub that can be mostly found in arid areas with high temperature (Mishra, 2015). L. camara was observed to exhibit allelopathy, wherein it invades the growth of pastures, thus decreasing plant productivity (Joshi, 2006). Allelochemicals, such as sesquiterpenes, flavonoids, glycosides, and triterpenes, have been found to be present in L. camara (Mishra, 2015). C. odorata, on the other hand, is a perennial shrub that commonly grow in well-drained soils with low fertility (Mandal and Joshi, 2014). Armed with an exceedingly huge amount of allelochemicals, such as saponins, tannins, terpenoids, alkaloids, flavonoids, and glycosides, C. odorata has strong allelopathic effect wherein it invades native plant communities due to its ability to grow rapidly and its high reproducibility of seeds (Hu and Zhang, 2013).

Although natural herbicides seem promising, only limited scientific evidences have been presented to support this claim. Thus, further researches about the potential of IAS as natural herbicide should be conducted. Generally, this study compared the allelopathic effect of *L. camara* and *C. odorata* on plant germination by determining which between the two noxious invasive weeds exhibits greater inhibitory effect. Moreover, this study determined if there is a significant difference on allelopathic effect between control and increasing concentrations as well as to record presence of allelochemicals in the leaf extracts. With these, a firmer findings for more environmentalfriendly herbicides against IAS proliferation comes into availability for future researchers.

Material and Methods

Plant acquisition

Mature leaves of *L. camara* and *C. odorata*, weighing 500 g each, were acquired from infested areas in Mount Zion, Pangasinan Province, Philippines and placed in polyethylene bags until utilization. Due to easy replication, rapid growth, and observable traits in germination and growth, *Vignata radiata* was chosen as test plant. Seeds of *V. radiata* were obtained from local markets and screened for seed quality. Damaged seeds were then discarded upon visual examination. The plant identification was verified and properly

authenticated at University of Santo Tomas Herbarium (USTH).

Extract preparation

The preparation of the leaf extracts followed that of P et al. (2010). Leaves were sun-dried for 14 days and ground afterwards. The resulting fine particles were dipped into two separate pails for 24 hours, each containing 5 L of methanol, from a basis ratio of 1 g per 10 mL. The produced mixture was filtered by a fluted Whatman No. 1 filter paper (particle retention of 11 μ m). The filtrates underwent rotary evaporation for solvent separation at about 70°C, resulting to about 160 mL per leaf extract. Using the extract as stock solution, 10%, 25%, 50%, 75%, and 100% concentrations were prepared by dilution, and their allelopathic effect was tested through seed germination bioassay (Hu and Zhang, 2013).

Petri dish bioassay

Six set-ups were prepared for the germination bioassay; triplicates for L. camara leaf extract (LCLE) and triplicates for C. odorata leaf extract (COLE). Each set-up constituted six 9 cm-diameter petri dishes for different concentrations, each of which contained five V. radiata seeds with tissue as seedbed. The first petri dish was used for application of distilled water (control), and the succeeding received 10%, 25%, 50%, 75% and 100% extract concentrations. About 5 mL of each concentration was added to each petri dish for the first day. The seeds were kept moist by applying about 1 mL of each extract daily for the next 13 days. After two weeks of observation, germinated seeds were counted based on the presence of radicle (Ahmed et al., 2007). Root, epicotyl, and hypocotyl of seedlings were also measured using a foot ruler.

Phytochemical analysis

LCLE and COLE were subjected to phytochemical analysis using Molisch, Fehling, Alkaline Reagent, Lead Acetate, Hager, Mayer, Wagner, Ferric Chloride, Borntrager, Keller-Kiliani, Froth, Resin, Liebermann-Buchard, and Salkowski tests to assess the presence of several allelochemicals (Usunomena and Efosa, 2016; Yadav and Agarwala, 2011).

Data analysis

Germination percentage and marginal mean length were used to determine the magnitude of inhibitory effect on *V. radiata* seedlings. Seedling measurements underwent two-way MANOVA to test the differences



between treatments and the control using SPSS version 23. Statistical significance was accepted at p<0.05; otherwise, rejected.

Results and Discussion

Allelopathic effect is exhibited when there is a negative effect in seedling growth and germination, compared to seedlings under the control. Results indicated that there is accumulating evidence on the inhibitory effect of LCLE and COLE on *V. radiata* seedling conditions, seen in the decrease in seedling growth and germination (Abhilasha et al., 2008; Maghrebi and Mirshekari, 2011; Orr et al., 2005). These negative effects showed that the responses of *V. radiata* seedlings are concentration dependent and species specific (Table 1). Findings in this study were found to be consistent with reports of Ahmed et al. (2007), and Hu and Zhang (2013).

Concentration dependence

Generally, there was a significant difference (p<0.05) between the control and increasing concentrations (Table 2). However, there was no significant

difference (p>0.05) between 50%, 75%, and 100% concentrations with respect to effect on root and hypocotyl length; and between 25%, 50%, 75%, and 100% in epicotyl length. This implies that very few to no seeds germinated under these treatments. Table 1 shows how germination percentage lowers with increasing concentrations. Maximum seed germination, wherein all the seeds germinated, was observed under the control, and 10% LCLE and COLE; and then lowered as the concentration was increased. No seed germination was evident under 50% COLE, and 75% LCLE. Moreover, Figure 1 show differences in marginal mean length of root, hypocotyl, and epicotyl of seedlings under the control, and different treatments. Seedling growth with respect to root, hypocotyl, and epicotyl length is maximum control, and lowers under with increasing concentration. Growth has already been impeded starting from 75% in root, 50% in hypocotyl, and 25% in epicotyl. Due to these negative effects, the relationship between increasing concentration and corresponding inhibitory effect is directly proportional.

Table 1. Germination percentage of V. radiata under different concentrations of LCLE and COLE

	Control	LCLE			COLE						
Concentration	0	10	25	50	75	100	10	25	50	75	100
No. of Germinated Seeds	15*	15*	15*	7*	0^*	0^*	15*	10^{*}	0^{*}	0^*	0^*
Germination Percentage	100	100	100	47	0	0	100	67	0	0	0

*15 total number of seeds were initially present.

Table 2. MANOVA results (Post Hoc Test – Tukey HSD) of petri dish bioassay of V. radiata seedling root,
hypocotyl, and epicotyl under different concentrations

		Root		Нуросо	tyl	Epicotyl	
(X) Concentration	(Y) Concentration	Std. Error	Sig.	Std. Error	Sig.	Std. Error	Sig.
	10	.04287 .000	.000	.15503	.000	.16214	.000
	25						
0	50						
	75						
	100				ļ		
10	25	.03500	.000	.12658	.000	.13238	.000
	50						
	75						
	100						
	50	.03500	.000	.12658	.010	.13238	1.000
25	75						
	100						
50	75	.03500	.766	.12658	1.000	.13238	1.000
	100						
75	100	.03500	1.000	.12658	1.000	.13238	1.000

The standard error and significant difference is directed to the corresponding X and Y concentration.





Figure 1. Estimated marginal mean lengths of V. radiata based on concentrations

Species specificity

Table 3 shows that significant differences (p<0.05)were present between the effects of control, LCLE, and COLE to V. radiata germination and growth. In Table 1, maximum seed germination was evident under the control, 10% LCLE and COLE, and 25% LCLE. Different from 25% LCLE, only ten seedlings germinated under 25% COLE. Moreover, COLE already obstructed seed germination at 50%; while LCLE stopped germination at 75%, with seven germinated seeds still present under 50%.

Table 3. MANOVA results (Post Hoc Test – Tukey HSD) of petri dish bioassay of V. radiata seedlings under different treatments

	(X)	(Y)	Std.	Sig.
	Treatment	Treatment	Error	
Deet	Control	LCLE	.03834	.000
Root	Control	COLE	.03034	.000
	LCLE	COLE	.02214	.000
	Control	LCLE	.13867	.000
Hypocotyl		COLE	.13807	.000
	LCLE	COLE	.08006	.000
Epicotyl	Control	LCLE	.14502	.000
		COLE	COLE .14502	
	LCLE	COLE	.08373	.000

The standard error and significant difference is directed to the corresponding X and Y concentration.

Furthermore, Figure 2 shows that growth of V. radiata seedlings is highest in control, lower in LCLE, and lowest at COLE. Due to lower germination and smaller mean lengths, greater inhibitory effect is



radiata based on extract source

Allelopathy

Allelochemicals drive allelopathy that serves as a plant's protection against herbivory, and way to eliminate competitors (Dewick, 2009). Phytochemical results showed presence of allelochemicals, such as flavonoids, alkaloids, tannins, saponins, and triterpenoids in LCLE and COLE, which caused allelopathic effect exhibited in the experiment (Table 4). However, glycosides were absent in COLE, contrary to Mishra's (2015) finding. This can be attributed to insufficient sensitivity of qualitative tests, and the amount of concentration that was tested by the laboratory, as glycosides can only be found at low concentrations (Chiejina and Onaebi, 2016). These allelochemicals present in LCLE and COLE impose mechanisms on plant growth and germination. Flavonoids disrupt photosynthetic processes by reducing the electron transport rate and inducing membrane depolarization causing the ineffectiveness of the selective permeability of cells (Huang et al., 2015). Alkaloids affect physiological processes in the plants by increasing the temperature of the DNA and preventing translation cleavage. and transcription of DNA (Cheng et al., 2015). Tannins directly affect cellular respiration through inhibiting peroxidase activities, catalase, and cellulose (Li et al., 2010). Saponins cause damage to cell membranes, by disrupting the membrane domain, creating pores, and vesiculation (Augustin et al., 2012) while triterpenoids inhibit seed germination, radicle growth, and the function of photosystem II (Wang et al, 2014).

Metabolites	LCLE	COLE		
Carbohydrates	-	-		
Reducing Sugars	-	-		
Flavonoids	+	+		
Alkaloids	+	+		
Tannins	+	+		
Glycosides	-	-		
Saponins	+	+		
Resins	-	-		
Sterols	+	+		
Triterpenoids	+	+		

Table 4. Phytochemical composition of LCLE and COLE based on presence (+) or absence (-)

Despite finding similar allelochemicals in LCLE and COLE, COLE exhibited greater allelopathic effect due to lower germination percentage and seedling growth. This may be attributed to higher amounts of allelochemical concentration in COLE than LCLE, but quantitative phytochemical analysis must be performed for confirmation. In Rusdy (2015), C. odorata was reported to have higher inhibitory effect than L. camara on the Leucaena leucocephala seedling growth, wherein findings in this research coincides with. However, Rusdy and Ako (2017) revealed that the inhibitory effect observed in the germination and seedling growth of Centrosema pubescens was more pronounced with L. camara than C. odorata. These differences in allelopathic effect indicate that the responses and corresponding effects are concentration dependent and species specific, varying depending on the plant source of allelochemicals and its level of concentration, and to what specific plant receives the effects (Ahmed et al., 2007; Hu and Zhang, 2013)

However, the petri dish bioassay conducted may overestimate the allelopathic effect of allelochemicals compared to actual field situations wherein several extraneous factors are present, such as compounds in soil, water, and temperature (Hierro and Callaway, 2003). Therefore, a combination of laboratory and field experiments are recommended to determine the persistence and bioactivity of allelochemicals *in situ* (Hu and Zhang, 2013; Inderjit & Foy, 2001).

Conclusion

This study shows that the leaf extracts of *Chromolaena odorata* and *Lantana camara* can

suppress the growth and germination of *Vigna radiata* seedlings. The inhibitory effect is directly proportional with the amount of extract concentration with *C. odorata* causing greater inhibition than *L. camara*. Several allelochemicals were found on both plants, confirming their allelopathic ability. Moreover, quantitative analysis, and combination of laboratory and field experiments are recommended to efficiently explore the allelopathic effect of these plants and their potential application as natural herbicides.

Acknowledgment

This study would have been impossible if not for the following people who had crucial contribution to our endeavor: Jennifer D. Julio, Roel C. Julio, Winston P. Tandoc, Maria Cecille R. Tandoc, Mary Ann D. Tipace, Edwin M. Tipace, Mercedita T. Vendivil, Narciso V. Yanesa Jr., and Nesie V. Yanesa. We would also like to express our gratitude to the administration of the University of Santo Tomas and the STEM Strand Chairperson, Mr. Louie B. Dasas, for their support.

Contribution of Authors

Julio A: Conceived Idea, Literature Search, Data Collection, Literature Review, Data Interpretation, Statistical Analysis, Manuscript Writing Tandoc WC: Designed Research Methodology, Literature Search, Data Collection Literature Review, Manuscript Writing Tipace HD: Designed Research Methodology, Literature Search, Data Collection, Literature Review, Manuscript Writing Vendivil YF: Literature Search, Literature Review, Manuscript Writing Yanesa Z: Designed Research Methodology, Literature Search, Literature Review, Manuscript Writing Tare MV: Manuscript Analysis and Approval Lactaoen EJ: Manuscript Analysis and Approval Clemente KJ: Manuscript Analysis, and Approval, Publication Correspondence

Disclaimer: None. **Conflict of Interest:** None. **Source of Funding:** None.

References

- Abhilasha D, Quintana N, Vivanco J and Joshi J, 2008.Do allelopathic compounds in invasive *Solidago canadensis* s.L. restrain the native European flora?J. Ecol. 96(5): 993-1001.
- Ahmed R, Udin MB, Khan MASA, Mukul SA and Hossain MK, 2007. Allelopathic effects of *Lantana camara* on germination and growth behavior of some agricultural crops in Bangladesh. J. For. Res. 18(4): 301-304.
- Augustin JM, Drok S, Shinoda T, Sanmiya K, Nielsen JK, Khakimov B, Olsen CE, Hansen EH, Kuzina V, Ekstrøm CT, Hauser T and Bak S, 2012. UDP-glycosyltransferases from theUGT73C subfamily in *Barbarea vulgaris* catalyse Sapogenin 3-O-glucosylation in Saponin-mediated Insect resistance. Plant Phys. 160(4): 1881-1895.
- Callaway R and Aschehoug E, 2000. Invasive plants versus their new and old neighbors: a mechanism for exotic invasion. Sci. 290(5491): 521-523.
- Cheng F and Cheng Z, 2015. Research progress on the use of plant allelopathy in agriculture and the physiological and ecological mechanisms of allelopathy. Front. Plant Sci. 6(1020): 1-16.
- Chiejina NV and Onaebi CN, 2016. Phytochemical constituents and antifungal properties of *Chromolaena odorat*a L. and *Moringa oleifera* Lam on fungal rot of cucumber (*Cucumis sativus* L.) Fruit. J. Asian. Plant Sci. 15(1): 35-41.
- Dewick PM, 2009. Medicinal natural products: a biosynthetic approach, 3rd ed. Wiley, United Kingdom.
- El-Kenany E and El-Darier S, 2013. Suppression effects of *Lantana camara* L. aqueous extracts on germination efficiency of *Phalaris minor* Retz. and *Sorghum bicolor* L. (Moench). J. of Taibah Univ. for Sci. 7: 64-71.
- Hierro JL and Callaway RM, 2003. Allelopathy and exotic plant invasion. Plant Soil. 256(1): 29-39.
- Hu G and Zhang Z, 2013. Allelopathic effects of *Chromolaena odorata* on native and non-native invasive herbs. J. Food Agric. Environ. 11(1): 878-882.
- Huang H, Xiao X, Ghadouani A, Wu J, Nie Z, Peng C, Xu X and Shi J, 2015. Effects of natural flavonoids on photosynthetic activity and cell integrity in *Microcystis aeruginosa*. Toxins. 7(1): 66-80.

- Inderjit M and Foy CL, 2001. On the significance of field studies in allelopathy. Weed Tech. 15(4): 792-797.
- Jabran K and Farooq M, 2013. Implications of potential allelopathic crops in agricultural systems, pp. 349-385. In Z.A. Cheema, M. Farooq and A. Wahid (eds.), Allelopathy -Current Trends and Future Applications. Springer-Verlag Berlin Heidelberg, Germany.
- Joshi R, 2006. Invasive alien species (IAS): concerns and status in the Philippines. Phil. Rice Res. Ins. 11: 1-23.
- Li ZH, Wang Q, Ruan X, Pan CD and Jiang DA, 2010. Phenolics and plant allelopathy. Molecules. 15(12): 8933-8952.
- Maghrebi T and Mirshekari B, 2011. Allelopathic effects of rapeseed residue on germination and seedling growth of redroot pigweed (*Amaranthus retroflexus*). J. Food Agric. Environ. 9(3): 646-648.
- Mandal G and Joshi S, 2014. Invasion establishment and habitat suitability of *Chromolaena odorata* (L.) King and Robinson over time and space in the western Himalayan forests of India. J. Asia Pac. Biodivers. 7(4): 391-400.
- Mishra A, 2015. Allelopathic properties of *Lantana camara*. Int. Res. J. Basic Clin. Stud. 3(1): 13-28.
- Orr SP, Rudgers JA and Clay K, 2005. Invasive plants can inhibit native tree seedlings: Testing potential allelopathic mechanisms. Plant Ecol. 181(2): 153-165.
- P MK, Rathinam X, Kasi M, Ayyalu D, Surash R, Sadasivam K and Subramaniam S, 2010. A comparative study on the antioxidant activity of methanolic leaf extracts of *Ficus religiosa* L., *Chromolaena odorata* (L.) King & Robinson, *Cynodon dactylon* (L.) Pers. and *Tridax procumbers* L. Asian Pac. J. Trop. Med. 3(5): 348-350.
- Rusdy M, 2015. Allelopathic effect of aqueous extracts of *Lantana camara* and *Chromolaena odorata* on germination and seedling growth of *Leucaena leucocephala*. Int. J. Sci. Res. 4(7): 2588-2591.
- Rusdy M and Ako A, 2017. Allelopathic effect of *Lantana camara* and *Chromolaena odorata* on germination and seedling growth of *Centrosema pubescens*. Int. J. Appl. Environ. Sci. 12(10): 1769-1776.

Asian J Agric & Biol. 2019;7(2):190-196. 195

- Usunomena U and Efosa EG, 2016. Phytochemical analysis, mineral composition and In Vitro antioxidant activities of *Chromolaena odorata* leaves. ARC J. Pharm. Sci. 2(2): 16-20.
- Wang CM, Chen HT, Li TC, Weng JH, Jhan YL, Lin SX and Chou CH, 2014. The role of pentacyclic triterpenoids in the allelopathic effects of *Alstonia scholaris*. J. Chem. Ecol. 40(1): 90-98.
- Sodaeizadeh H and Hosseini Z, 2012. Allelopathy: an environmentally friendly method for weed control. Int. Conf. Appl. Life Sci. 387-382.
- Yadav R and Agarwala M, 2011. Phytochemical analysis of some medicinal plants. J. Phyt. 3(12): 10-14.