ANTIMICROBIAL SUSCEPTIBILITY OF SOME NATURAL DYES

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خلاصه

اس موجودہ تحقیقی کام میں پانچ مختلف پودوں سے بیکٹیر یائی مخالف طاقت کا مطالعہ کیا گیا جس میں غیر نا میاتی محلل (پانی) اور نا میاتی محلل (ایسی ثون اور بیگر ین) شامل ہیں ۔ ان تما م پودوں سے مخالف بیکٹیر یائی کار کردگی کی جانچ سے لئے آگر وال نفوذ ی طریقہ استعال کیا گیا ۔ جس سے بہترین نتیجہ حاصل ہوا ۔ جبکہ گرام شبت اور گرام منفی بیکٹیر یا میں نیر پل کوتھی کا عرق بیکٹیر یا سے اکا ڈ سے لئے نا موافق کا بت ہوا۔ اس سے علاوہ نارنگی سے حصلے ہوا ۔ جبکہ گرام شبت اور گرام منفی بیکٹیر یا میں نیر پل کوتھی کا عرق بیکٹیر یا ہے اکا ڈ سے لئے نا موافق کا بت ہوا۔ اس سے علاوہ نارنگی سے حصلے ہوند کی سے چوں اور دار چینی سے عرق میں تو نال کوتھی کا کر والے زہر ہے بیکٹیر یا سے خلاف بہتر کار کردگی ظاہر ہوتی ۔ محتلف پو دوں میں ضروری تیل جیس کی سالد پی موجود پی محلس اور ایس اور لیس میں فلیو ینوا ئیڈ کی موجود گی لوہ و نیا نرش میں میٹا ایس ورز کی میکٹور کی موجود گی جبکہ مرس میں ایل پر والی موجود کی محلس اور حول کار ایس وی میں محلود ہو ہو تیا نرش میں میٹا ایسوروز کی میکٹو اکھلو کی موجود گی جبکہ مرس میں ایل پر والی موجود حول کی کار پس جبکہ دار چینی موجود گی اور و نیا نرش میں میٹان محلو کی موجود کی جبکہ مرس میں ایل پر واجو کی محل اور خالف کر ہو جبکہ دار چینی میں بین سال ایلڈ کی بائیڈ اور ایم اینڈ دیکا میں میٹنی فیرن کی موجود گی جبکہ مرس میں اور

Abstract

Antibacterial potency of aqueous and non aqueous extract of five plants were studied in vitro. Methanol, Acetone and Hexane were used as a non aqueous solvents for the extraction of dyes. The agar well diffusion method were used during the entire study for the evaluation of antibacterial activities of different plants. The remarkable results were obtained from aqueous as well as non aqueous extracts against gram positive and against gram negative bacteria respectively. Furthermore, among them the aqueous and non aqueous purple cabbage extract exhibit most inhibitory effect on bacteria. It suppressed the bacterial growth by effecting the metabolite of certain bacteria. Orange peel, Henna leaf, Mango Bark and Cinnamon extract showed antibacterial activities against 1 to 4 pathogen as well as food poising bacteria. The growth inhibition effect of methanolic cinnamon extract against *M. luteus*, *B. subtilis S. aureus* almost equal about 12mm. The presence of flavonoids in *Brassicaoleracea var. capitata f. rubra*, β -asarones in *Lawsoniainermis*, nutraceuticals in albedo and flevedo of pericarp of Citrus fruits, Cinnmaldehyde in *CinnamomumVerum*, essential oils in different plant and mangiferin in *M.indica*, make them to survived against pathogenic bacteria.

Keywords: Gram Positive bacteria, Gram Negative bacteria, Essential oils, albedo, flevedo,

Introduction

Now a days, finding the methods that free the textile industries from pathogenic micro-organisms is a challenge. In modern world the upsurge in advanced technology demanding functionality in products like water repelling, fade resistance and resistance to microbial invasion (Kamel *et al.*, 2015). Consumer worldwide looking for the food remain fresh and odour free able to be use. This is all beholden to protect the materials from microorganisms. The plant is one major source of food. Majority of plants contain aromatic compounds. Aromatic plants constitute large part of natural flora and different part of plant materials used in various field such as pharmaceutical, flavour, fragrance, perfumery and cosmetic industries (Swany *et al.*, 2016). Moreover these plant are also the source of natural dyes. Dyes obtained from nature are safe and eco-friendly but mostly unstable and oxidize easily, resulting discoloration (Chao *et al.*, 2017) and susceptible to micro organisms. Many natural dyes have medicinal properties (Gupta *et al.*, 2006). Natural dyes are used as a textile material which are sensitize to microbial attack (Singh *et al.*, 2004). Due to physical properties of natural dyes such as Large surface area and absorption of moisture make natural dyes hypersensitive for the microbial onslaught. Literature review revealed that the massive work was done on antimicrobial properties of natural as well as synthetic dyes. Similitude between antibacterial activities of some natural dyes and silver nano particles were

synthetic dyes. Similitude between antibacterial activities of some natural dyes and silver nano particles were studied by (Mirjalili and Abbasipour, 2013). Synthesis, fastness properties, colour assessment and antimicrobial activity of some azo reactive dyes heaving pyrazole moiety was researched by (Rizk *et al.*, 2015). Dyeing phenomenon was carried out on acrylic fiber through antimicrobial cationic dyes (Ma *et al.*, 2005). The antimicrobial properties of photosensitizer cyanine dyes were examined by (Shindy *et al.*, 2006). Antimicrobial activities of synthesized monomethinecyanin dyes were researched out by (Abd-El-Aal, 2004). Antimicrobial

potency and absorption of some novel hetrocyclicdisazo dyes were studied by researcher team of (Karchi *et al.*, 2009).

The main interest of this work was to analyse the antimicrobial characteristics of natural dyes. The study on susceptibility of natural dyes towards microorganisms give broad spectrum information about the sustainability toward microbial attack as well as stability of natural dyes. The information from antimicrobial surveillance studies can be use to establish a trend in pathogen antimicrobial resistance and emerging pathogens at the national and global level (Masterton, 2008). The information about development of targeted approaches of microorganisms enable us to control the microorganisms. By controlling microorganisms in dyes the shelf life of dye in foods, dyeing and cosmetics increase because dyes are in several ways.

Materials and Methods

Plant extraction preparation

Plant material of five plants species included in this study presented in (**Table 1**) were collected from different local market of Karachi, Pakistan. These plants were watery washed, disinfected and rinsed by distilled water. Plant materials were dried and grounded in fine powder. After washing and converting into powder form the dyes were extracted in different solvents (**Table 2**) by using soaking method of extraction.

S.No	Plant	Family	Local Name	Plant part used	
1	M.indica	Anacardiaceae	Mango	Fruit	
2	CinnamomumVerum	Lauraceae	Cinnamon	Bark	
3	Lawsoniainermis	Lythraceae	Henna	Leaf	
	Brassica oleracea var.		Purple		
4	capitata f. rubra	Brassicaceae	Cabbage	Vegetable	
5	Citrus X sinensis	Rutaceae	Orange	Fruit peel	

Table 1: Plant Species with their Specific parts were used in study

	Table 2 Non aqueous solvent for extraction of uyes					
S.No	Samples	Solvents				
		Water				
1	Mango Bark	Methanol				
-		Acetone				
	Cinnamon Bark	Water				
2		Methanol				
		Hexane				
	Orange Peel	Water				
3		Methanol				
4	Henna	Water				
		Water				
5	Purple Cabbage	Acid				
		Methanol				

Table 2:- Non aqueous solvent for extraction of dyes

Antimicrobial activity of plant extract

Bacterial Strain: The antibacterial potency of dyes examined by using seven bacterial species which have ability to food poising and life threatening diseases in human beings. Three species of Gram Positive bacteria (Micrococcus luteus, Staphylococcus aureus and Bacillus subtilis) and five species of Gram negative bacteria (Escherichia coli,Pseudomonas aeruginosa, Klebsiella pneumoniae, *Proteus mirabilis* and Salmonella typhi) were used to study the antibacterial activity of natural dyes in presented work.

Inoculum Method :The 24h fresh Cultures were used. Fresh culture transferred aseptically into sterile saline. Then vortex the saline tube and measured with McFarland standard no. **0.5**, turbidity which adjusted to give 1.5×10^8 CFU/mL count.

Antibacterial activity of plant extract: The agar well diffusion method was used for evaluation of antimicrobial activity of each plant extract. The inoculum of culture was spread into surface of nutrient agar plates and then made a 100μ L hole by using cork borer. Sample of interest transferred with the help of 100μ L juster pipette aseptically. Then nutrient plates were incubated at 37° C for 24h.

Results and discussion

Antibacterial Activities of Aqueous extract

Potency of aqueous dyes against the gram positive as well as gram negative bacteria species were depicted in Table 3 and Table 4 respectively. It was observed that henna leaf dye was susceptible to Staphylococcus aureus. Staphylococcus aureus cause severe disease in human such as bacteremia, infective endocarditis, skin and soft tissue infections (Tracey et al., 2018). It was notice that Henna leaf showed resistance against two species of bacteria that were Micrococcus luteus and Bacillus subtilis. The potency of henna leaf extract due to presence of β -asarones active ingredient present in leaf, root and rizhomes tissues active against bacteria (Kannah and Vinotha, 2013). While extract of orange peel showed resistance against Bacillus subtilis. This was due to presence of magnesium, Calcium, Copper, Vitamin A, folate and phytochemicals in orange pericarp which make it defy towards gram positive bacteria. There was no zone of growth inhibition observed by orange peel extract against Micrococcus luteusand Staphylococcus aureus. The antimicrobial activity of samples against mentioned pathogens were due to presence of tennins, Saponins, phenolic compounds, essential oil and flavonoids (Yerou et al., 2017). Flavonoids, phenolic compounds, phytochemicals and essential oils are bioactive ingredient of any fruit, leaf and vegetable which have antimicrobial properties. Antibacterial potency of henna and orange peel dyes were negative against three gram negative bacteria that were Proteus mirabilis, Salmonella typhiand Pseudomonas aeruginosa. Pseudomonas aeruginosaishuman pathogen which cause life threatening acute infection and chronic airway colonization during cystic fibrosis. *P.aeruginosa* form a bio-film characterized by secretion of extra-cellular matrix of various poly-saccharides and grow in number of genes involved in surface adherence (Eckert et al., 2016). Cell related to bio-film less active metabolically than their planktonics counterparts and especially resist to small molecule antibiotics as a result of this, potency of extracts against P.aeruginosa was negative.

It was also ascertained the mango bark dye was showed zone of growth inhibition against *Bacillus subtilis* and *Staphylococcus aureus*. It was due to mangiferin. Mangiferin(1,3,6,7-tetrahydroxyxnthone-2-C- β -D-glucopyranoside), a C-glucosylxanthone was isolated from mango bark (Castellanos *et al.*, 2014) which is antibacterial agent (Hannan *et al.*, 2013) and showed their potency against gram positive bacteria's. Cinnamon bark dye appeared their antibacterial activity against only Micrococcus luteus. there was no zone of growth of inhibition observed for gram negative bacteria's. In the studied of antibacterial activity of cinnamon aqueous extract studied by researchers showed antibacterial activity against *E.coli* (Puangpronpitag and Sittiwet, 2009).Cinnamaldehyde and eugnol bio-active compounds showed antibacterial activity (Nabavi *et al.*, 2015) against Micrococcus luteus. The purple cabbage showed zone of growth inhibition against three gram positive and one gram negative bacteria which where depicted in table 3 and 4 respectively. The presence of phytochemical in brassica oleracea var. capitata f. rubra were responsible for their antimicrobial potency.

S.NO	Samples	Zone of Growth Inhibition (mm)				
		M. luteus	B. subtilis	S. aureus		
1	Henna leaf	20mm	10mm	-tive		
2	Orange Peel	-tive	20mm	-tive		
3	Mango Bark	-tive	11mm	10mm		
4	Cinnamon	10mm	-tive	-tive		
5	Purple Cabbage	12mm	11mm	10mm		

Table 3: Antibacterial activity of aqueous extracts against Gram Positive bacteria's

Key: -tive means no zone of growth of inhibition



Fig (1A): Antibacterial activity of different compounds against *Micrococcus luteus*, (1B) Antibacterial activity of different compounds against *Bacillus subtilis*, (1C) Antibacterial activity of different compounds against *Staphylococcus aureus*



Fig 02: Antibacterial activity of different compounds againstProteus mirabilis

S.NO	Sample	Zone of Growth Inhibition (mm)					
		P.mirabilis	S. typhi	P. aeruginosa	E.coli	K.pneumoniae	
1	Henna	-tive	-tive	-tive	-	-	
2	Orange Peel	-tive	-tive	-tive	-	-	
3	Mango Bark	-tive	-tive	-tive	-	-	
4	Cinnamon	-tive	-tive	-tive	-	-	
5	Purple Cabbage	-	-tive	-tive	8mm	-tive	

Table 4: Antibacterial activities of aqueous extracts against Gram negative bacteria's

Key: -tive means no zone of growth inhibition

Key: - means sample not studied against these bacterial strains

Antibacterial activity of non aqueous extract: By using the non aqueous extraction the antibacterial potency of extracts were also studied presented in (Table 5). It was ascertained that the methanolic extract of orange peel exhibit antibacterial activity against *M. luteus* and *B. subtilis*. The zone of growth inhibition in *M. luteus* was 10mm than *B. subtilis* that was 9mm. The orange is citrus fruit. The pericarp of orange consist bio-active compounds, nutraceuticals and functional compounds in albedo and flavedo of peel (Yashaswini and Arvind, 2018). In orange the flavonoids are present as flavanones (neohesperidosides, rutinosoides), flvanol glycoside, flavones (polymethoxyflavones, hydroxylated polymethoxyflavones) with frequent bio active compounds like

naringin and hesperidin. Phenolic compound have antibacterial as well as antiviral activities in orange. The citrus fruit also contain the essential oils (EO_s) that were also responsible for their antibacterial activities. It was also observed three strains of gram negative bacteria survived against methanolic extract of orange peel was depicted in (Table 6). The other authors also studied the antibacterial activities of orange EO_s (Espina *et al.*, 2011) which ratified our results.

Methanolic extract of mango bark also resist against *B. subtilis.* While there was no zone of growth inhibition observed against gram negative bacteria. The bark of mango consist Tannins, Saponins, Phenol, Glycogen, Alkaloids and Cardiac glycoside (Abubakar, 2009). These are bio-active bio compounds which protect the plant against pathogens and predation by animals. The *Micrococcusluteus* and *Staphylococcus aureus* showed resistance against the methanolic and acetone mango bark extract. The difference in activities observed with the variation in solvents. This associated with presence of oil, fatty acids, wax, resins, and pigments which capable to blocked the active ingredient in the extract, preventing the bark extract from reaching the bacterial cell wall (Ashok *et al.*, 2014). The *Micrococcusluteus Staphylococcus aureus* showed resistance against the methanolic and acetone mango bark extract.

The methanolic extract of Cinnamon showed resistance against three gram positive and one gram negative bacteria. The methanolic extract clearly indicate the antibacterial characteristic of cinnamon bark. There is a presence of tennin which has phenol group have a properties like alcohol is an antiseptic that use as antimicrobial component (Suliman *et al.*, 2017). Methanolic extract showed almost equal antibacterial activity against *M. luteus*, *B. subtilis*and *S. aureus*. 10mm length of growth inhibition observed against *P. mirabilis*. The cinnamaldehyde in cinnamon proven most active against fungi, virus and bacteria (Ali *et al.*, 2005). The cinnamaldehyde in cinnamon resits against *S. aureus* (Aneja *et al.*, 2009). It was among the most potent chemical against gram positive bacteria. Hexane extract of Cinnamon was capability to resist against only *B. subtilis* while the antibacterial activity of hexane extract of cinnamon against gram negative bacteria was negative.

The acidic extract of purple cabbage resist againt all gram positive and gram negative bacteria's except *S.typhi*which resist against acidic extract. The acid break the cell wall of gram positive as well as gram negative bacteria's.

S.NO	Solvents	Samples	Zone of Growth Inhibition (mm)			
5.10	Solvenis		M. luteus	B. subtilis	S. aureus	
1		Orange Peel	10mm	9mm	-tive	
2	Methanol	Mango Bark	-tive	10mm	-tive	
3	memunoi	Cinnamon	12mm	12mm	12mm	
4	Hexane	Cinnamon	-tive	10mm	-tive	
5	Acetone	Mango Bark	-tive	12mm	-tive	
6	Acid	Purple	15mm	18mm	18mm	

Table 5: Antibacterial activities of non aqueous samples against Gram positive bacteria

Key: -tive means no zone of growth of inhibition

Table 6: Antibacterial activities of non aq	ueous extracts against	Gram negative bacteria

	Solvent	Sample	Zone of Growth Inhibition (mm)				
S.No			P. mirabilis	S.typhi	P. aeruginosa	E.coli	K. Pneumonia
1		Orange Peel	-tive	-type	-tive	-	-
2	Methano	Mango Bark	-tive	-tive	-tive	-	-
3		Cinnamon	10mm	-tive	-tive	-	-
4	Hexane	Cinnamon	-tive	-tive	-tive	-	-
5	Acetone	Mango Bark	9mm	-tive	-tive	-	-
6	Acid	Purple Cabbage	-	-tive	20mm	18mm	17mm

Key: -tive means no zone of growth inhibition

Key: - means sample not studied against these bacterial strains

It was concluded Thatthe aqueous extract exhibit great potential against gram positive bacteria than non aqueous extract. The potency of non aqueous extracts against gram negative bacteria was greater than aqueous extracts. Mango bark non aqueous extract showed resistance against *P.mirabilis*. More over the Cinnamon also exhibited antibacterial activity against *P.mirabilis*. Aqueous extract of purple cabbage resist against all gram positive bacteria which were under studied. While the potency of purple cabbage aqueous extract against *E.coli* was 8mm suggest that the purple cabbage can be use in stomach relief pain drugs. The all non aqueous extracts which were under studied showed great antibacterial activity against *B. subtilis*. Further more the acid extract of purple cabbage possessed potency against all gram positive as well as all gram negative bacteria except *S.typhi*. This was notable that the *S.typhi* resist in acidic condition. The presence of flavonoids, essential oils, flavanones tennins and other compounds make fruits and vegetable to resist against wide range of microorganisms.

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References

- Abd-El-Aal., (2004). Synthesis and antimicrobial activity of certain noval monomethin Cyanin dyes.J. Dyes and Pigment.60(3):205-214.
- Castellanos, E., Gonzalez, R., Nuevas, L. and Hoogmart, J. (2014). Determination of MangiferinIndicaL.Stem Bark extract (Vimang) and Pharmaceutical by liquid Chromatography .*J.Food.Agric* 26(7):592-601.
- Deepti,G., Sudhir, K. .K. and Ankhur. L. (2006). Antimicrobial properties of natural dyes against Gram negative bacteria, J. *Coloration and Technology*. 120(4):167-171.
- Eckert, R., Keith, M.B., Peter, E., Fengxia, Q., Dania, K., Jin, H., Lan.M., Anderson, H. and Wenyuan, S. (2006). Enhancement of antimicrobial activity against Pseudomonas aeruginosa by Co administration of GloKHc and tobramycin. *J. antimicrobial agent and chemotherapy*. 50(11):3833-3838.
- Espina, L., Somolinos, M., Loran, S., Conchello, P., Garcia, D. and Pagan, R. (2011). Chemical Composition of Commercial Citrus Fruits essential oils and evaluation of their antimicrobial activity acting alone or in Combined process. *J.FoodControl.* 22(6): 896-902.
- Fazal, S., Lorenzo, A., Izadi, M., Sobarzo-Sanchez, E., Daglia, M. and Muhammad, S. (2015). Antibacterial effect of Cinnamon : from Farm to food, Cosmetic and Pharmaceutical Industries. J. Nutrients.7: 7729-7748.
- Gomashe, V., Balge, S. and Gulhane, A. (2014). Evaluation of antibacterial and mangifera indica bark extract. J. *Curr.Microbiology.App.Sci.* 3(5): 567-580.
- Haman, A., Naeem, T., Ikam, M., Ahmad, I., Aneela,S. and Hussain, S. (2013). Antibacterial activity of Mango (Mangiferaindicalinn) leaf extract against antibiotic sensitive and multi Drug resistance Salmonella typhi. J.Pak.Pharm.Sci. 26(4): 715-719.
- Kamel, M. M., Helmy, H.M., Meshaly, H.M. and Abou-okeil, A. (2015). Antibacterial activity of cationised cotton dyed with some natural dyes, *J.textile Science and Engineering*. 5(1):1-5.
- Kannahi, M. and Vinotha, K. (2013). Antimicrobial Activity of Lawsoniainermis leaf extracts against some human pathogens. J.Curr.Microbial.App.Sci. 2(5):342-349.
- Kret, K. F., Nesrin, S., Mustafa, Y., Izzet, S. and Aykut, D. (2009). The Synthesis, Antimicrobial activity and absorption characteristics of some novalHetrocyclic Diazo Dyes. J. Dyes and Pigment. 80(1): 47-52.
- Mahaboob,S., Aleemkhan, A., Ahed, I., Musaddiq, M., Khaja, S., Polasa, H., Venkate, L., Habibullah, M., Leonardo, A. and Ahmad, N. (2005). AntibacterialActivity of eugenol and Cinnamaldehyde against human gastric Pathogen Helicobacter Pylori, *J.Annals of Clinical Microbiology and Anti-microbiology*. 4(20): 1-7.
- Mallappa, K.S., Mohd, S.A. and Uma, R.S. (2016). Antimicrobial properties of plant essential oil against human pathogens and their mode of action, *J. Evidence based Complementary and Alternative Medicines*. 2016:1-21.
- Masterten, R. (2008). The importance and Future of Antimicrobial Surveillance Studies. J. Clinical Infectious Disease. 47(51):521-531.
- Meddeh, B. and Bouchadi, D. (2017). The use of Orange peel as antimicrobial and anti oxidantagent. *J.Fundamental and Applied Science*. 9(3):1351-1357.
- Mehmood, E. (2009). Antimicrobial efficiency of stem bark extracts of Mangoferaindica against some bacteria associated with respiratory tract infections. *J.Scientific research and Essay.* 4(10): 1031-1037.

- Minghua, M. and Gang. S. (2005).antibacterial cationic dyes part 3 simultaneous dyeing and antimicrobial finishing of acrylic fiber. *J. dyes and pigment*. 66(1):33-41.
- Mirjalili, M. and Abbasipour. M. (2013). Comparison between antibacterial activity of some natural dyes and sliver nano particles, *J. Nano Structure in Chemistry*.3(37):1-3.
- Puangpronpitag, D. and Sittiwet, K. (2009). Antimicrobial Properties of Cinnamomum Verum aqueous extract. J. Biological Science. 2(2):49-53.
- Rai, K., Joshi, R. and Sharma, C. (2009). Antimicrobial activity of Dalchini (C.Zeylanicum bark) extract on some dental Caries Pathpgens. J. Pharmacy Research. 2(9): 1387-1390.
- Rajni, S., Astha, J., Shikapanwar, D. G. and Khare, S.K. (2004). Antimicrobial activity of some natural dyes, J. *Dyes and Pigments*. 66(2):99-102.
- Rizk, H. F., Ibrahim, S.A. and Elborni, M.A. (2015). Synthesis, Fastness properties, Colour assessment and antimicrobial activity of some azo reactive dyes heaving Pyrazole moiety, *J.Dye and Pigment*.112:86-92.
- Saad, R., Ali, H., Nurulam, I., Nurshamiha, N., Nazim, M., Budiash, S., Suliman, R. and Gebaly, A. (2017). Cinnamon bark extract for formulation nd Characterization of antimicrobial Cream. J. Ayurveda Pharm.8(2): 1-7.
- Shindy, H., Elmaghraby, M, and Eissa, F. (2006). Synthesis, Photosensitization and antimicrobial activity of OxidiazineCyanin Dyes. J. Dyes and Pigment, 70(2): 110-116.
- Taylor, T.A. and Unakal, C.G. (2019). Staphylococcus Aurus, State Pearl Publishing.
- Yashaswiniard, P. and Arvind, (2018). Antimicrobial properties of orange (Citrus Reticulata.Kinnow) Peel extract against pathogenic bacteria. *J.*.*App.Sci. Curr.Microbiol.* 7(3): 737-746.
- Yu, C. C. Tsung-hun, H. Jiao, .Z. hengKao, L. and Szu, T. P. (2017). A study on combining natural dyes and environmentally friendly mordant to improve colour strength and ultra violate protection of textile, *J. Fiber* and Polymers.18(8): 1523-1530.