Effect of berberis vulgaris fruit extract on Gentamicin induced Histopathological changes in liver of Albino rats

Saima Athar, Lubna Faisal, Zia-ul-Islam, Asma Basharat, Muhammad Mohtasheem-ul-Hasan, Bushra Shaikh

Department of Anatomy, Liaquat National Hospital Medical College, Jinnah Medical Dental College, University of Karachi, Ghulam Muhammad Mahar Medical College, Sukkur, Pakistan

Objective: To evaluate the effect of the Berberis vulgaris fruit extract (BVFE) on the hepatic cells against cytotoxicity induced by gentamicin.

Methodology: This experimental study was conducted at department of Anatomy, Liaquat National Hospital &Medical College, Karachi, Pakistan for a period of four weeks in March 2020. The severity of liver damage was observed in randomly selected four different visual fields of each slide under light microscopy, using the following parameters: hepatolobular architecture, arrangement of hepatocytes within lobules, presence of congestion, hydropic degeneration & vacuolization, inflammatory

infiltrate, mean hepatocyte count and mean nuclear diameter.

Results: There was statistically significant difference in mean hepatocyte count (p = 0.00) while no significant difference in the mean nuclear diameter among various groups.

Conclusion: Berberis vulgaris fruit extract is hepatoprotective against the damaging effects of drugs like gentamicin in albino rats and the protective effects are better at higher doses.

Keywords: Berberis vulgaris fruit extract, gentamicin, albino rats, hepatoxicity.

INTRODUCTION

Liver regulates metabolic functions of the body including detoxification of drugs. Drug induced hepatotoxicity is a potentially fatal and a major clinical challenge for the physicians. ^{1,2} It is caused by medications, herbal and dietary supplements or other xenobiotics. ³ Drug induced hepatotoxicity can be Intrinsic or Idiosyncratic. Intrinsic refers to liver toxicity induced by a drug in a predictable and dose-related manner while Idiosyncratic is associated with a less consistent dose-toxicity relationship. ^{4,5} More than 900 drugs, toxins, and herbs have been reported to cause liver injury, among them antibiotics are the most common cause. ⁶

Clinical manifestation range from elevation of liver enzymes with or without symptoms to development of severe liver injury leading to acute hepatic failure. Gentamicin is a potent bactericidal agent belonging to aminoglycoside group of antibiotics. It is used against aerobic gram negative bacteria and is a widely used for the prophylaxis and treatment of urinary tract and abdominal infections. It is a hepatotoxic and produces tissue injury by free radical damageand by apoptosis. It produces marked elevation in the levels of ALT, AST and a decrease in total serum protein and albumin level. 12,13

Liver damage produced by the hepatotoxic drugs can be

reverted or treated by plant extracts. ¹⁴ Berberis vulgaris is one such medicinal plant which is used to treat hepatic disorders. ¹² There are variety of alkaloids in the various parts of this plant like fruit, leaves, root, the most important of which is berberine; this alkaloid can exert different effects including antioxidant, anti-inflammatory, hypoglycemic, hypotensive, and hypolipidemic. ¹⁵ Berberine protects from hepatic tissue destruction by reducing steatosis, necrosis and myofibroblast and inflammatory cell proliferation in hepatocytes. ¹⁶

METHODOLOGY

This experimental study was conducted at department of Anatomy, Liaquat National Hospital &Medical College, Karachi, Pakistan for a period of four weeks in March 2020. The study was approved by the institutional Research & Ethical review committee (Letter No: 0436-2020LNH-ERC). Thirty two male albino rats, weighing 160-230 gm were housed in clean, properly ventilated cages in a temperature-controlled room and had free access to standard animal diet and water throughout the experiment.

The Berberis Vulgaris fruit was purchased from local herbal market of Karachi and confirmed by a botanist. 1000 gm of dried and powdered fruit was soaked in adequate volume of ethanol and water 70:30, stirred in

an orbital shaker and the extract was obtained using percolation method. The extract was filtered and evaporated to obtain semisolid syrup.¹⁷ The rats were randomly divided into four groups (n = 8) and the drugs were administered using animal feeding intubation needle (BVFE) and intra-peritoneally using insulin syringe (gentamicin). Group A (control group) received normal saline daily for 21 days. Group B (gentamicin group) received 80 mg/kg injection gentamicin intraperitoneally daily for 21 days. Group C (gentamicin + received gentamicin BVFE100) 80 intraperitoneally and BVFE 100 mg/kg orally for 21 days. Group D (gentamicin + BVFE200) received gentamicin 80 mg/kg intraperitoneally and BVFE 200 mg/kg orally for 21 days.

At the end of the experimental period, all animals were weighed for final body weight, anesthetized and sacrificed. Livers were removed after careful dissection, washed with saline and photographed for gross appearance. The specimen was preserved in 10% formalin solution. The specimens embedded in paraffin were cut into 5 micron thick sections, stained with hematoxylin and eosin (H&E) stain, examined under light microscopy and photographed with digital camera using Moticam 1080 HDMI & USB. 18,19

The severity of liver damage was observed in randomly selected four visual fields of each slide under light microscopy, following using the parameters: hepatolobular architecture, the arrangement hepatocytes within lobules, presence of congestion, hydropic degeneration & vacuolization, inflammatory infiltrate, mean hepatocyte count and mean nuclear diameter. These microscopic findings were calculated separately and labeled as normal or absent (0), mild (1), moderate (2), and severe (3) for each parameter. 19

Statistical Analysis: The data were analysed using SPSS 22. Morphometric parameters like hepatocyte count and nuclear diameter of various treatment groups was performed using one-way analysis of variance (ANOVA). Chi square test was used to compare the histopathological changes in the liver tissue.p<0.05 was considered significant.

RESULTS

Mean initial weight of groups A, B, C & D was 173.5, 200.37, 200.75 & 174.75 gm, respectively (Table1). Mean liver weight of group A and D was 6.48 ± 0.30 & 6.54 ± 0.59 gm while group B & C was 7.85 ± 1.27 & 7.34 ± 0.59 gm, respectively.

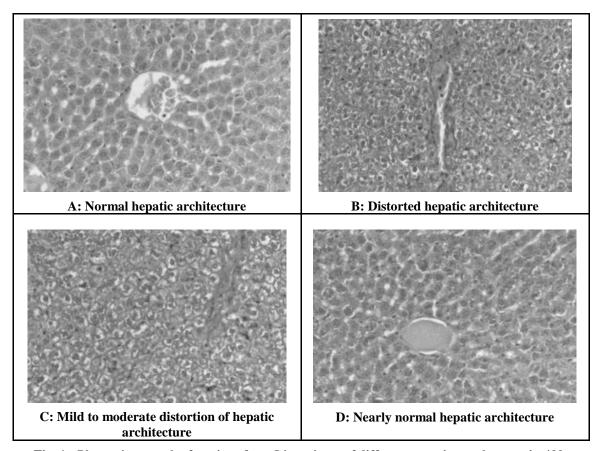


Fig. 1: Photomicrograph of sections from Liver tissue of different experimental groups' ×400.

Table 1: Statistical comparison of weight between groups at the start of experiment.

Groups	N	Mean	Std. Deviation	sig	
Group A	8	173.5000	7.59699		
Group B	8	200.3750	24.40104	0.016*	
Group C	8	200.7500	32.49066		
Group D	8	174.7500	10.44373		
Total	32	187.3438	24.31196		

Table 2 shows the comparison of histopathologic features among different experimental groups. The microscopic examination of liver sections of the animals in the control group (Group A) showed normal hepatic architecture, with hepatic lobules containing regularly arranged hepatocytes and no lymphocytic infiltration (Fig. 2-A), while group B (gentamicin) showed moderate to severe changes with loss of normal lobular architecture. There was statistically significant difference in mean hepatocyte count (p = 0.00) while no significant difference was found in the mean nuclear diameter among various groups (Table 3).

Table 2: Comparison of histopathological changes in liver within different experimental groups (n = 8).

	Group A Control Group	Group B Gentamicin Group	Group C Gentamicin+ BVFE 100 mg/kg Group	Group D Gentamicin + BVFE 200 mg/kg Group	P-value
Classic hexagonal lobule					
Normal	8 (100%)	0 (0.0%)	0 (0.0%)	2 (25.0%)	
Mild distortion	0 (0.0%)	0 (0.0%)	1 (12.5%)	6 (75.0%)	0.000*
Moderate distortion	0 (0.0%)	3 (37.5%)	5 (62.5%)	0 (0.0%)	
severe distortion	0 (0.0%)	5 (62.5%)	2 (25.0%)	0 (0.0%)	
Hepatocyte in cords					
Normal	8 (100%)	0 (0.0%)	0 (0.0%)	4 (50.0%)	
Mild distortion	0 (0.0%)	0 (0.0%)	1 (12.5%)	3 (37.5%)	0.000*
Moderate distortion	0 (0.0%)	3 (37.5%)	5 (62.5%)	1 (12.5%)	
severe distortion	0 (0.0%)	5 (62.5%)	2 (25.0%)	0 (0.0%)	
Portal vein, Hepatic artery					
Bile duct					0.001*
Normal	8 (100%)	2 (25.0%)	1 (12.5%)	6 (75.0%)	0.001*
Compressed	0(0.0%)	6 (75.0%)	7 (87.5%)	2 (25.0%)	
Mono nuclear cell infiltrate					
None	8 (100%)	0 (0.0%)	1 (12.5%)	5 (62.5%)	
Mild	0 (0.0%)	0 (0.0%)	2 (25.0%)	3 (37.5%)	0.000*
Moderate	0 (0.0%)	5 (62.5%)	4 (50.0%)	0 (0.0%)	
Severe	0 (0.0%)	3 (37.5%)	1 (12.5%)	0 (0.0%)	
Sinusoidal Congestion		,	, ,	,	
None	8 (100%)	0 (0.0%)	0 (0.0%)	4 (50.0%)	
Mild	0 (0.0%)	0 (0.0%)	4 (50.0%)	2 (25.0%)	0.000*
Moderate	0 (0.0%)	4 (50.0%)	4 (50.0%)	2 (25.0%)	
Severe	0 (0.0%)	4 (50.0%)	2 (25.0%)	0 (0.0%)	
Vacuolated Hepatocytes			, ,	, ,	
None	8 (100%)	0 (0.0%)	0 (0.0%)	1 (12.5%)	
Mild	0 (0.0%)	0 (0.0%)	4 (50.0%)	7 (87.5%)	0.000*
Moderate	0 (0.0%)	4 (50.0%)	4 (50.0%)	0 (0.0%)	
Severe	0 (0.0%)	4 (50.0%)	0 (0.0%)	0 (0.0%)	

^{*}Significant at 0.05.

Table 3: Mean comparison of Morphometric parameters.

		Mean	Std. Deviation	P-Value	
	Group A	34.8750	9.14076		
	Group B	20.1250	2.74838		
Hepatocyte Count	Group C	16.0000	2.50713	0.000*	
	Group D	26.5000	2.61861		
	Total	24.3750	8.71317		
	Group A	28.0425	3.36721		
	Group B	30.0713	3.77871		
Nuclear Diameter	Group C	30.0950	3.61254	0.579	
	Group D	29.1600	2.54000		
	Total	29.3422	3.30394		

ANOVA was applied. *Significant at 0.05.

DISCUSSION

Liver is the main organ at risk along with kidneys, owing to its role in detoxification of drugs. ²⁰ This study confirmed the harmful effects of gentamicin on the liver of albino rats as shown by the histopathologic examination, similar findings were observed in other studies after administration of gentamicin at a dose of 80 mg/kg. ²¹ There was a marked decrease in the final body weight of the rats receiving gentamicin as compared to control and BVFE – treated groups which is compatible with other studies confirming the damage produced by oxidative stress due to this drug. ^{22,23}

Gentamicin produces free radicals which generates toxic effects on the cellular structure of liver tissue which was observed in the present study as loss of normal lobular architecture, irregular arrangement of liver hepatocytes and vacuolar degeneration.²⁴ This is in agreement with the earlier studies reported by Serges et al.²³ Findings of this study are consistent with an earlier study done by Rahimi etal documenting the antioxidant properties of berberine present in BVFE at a higher dose.¹⁶

Previous studies have shown that rats treated with BVFE showed mild inflammatory changes like mononuclear infiltrate and glycogen deposition within the hepatocytes, ¹⁶ which was also observed in the present study. These results are in contrast to a study which showed that BVFE could not cure the damage in the liver tissue of tested rats. ²⁰

The findings of this study have shown that the appropriate dose of BVFE can accelerate recovery in the parenchyma of the organ in albino rats against gentamicin induced hepatoxicity. Further studies are

required at a larger scale to investigate, the mechanism of action of BVFE against gentamicin induced biochemical and histopathologic disturbances in the liver.

CONCLUSION

This study showed that Berberis vulgaris fruit extract was hepatoprotective against the damaging effects of gentamicin in albino rats and the protective effects were better at higher doses.

Author Contributions:

Conception and design: Saima Athar, Lubna Faisal, Zia-ul-Islam.
Collection and assembly of data: Lubna Faisal, Asma Basharat.
Analysis and interpretation of data: Saima Athar, Bushra Sheikh.
Drafting of the article: Lubna Faisal, Muhammad Mohtasheem-ul-Hasan
Critical revision of article for important intellectual content: Zia-ul-Islam.
Statistical expertise: Saima Athar, Bushra Sheikh.
Final approval and guarantor of the article: Saima Athar.

Corresponding author email: Saima: saimaathershaikh@gmail.com Conflict of Interest: None declared.

Rec. Date: Feb 10, 2021 Revision Rec. Date: Aug 17, 2021 Accept Date: Feb 2, 2022.

REFERENCES

- Karimi-Khouzani O, Heidarian E, Amini SA. Antiinflammatory and ameliorative effects of gallic acid on fluoxetine-induced oxidative stress and liver damage in rats. Pharmacol Rep. 2017; 69: 830-5.
- Mahmoodzadeh Y, Mazani M, Rezagholizadeh L. Hepatoprotective effect of methanolic Tanacetumparthenium extract on CCl4-induced liver damage in rats. Toxicol Rep. 2017; 4: 455-62.
- Andrade RJ, Chalasani N, Björnsson ES, Suzuki A, Kullak-Ublick GA, Watkins PB, et al. Drug-induced liver

- injury. Nat Rev Dis Prim. 2019; 5: 1-22.
- Yokoi T, Oda S. Models of idiosyncratic drug-induced liver injury. Ann Rev Pharmacol Toxicol. 2021; 61: 247-68.
- 5. McGill MR, Jaeschke H. Animal models of drug-induced liver injury. Biochimica et Biophysica Acta (BBA). Mol Bas Dis. 2019; 1865: 1031-9.
- Hoofnagle JH, Björnsson ES. Drug-induced liver injury—types and phenotypes. New Eng J Med. 2019; 381: 264-73.
- 7. Garcia-Cortes M, Robles-Diaz M, Stephens C, Ortega-Alonso A, Lucena MI, Andrade RJ. Drug induced liver injury: An update. Arch Toxicol. 2020; 27: 1-27.
- 8. Al-Doaiss AA, Jarrar YB. Investigation of in vivo protective effect of orally administered vitamin E and selenium against gentamicin-induced renal and hepatic toxicity. Trop J Pharm Res. 2019; 18: 23-5.
- 9. Mourad AM, de Carvalho Bricola SA, Sasagawa SM, Berezin EN, Ueda SM. Pharmaceutical care supported by the evidencing of in vitro interactions with the use of aminoglycosides and penicillins in patients affected by Enterococcus sp. African J Pharm Pharmacol. 2017; 11: 271-8.
- Yarijani ZM, Najafi H, Shackebaei D, Madani SH, Modarresi M, Jassemi SV. Amelioration of renal and hepatic function, oxidative stress, inflammation and histopathologic damages by Malvasylvestris extract in gentamicin induced renal toxicity. Biomed Pharm. 2019; 112: 108635.
- 11. Ali FE, Hassanein EH, Bakr AG, El-Shoura EA, El-Gamal DA, Mahmoud AR. Ursodeoxycholic acid abrogates gentamicin-induced hepatotoxicity in rats: Role of NF-κB-p65/TNF-α, Bax/Bcl-xl/Caspase-3, and eNOS/iNOS pathways. Life Sci. 2020; 254: 117760.
- 12. Afsharinasab M, Mohammad-Sadeghipour M, Hajizadeh MR, Khoshdel A, Mirzaiey V, Mahmoodi M. The effect of hydroalcoholic Berberis integerrima fruits extract on the lipid profile, antioxidant parameters and liver and kidney function tests in patients with nonalcoholic fatty liver disease. Saudi J Biol Sci. 2020; 27: 2031-7.
- 13. Arjinajarn P, Chueakula N, Pongchaidecha A, Jaikumkao K, Chatsudthipong V, Mahatheeranont S, et al. Anthocyanin-rich Riceberry bran extract attenuates gentamicin-induced hepatotoxicity by reducing oxidative stress, inflammation and apoptosis in rats. Biomed Pharm. 2017; 92: 412-20.

- 14. Niazi M, Veiskaramian A, Mokhayeri Y. Toward Nonalcoholic Fatty Liver Treatment; A Review on Herbal Medicine Treatment. J Crit Rev. 2020; 7: 554-64.
- Laamech J, El-Hilaly J, Fetoui H, Chtourou Y, Gouitaa H, Tahraoui A, et al. Effects on oxidative stress and liver injury in lead-intoxicated mice. J Comp Int Med. 2017; 14: 57-9.
- 16. Rahimi-Madiseh M, Karimian P, Kafeshani M, Rafieian-Kopaei M. The effects of ethanol extract of Berberis vulgaris fruit on histopathological changes and biochemical markers of the liver damage in diabetic rats. Iranian J Basic Med Sci. 2017; 20: 552-7.
- 17. Minaiyan M, Ghannadi A, Mahzouni P, Jaffari-Shirazi E. Comparative study of Berberis vulgaris fruit extract and berberine chloride effects on acetic acid-induced colitis in rats. Iranian journal of pharmaceutical research: IJPR. 2011; 10: 97-9.
- 18. Hassan NF, Soliman GM, Okasha EF, Shalaby AM. Histological, immunohistochemical, and biochemical study of experimentally induced fatty liver in adult male albino rat and the possible protective role of pomegranate. J Mic Ultras. 2018; 6: 44-7.
- 19. Abbasi MM, Hassanalilou T, Khordadmehr M, Vardin AM, Kohlan AB, Khalili L. Effects of Cornus mas Fruit Hydro-Methanolic Extract on Liver Antioxidants and Histopathologic Changes Induced by Cisplatin in Rats. Indian J Clin Biochem. 2020; 35: 218-24.
- Tahmasebi M, Sadeghi H, Nazem H, Kokhdan EP, Omidifar N. Hepatoprotective effects of Berberis vulgaris leaf extract on carbon tetrachloride-induced hepatotoxicity in rats. J Educ Health Promot. 2018; 7: 147.
- 21. Aboubakr M, Abdelazem MA. Hepatoprotective effect of aqueous extract cardamom against gentamicin induced hepatic damage in rats. Int J Basic Appl Sci. 2016; 5: 1-4.
- 22. Aly HA, Hassan MH. Potential testicular toxicity of gentamicin in adult rats. Biochem Biop Res Comm. 2018; 497: 362-7.
- 23. Serges KD, Laure PK, Legentil NM, Norbert K, Albert K, SL WN. Hepatoprotective and antioxidant effects of CommelinadiffusaBurm extracts on gentamicin-induced liver damage in rats. J Pharm Biol Sci. 2020; 8: 23-5.
- 24. Chandel SS, Sahu RK. Protective effect of dietary inclusion of Aegle marmelos fruit on gentamicin induced hepatotoxicity in rats. Intl J Green Pharm. 2017; 11: 597-603.