

ESTIMATION OF ALKALINE PHOSPHATASE AND PROTEIN CONTENT IN DIFFERENT SPECIES OF CESTODE PARASITES OF DOMESTIC PIGEONS (*COLUMBA LIVIA*)(GMELIN,1789)

SHAZIA NISAR¹, MUHAMMAD ZAHID, SYED IKHLAQ HUSSAIN, AASIA KARIM AND UZMA MEHBOOB

¹Department of Zoology, Federal Urdu University of Arts, Science and Technology
Corresponding Author's e-mail : shaz_noman@live.com

خلاصہ

گھریلو کبوتر (*Columba Livia*) میں پائے جانے والے فیٹے (Cestodes) *Cotugnia streptopell*, *Raillietina galeritae* اور *Raillietina flaccida* میں الکلائن فاسفاٹیز اور پروٹین مندرجات (Protein Content) کی مقدار کا مشاہدہ کیا گیا۔ الکلائن فاسفاٹیز کی اوسط مقدار کا تخمینہ *R.flaccida* اور *R.galeritae*, *C.streptopell* میں بالترتیب 19.228، 19.504، 22.908 U/l جبکہ پروٹین مندرجات کی اوسط مقدار کا تخمینہ 0.950، 2.677، 4.117 g/dl بالترتیب لگایا گیا۔ الکلائن فاسفاٹیز کی زیادہ سے زیادہ اوسط مقدار *C.Streptopell* میں جبکہ کم ترین مقدار *R.flaccida* میں دیکھا گیا۔ پروٹین مندرجات (Protein Content) کا تخمینہ اس کے بالکل برعکس حاصل ہوا۔ زیادہ سے زیادہ پروٹین مندرجات *R.flaccida* میں اور کم سے کم *C.Streptopell* میں لگایا گیا۔

Abstract

Level of alkaline phosphatase and protein content were observed in *Cotugnia Streptopell*, *Raillietina galeritae* and *Raillietina flaccida* in whole cestode from domestic pigeons (*Columba livia*). Mean values of alkaline phosphatase in *C. streptopell*, *R.galeritae* and *R. flaccida* were observed as 22.908, 19.504, 19.228 U/l respectively, while mean values of protein content were observed as 0.950, 2.677, 4.117g/dl respectively. The maximum values of alkaline phosphatase was obtained in *C. streptopell* while *R. flaccid* showed lowest mean values. The reverse result was observed in protein content estimation. The highest protein content was observed in *R. flaccida* while reduced protein content was observed in *C. streptopell*.

Key words: Cestodes, Pigeon, Alkaline phosphatase, protein content

Introduction

Biochemistry of parasites has great importance and some work has been done on helminth's physiology. Maximum parasites exist in connotation of animals, birds, and fishes of economic significance. Parasitic biochemistry has abundant practical significance through chemotherapy and vaccine manufacture and in considerate of the multifarious link involved in the host parasite relationship with helminth parasites enzyme (Alkaline phosphatase) is significant enzyme that are originated to be allied with consumption of nutriment ingredients. Some intervention to this enzyme might cause decrease and paralysis of the worms (Swargiary *et al.*, 2013). The enzyme is established in the extracellular liquids and cells of a comprehensive variety of worms with helminths. (Kar and Tendon 2004) made histochemical and biochemical observations which confirmed the occurrence of alkaline phosphatase in adjacent relations with somatic musculature, sub-tegment, tegument, cuticle and gut of numerous helminths, mainly in the adhesive structures like oral suckers and acetabulum. The farthest body layer of trematode and cestodes Tegument parasites through which absorption and digestion of several food stuff takes place (Roy 1982). In cestodes, alkaline phosphatase is present in walls of excretory ducts and subcuticula (Bogitsh 1963). Protein content of the cuticles of cestodes is synthesized in subcuticular matrix (Lumsden 1966). (Rogers 1969) studied on the metabolism of protein, amino acids and other nitrogen compounds in nematode. Protein and enzyme activity differs in different parasitic species due to capability of absorption of food from host and intestinal environment (Akhter *et al.*, 2006). Diet protein effects the growth of parasites and involve in protein metabolism of cestodes. (Mettrick and Munro 1965) observed free nutritional amino acid supplements effects on the *Hymenolepis diminuta* growth.

Materials and Methods

Cotugnia Streptopell, *Raillietina galeritae* and *Raillietina flaccid* of different size were obtained from intestine of infected pigeons. Level of alkaline phosphatase and protein content estimated in these worms. 2g samples were crushed in 2ml of distilled water and homogenized in Teflon Pyrex soft tissue grinder for 5 min at 1000 rpm. The homogenates were centrifuged at 3500 rpm in Labofuge 15000 for 15 min. Supernatants were used for biochemical analysis.

Estimation of Alkaline phosphatase (ALP): Alkaline phosphatase was determined by the colorimetric kit method of Randox cat No. 307. One test tube was taken and add 0.02 ml of homogenate/supernatant and then 1ml of substrate was added and mixed. Reading was taken after 1, 2 and 3 minutes against the air blank at 405nm on Shimadzu spectrophotometer UV-120. Finally these values were kept in the following formula to calculate the alkaline phosphatase activity.

$$U/l = 2760 \times \Delta A_{405nm/min}$$

Estimation of Total Protein: Total protein was estimated by Biuret method using Randox diagnostic kit Cat No. TP 245. Three test tubes were taken and marked as sample, standard and reagent blank. 1 ml of solution was added to each test tube then 0.02 ml of supernatant was added to their respective test tube and 0.02 ml of distilled water to the reagent blank's test tube and 0.02 ml standard in respective test tube. Test Tubes were incubated for 30 min. in water bath at 25 °C. Absorbance of sample was measured against reagent blank at 546 nm on shimadzu spectrophotometer U V-120.

For the estimation of total protein concentration, following formula was used.

$$\text{Total protein concentration} = 190 \times A_{\text{sample}}(g/l)$$

Results and Discussion

Biochemical estimation in cestode parasites are shown in Table.1 and Table.2 Estimation of alkaline phosphatase and protein content were observed in *C. Streptopell*, *R.galeritae* and *Raillietina flaccida* in whole body. Mean values of alkaline phosphatase in *C.streptopell*, *R.galeritae* and *R.flaccida* were observed as 22.908, 19.504, 19.228U/l respectively. The maximum value of was obtained in *C. Streptopell* while *R.flaccida* and *R.galeritae* showed very close levels of alkaline phosphatase (Table 1).

Table 1. Level of Alkaline Phosphatase (ALP) in different cestode parasites.

	Mean (U/l)	S.D	S.E	Range(U/l)
<i>C.streptopell</i>	22.908	0.730	0.422	22.082-23.734
<i>R.galeritae</i>	19.504	0.575	0.332	18.854-20.154
<i>R.flaccida</i>	19.228	1.149	0.663	17.928-20.528

Mean values of protein content in *C.streptopell*, *R.galeritae* and *R.flaccida* were observed as 0.950, 2.677 and 4.117 g/dl respectively. The maximum value of protein content was obtained in *R.flaccida* while *C. Streptopell* showed lowest mean value of protein contents (Table 2).

Table 2. Level of protein content in different cestode parasites.

	Mean(g/dl)	S.D	S.E	Range(g/dl)
<i>C.streptopell</i>	0.950	0.380	0.219	0.520-1.380
<i>R.galeritae</i>	2.677	0.366	0.211	2.263-3.090
<i>R.flaccida</i>	4.117	0.611	0.353	3.426-4.808

Biochemical estimation in cestode parasites i.e *C.streptopell*, *R. galeritae* and *R. flaccida* are expressed in terms of unites. *Swargiary et al.*, (2013) observed alkaline phosphatase's characterization in an intestinal fluke *fasiolopsis buski* treated with alpinianigra's crude extract Temperature, pH and incubation time effect alkaline phosphatase activity. Enzyme activity showed negative effect with increasing pH. By the temperature fluctuation activity of enzyme can be controlled. A huge quantity of chemicals have been seen to modify alkaline phosphatase activities. According to Mahanty *et al.*, (2011) in the culture medium alkaline phosphates secretion found to be reduced when albendazole and PZQ was used to treat *Taeniasolium* Cysts. Pappas 1991 observed divalent cations was used to check inhibition and activation brush-border membrane-bound alkaline phosphatase activity on *Hymenolepis diminuta* (Cestoda). Dusanic (1959) demonstrated the alkaline phosphatase histochemical localization in different stages of parasitic trematode, *Schistosomamansonii* too observed significant

difference from stage to stage. Krasnosnoschchekov and Tomilovskala (1975) observed distribution of alkaline phosphatase by means of the incubation of whole cestode. Alkaline phosphatase was associated with calcareous corpuscles, integumental tissue and excretory system.

Rothman (1966) observed alkaline phosphatase in the cestode *Hymenolepis citelli*, on outer membrane of the proximal microthrix. In present investigation highest level of alkaline phosphatase observed in *C.streptopell* and almost same level of alkaline phosphatase observed in *R. galeritae* and *R. flaccida*. Many factors like absorption of nutrients, pH and temperature effect the level of alkaline phosphatase in different cestodes. Proteins consume various altered biological functions. They are ubiquitous in their distribution and there is certainly no suitable outline of categorizing them. The principal group of proteins are the enzyme proteins that offer ridiculous setting for the nourishment of cestodes parasites (Sonune 2012). The cestodes consume diverse grades of protein for generating energy. Literature has exposed that the parasites are capable of adopting themselves to the parasitic manner of life, only due to protein. In helminths parasites, the protein generally present between 20 – 40 % of the dry weight (Sharma, 1979) but protein content value, as high as 70% of the aridmass have been observed for *Nippostrongylus brasiliensis* (infective larvae) and *Macrachanthorhynchushirudinaceus* (Barrett, 1981). Jadhav *et al.* (2008) reported biochemical contents from Davaineashindei. Amount of protein present in Davaineashindei 13.20 mg/gm wt. of tissue. Bhure *et al.* (2016) studied protein's amount present in nematode parasites is 24.80 mg / 100mg collected from Wallagoattu. Bhure *et al.*, (2012) observed reduced amount of protein (15.88 mg/gm) in Ascardiagalli as associated to healthy intestine (19.77 mg/gm) and diseased host intestine (19.33 mg/gm). Pallewad *et al.* (2015) reported Proteinin Cotylophoron cotylophorum 23.60 mg / 100 mg in Capra hircus. Nanware *et al.* (2012) observed protein's amount present in Cestode Cotugnia sp. is 5.77mg/gm. Dhondge *et al.* (2010) reported amount of Protein was lower in the body of the parasites than infected and normal intestinal tissue of host. In present study showed that highest protein content found in *R. flaccida*. Pathan *et al.*, 2011 were observed worms were skilled of extracting nutritious material from their hosts and thus characterized elevated level in protein.

Conclusion

Different species have different protein content depend on parasites capability to absorb protein from nourishment of host. In present study the highest protein content was found in *R. flaccid*. Hence there is relationship between the protein content of parasites and nutrient protein content of host. Parasites depend on protein consumptions from host for energy. Many factors can effect the level of alkaline phosphatase in parasites.

References

- Akhter, K., Naqvi, S. H. N., Azmi, M. A., Tariq, R. M. and Btool, F. (2006). Study of glucose and protein activity in some common cestodes and nematodes of *Columba livia*. *Pakistanj.entomol*, 21: 19-21.
- Barrett, J., 1981. *Biochemistry of parasitic helminths* pp. 308. Macmillan pub. Ltd., London.
- Bogitsh, B. J. (1963). Histochemical studies on *Hymenolepismicrostoma* (cestoda *Hymenolepididae*.) *The journal for parasitology*, 49 (6): 989 – 997.
- Bhure, D. B., Nanware, S. S., Kardile, S. P. and Barshe, M. U. (2016). Taxonomic and Biochemical Studies of Piscean Nematode CamallanusJadhavii (Jadhav and Khadap, 2013) Parasitic in Wallago Attu (Bleeker, 1857). *World Scientific News* 34: 98-108.
- Bhure DB, KadamNima, Nanware SS. and Garad VB. (2012). Studies on protein profile of Ascardiagalli and its host Gallus gallusdomesticus. *International Multidisciplinary Research Journal* 2(6): 60-61.
- Dhondge, R. M., Nanware, S.S., Bhure, D. B. and Kadam, M. S. (2010). Protein profile of avainCestodes - A case Study. *The Biosphere An International Journal of Life Sciences*. 2(2): 133-136.
- Dusanic, D. G. (1959). Histochemical observations of alkaline phosphatase in *Schistosomamansoni*. *J Infect Dis*, 105: 1–8.
- Jadhav, B., Shivesh, V., Singh, P., Bhure, D. B. and Padwal, N. D. (2008). Biosystematic studies of Davaineashindei.sp. (Cestoda- Davainidae) Fuhrmann, 1907 from Gallus gallusdomesticus. *National Academy of Science Letter* Vol.-31 No.7 & 8 pp 245-250.
- Kar, PK. and Tandon, V. (2004) Anthelmintic efficacy of genistein, the active principle of *Flemingiavestita* (Fabaceae): Alterations in the activity of the enzymes associated with the tegumental and gastrodermal interfaces of the trematode, *Fasciolopsisbuski*. *Journal of Parasitic Diseases*, 28:45-56.
- Krasnosnoschchekov, G. P. and Tomilovskala, N. S. (1975). Distribution of certain enzymes in totally stained cestode preparations. *Parazitologiya*, 9 (3): 227-31.
- Lumsden, R. D (1966). Cytological studies on absorptive surfaces of cestodes. *Parasitology Research*, 28 (1): 1-13.

- Mahanty, S., Paredes, A., Marzal, M., Gonzalez, E., Rodriguez, S., Dorny, P., Guerra-Giraldez, C., Garcia, HH. and Nash, T. (2011). Sensitive invitro system to assess morphological and biochemical effects of praziquantel and al-bendazole on *Taeniasolium* cysts. And *microbial Agents and Chemo-therapy*, 55:211-217.
- Mettrick, D. F. and Munro, H. N. (1965). Studies on the protein metabolism of cestodes. *Parasitology*, 55: 453-466.
- Nanware, S. S, Nazneen, U., Bhure, D. B. and Garad, V. B. (2012). Studies on protein content of cestode *Cotugnia* and its host *Gallus gallusdomesticus*. *Journal of Experimental Sciences*, 3(1): 40-41 ISSN: 2218-1768
- Pappas PW: (1991). Activation and inhibition of the brush border membrane bound alkaline phosphatase activity of *Hymenolepisdiminuta* Cestoda. *Parasitology*, 10:141-146.
- Pallewad. S., Nanware. S. and Bhure, D. B. (2015). Biochemical contents of *Cotylophoroncotylophorum* (Fischöeder, 1901) stiles et Goldberger, 1910 and its host intestinal tissue. *Biolife*. 3(1): 192-195.
- Pathan, D. M., Bhure, D. B. and Rajput, K.H. (2011). Redescription of *Camallanusjadhavii* from freshwater fish, Wallagaattu from Aurangabad. *Flora and Fauna An International Research Journal of Biological Sciences*. 17(2): 338-342
- Rothman, A. H. (1966). Ultrastructural studies of enzyme activity in the cestode cuticle. *Experimental parasitology*, 19 (3): 332-338.
- Rogers, W. P. (1969). Nitrogenous components and their metabolism *Acanthocephala* and *Nematoda*. *Chemical Zoology. New york and London: Academic Press Inc.*, 3: 379-428.
- Roy, T. K. (1982) Histochemicalstudies on *Raillietina (Rail-lietina) johri (Cestoda: Davaineidae)*. I. Nonspecific and specific phosphatases. *Journal of Helminthology* 53:45-49.
- Sharma, P. N. (1979). Histochemical localization of glycogen, lipids, protein and phosphatase in the parenchyma and other tissues of some digenetic trematodes. *Indian Journal of Experimental Biology*, 17, 479–483.
- Sonune, M. B. (2012). Biochemical studies of gastrointestinal cestode parasites in *OvisBharal* (L.) from Vidharbha region. *Bioscience Discovery* 3: 321-322.
- Swargiary. A., Roy, B. and Ronghang, B (2013). Partial characterisation of alkaline phosphatase in *Fasciolopsisbuski* an intestinal fluke treated with crude extract of *Alpinianigra* (Zingiberaceae). *Journal of Pharmaceutical Technology & Drug Research* 1-7.