# Evaluation of Antioxidant and Antidiabetic Activity of *Phyllanthus emblica* (Fruit)

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#### ABSTRACT

The objective of this study was to explore the scientific basis of traditional usage of *Phyllanthus emblica* as an anti-diabetic agent in hyperglycemic patients. Ethanolic extract of its dried fruits was analyzed both qualitatively and quantitatively for evaluation of phytochemical agents. Then its antioxidant and anti-diabetic activities were found. The results indicated the presence of carbohydrates, tannins, phenols, alkaloids, flavonoids, saponins, glycosides, amino acids and proteins in crude extract with phenolic and flavonoid contents observed as 9.9 mg/g equivalent of gallic acid and 100 mg/g equivalent of rutin, respectively. Free radical scavenging activity by plant extract was observed as  $68\pm0.33$  % as compared to rutin ( $58\pm1.15$  %) at concentration 2.56 mg/ml. Similarly, *in vivo* studies indicated that *Phyllanthus emblica* 80 mg/kg significantly reduced the glucose level in diabetic rats up to  $166 \pm 0.7$  mg/dl on day 8<sup>th</sup> at 8<sup>th</sup> h as compared to 310.71 ± 0.57 g which was recorded before treatment. It was thus concluded that *Phyllanthus emblica* is a potent antioxidant and antidiabetic agent.

Key words: Phyllanthus emblica, antidiabetic, Amla, diabetes mellitus, antioxidant

### INTRODUCTION

The prevalence of diabetes mellitus is increasing day by day which needs to be controlled on priority basis. Trend is shifting towards the use of herbal remedies so various plants (*Pterocarpus marsupium*, *Momordica charantia* and *Trigonella foenum-graecum* etc) have been scientifically studied for their anti-diabetic effect (Jung *et al.*, 2006).

Medicinal plants endure to be of an essential therapeutic benefitfor relieving disorders of mankind. From the previous 2500 years, a variety of traditional medical systems including Ayurvedic, Unani and Chinese are in practice. These traditional medicinal systems have been created and largely practiced in the eastern countries. On the whole, nearly 80% population of the developing countries largely depend upon these systems of traditional medicine for their basic health needs (Tsay & Agrawal, 2005).

Herbal preparations and plants are widely used for the treatment and management of diabetes mellitus all over the world instead of the availability of modern glucose lowering agents. The current research was performed to investigate the hypoglycaemic activity of the fruit extract of *Phyllanthus emblica* in insulin resistance diabetic rat models and the effect of plant extract on the fasting glucose level was also found.

*Phyllanthus emblica* (Euphorbiaceae) is commonly known as Amla and generally grows in subtropical as well as tropical areas in China, India, Asia, Indonesia and also on the Malay Peninsula. All parts of plant like roots, stem, leaves and fruits are used both in fresh and dry forms (Unander *et al.*, 1990).

It has been increasingly used in Ayurvedic system of medicines as a diuretic, laxative and coolant. In food stuffs, it is used to make jellies and pickles (Shankar et al., 1996). It possesses strong antioxidant activities (Naik et al.. 2005). Traditionally, it has been used as anti-inflammatory, antiulcer. anti-cancer. hepatoprotective and immune-modulating agent (Dang et al., 2011).

Chemical studies have shown that Amla is enriched in vitamin-C (Nisha *et al.*, 2004) and gallic acid. It also contains ellagic acid, gallic acid, corilagin, furosin, flavonoids, glycosides and tannins which are responsible for anti-inflammatory, antimicrobial, anti-ulcer, nephroprotective, analgesic and antipyretic, anti-tussive, and immune modulating activities (Dang *et al.*, 2011; Kumaran & Karunakaran, 2006; Zhang *et al.*, 2001).

Author's Contribution: A.B., Did experimental work; A.M., Designed and supervised the study; T.M., Compiled the data and assisted in manuscript writing

### MATERIALS AND METHODS

#### **Plant Material and Extraction**

The Amla (Phyllanthus emblica) fruits were obtained from the local market of Lahore, Pakistanand duly authenticated by Department of Botany, University of the Punjab, Lahore. They were then dried in the shade and carefully grinded to a coarse sizepowder by electrical grinder and then the powder was passed through 24 mesh size sieve. The powder of Amla fruit (500 g) was then allowed to dip in 70% aqueous ethanolsolution for three days with occasional stirring and mixture was drained first with muslin cloth and after that it was strained by Whatmann filter paper in order to achieve the extract in the fine form. The filtrate was then concentrated by using rotary evaporator and finally a thick viscous paste of dark brown color was obtained. It was then weighed to obtain percentage vield of the extract and extract was then stored in an air lockcontainer and was appropriately labeled. Container was placed in a refrigerator at low 4°C.

### Phytochemical analysis

Many kinds of secondary metabolites or constituents like tannins, alkaloids, phenols, flavonoids, nitrogen compounds, vitamins, terpenoids, steroids present in plant extract wereanalyzed by using simple qualitative tests (Mushtaq *et al.*, 2013).

#### Quantitative analysis of phenols in plant extract

Folin-Ciocalteu reagent method Kumaran & Karunakaran, (2007) with some modifications was used toassess the quantity of phenol in the aqueous crude extract of Phyllanthus emblica. In 1ml of plantextract 2.5ml of 10% Folin-Ciocalteu reagent and 2ml of 2% solution Na2CO3 were added. The resulting mixture was allowed to incubate for 15 minutes at room temperature .The sampleabsorbance was determined at 765nm. Gallic acid was used as standard in concentration of 1mg/ml.The tests were performed in triplicates and results of the test were determined by thestandard curve and were mentioned as gallic acid equivalent (mg/g of extracted compound).Total phenols = Gallic acid Eq.  $(\mu q/mL) \times extract (vol.) /$ sample (g)

# Quantitative analysis of flavonoids in plant extract

To determine the total flavonoid content, a method by (Kumaran & Karunakaran, 2007)was used. Toa 10 ml test tube, 0.3 ml of extract, 3.4 ml of 30% methanol, 0.15 ml of NaNO<sub>2</sub> (0.5 M) and

0.15 ml of AlCl<sub>3</sub>.6H<sub>2</sub>O(0.3 M aluminum chloride) were combined and mixed. 1 ml of NaOH (1 M) solution was added after 5 mins. The resulting solution was mixed well and the absorbance of the solution was measured against the reagent blank at 506 nm. The standard curve for total flavonoids was made by using rutin as standard solution (0 to 100 mg/l). The total flavonoidswere expressed as milligrams of rutin equivalents per g of dried fraction.

# Antioxidant activity

Plant extract was dissolved in DMSO and stock solution was also prepared. The stock solution (1mg/ml) was taken to prepare different concentration. То measure the free radicalscavenging activity DPPH method was used. Different concentrations of the crude extract(0.02, 0.04, 0.08, 0.16, 0.32, 0.64, 1.28 and 2.56 mg/ml) were prepared in DMSO and a solution of 0.01 mM of DPPH in DMSO was also prepared. 1mL of DPPH solution was addedin each different concentration. The control was also prepared by addition of DPPH solution in3mL of DMSO. The test tubes were kept in darkness for 30 min at room temperature. Absorbance was measured at 517nm against blank. Different concentrations of Rutin was also prepared which was used as a standard(Blois, 1958). Each sample was tested three times. Graph was plotted between% scavenging activity and concentration by which the value of IC50 was calculated. DPPH radical scavenging activity was measured by following formula.% age scavenging = [Absorbance of control - Absorbance of sample/ Absorbance of control] x 100.

# Animals

Male albino rats weighing 200-300 g were used in this study which were procured and housed in the animal house of Riphah Institute of Pharmaceutical Sciences, Riphah International University Lahore. The rats were kept under standardroom temperature of 25±2 °C and a relative humidity of 45-55% and supplied with a light and dark cycle of 12:12 hr. The rats were placed in a hygienic environment during the entire study Before initiating the study the animals were acclimatized at least for one week of work. The experimental study design was provided as per the ethical considerations for animal management and handling. All the animals had free access to water and they were fed with commercially available rat food pellet.

### **Study Design**

Total of 36 rats were equally divided into six groups in such a way that G-1 was administered with normal food and water. G-II was diabetic control group administered with dexamethasone 10 mg/kg/i.p once daily. G-III to VI were administered with dexamethasone 10 mg/kg/i.p + glimiperide 20 dexamethasone mg/kg/i.p. 10 mg/kg/i.p+ Phyllanthus emblica 40 mg/kg/i.p, dexamethasone 10 mg/kg/i.p+ Phyllanthus emblica 60 mg/kg/i.p and dexamethasone 10 mg/kg/i.p+ Phyllanthus emblica respectively for eight days. 80 mg/kg/i.p. Hyperglycemia was induced in all the groups except normal control group by administration of dexamethasone 10 mg/kg/i.p daily for seven days and then later on animals were treated according to the protocols of study design for eight consecutive days and blood glucose was measured at 0 hr, 2 hr, 4 hr and 8 hr after dosing of animals. Blood was drawn from tail of rat and glucose level was measured by using glucometer. Similarly, weight of animals before and after the treatment was noted for each animal.

#### **Statistics Tools**

All the statistical data /results were assessed by using Graph Pad Prism 6. TWO WAYANOVA was applied and the values /data were presented in tabular form as a Mean  $\pm$  standarderror of mean. The results were summarized by comparing the experimental groups to disease groups and the normal group. The level of significance was shown in term of P-value (P  $\leq$  0.05)

#### RESULTS

# **Phytochemical Analysis**

Qualitative phytochemical analysis revealed the presence of carbohydrates, tannins, phenols, alkaloids, flavonoids, saponins, glycosides, amino acids and proteins in crude extract of *Phyllanthus emblica* as shown in Table I.

#### **Total Phenols and Flavonoids**

Total phenolic and flavonoids contents of ethanolic extracts of *Phylanthus emblica* were recorded as 9.9 mg/g equivalent of gallic acid and 100 mg/g equivalent of rutin, respectively.

## Free Radical Scavenging Activity

The percentage of DPPH free radical scavenging activity of the ethanolic extracts of *Phyllanthus emblica* indicated that by increasing the concentration, scavenging activity was also increased against Rutin. As IC<sub>50</sub> is increased, scavenging activity is decreased and vice versa.

# Effect of Ethanolic Extract of *Phyllanthus Emblica* on Blood Glucose Level

It was observed that high blood glucose levels were recorded in disease controlled animals in which diabetes was induced as compared to normal control group. The blood glucose level was observed to be decreased significantly in animals treated with ethanolic extract of *Phyllanthus emblica* and the reference drug glimepiride when compared with the disease control. Moreover, it was observed that on day 1<sup>st</sup> and 2<sup>nd</sup>, least reduction in glucose levels was observed at various time intervals. But the effect of different doses of plant extract was clearly observed on day 4<sup>th</sup>, 6<sup>th</sup> and 8<sup>th</sup> as shown in Table III.

Chemical Constituents	Tests	Presence
Protein and amino acids	i. Millon's test	+++
	ii. Ninhydrin	++
	i. Fehling Test	-
Carbohydrates	ii. Benedict test	++
	iii. Molisch's Test	++
	iv. Iodine Test	+
Tannins and phenols	i. Iodine test	-
	ii. Ferric chloride test	-
	iii. Nitric acid test	+
Flavonoids	i. Alkaline Reagent	++
	ii. Lead acetate	++
Saponins	Foam Test	++
	i. Liebermann's Test	-
Glycosides	ii. Salkowski's Test	++
	iii. Keller-kilani Test	+
Steroids	Test for Steroids	++
	i. Hager's Test	++
Alkaloids	ii. Wagner Test	+++
	iii. Mayer's Test	++
	iv. Dragendroff's Test	++
Terpenoids	Test for Terpenoids	-

+ sign indicates slightly present, ++ indicates moderately present, +++ represents highly present and - sign indicates absence

Table II: DPPH free radical	scavenging activity	of ethanolic	extract of	Phyllanthus e	mblica and
Rutin					

Sr. No.	Concentration mg/ml	% age scavenging by Rutin (%)	% age scavenging by Phyllanthus emblica (%)
1	0.02	14±0.66	58±1.155*
2	0.04	17±1.14	60±1.155*
3	0.08	18±1.16	61±1.155*
4	0.16	35±1.12	65±1.155*
5	0.32	40±1.11	66±1.155*
6	0.64	43±1.21	68±0.577*
7	1.28	48±1.24	67±1.155*
8	2.56	58±1.15	68±0.33*

Each Value was expressed as Mean ± SEM. n=3. \*P<0.05 was given as compared to the value of Rutin

Groups					Blucose Levels I) on Day 2 <sup>nd</sup>			Blood Glucose Levels (mg/dl) on Day 4 <sup>th</sup>			Blood Glucose Levels (mg/dl) on Day 6 <sup>th</sup>			Blood Glucose Levels (mg/dl) on Day 8 <sup>th</sup>						
	0 h	2 h	4 h	8h	0 h	2 h	4 h	8h	0 h	2 h	4 h	8h	0 h	2 h	4 h	8h	0 h	2 h	4 h	8h
G-I	99	100	101	99	99	99	100	98	99	100	99	100	98	103	104 ±	99.6	101	99	99.33	101
	±	±	±	±	±	±	±	±	±	±	±	±	±	±	0.5	±	±	±	± 0.4	±
	0.5	0.5	0.5	0.9	0.5	0.7	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.6		0.4	0.5	0.7		0.8
G-II	263	268	271	283	285	289	293	301	315	318	321	328	341	342	346.3	355	367	369	373 ±	380
	±	±	±	±	±	±	±	±	±	±	±	±	±	±	± 0.4	±	±	±	0.5	±
	0.9	0.5	0.5	0.7	0.7	0.7	0.7	0.5	0.7	0.5	0.5	0.5	1.3	1.2		1.1	0.5	0.7		0.7
G-III	281	278	271	263	285	281	278	267	248	246	240	232	215	209	207.1	210	178	171	163 ±	155
	±	±	±	±	±	±	±	±	±	±	±	±	±	±	± 0.6*	±	±	±	0.5*	±
	0.5*	0.5*	0.5	0.5*	0.5	0.7*	0.5*	0.5*	0.5*	0.5*	0.5*	0.5*	0.5*	0.4*		1.7*	0.5*	0.7*		1.1*
G-IV	298	291	287	278	283	278	271	261	260	257	250	237	221	218	216.6	207	189	181	178 ±	171
	±	±	±	±		±	±	±	±	±	±	±	±	±	± 0.7*	±	±	±	0.7*	±
	0.5*	0.6*	0.7*	0.7*	0.5	0.7*	0.7*	0.7*	0.5*	0.5*	0.5*	0.5*	1.1*	0.3*		0.5*	0.7*	1.6*		0.7*
G-V	299	295	291	283	301	298	291	275	251	248	241	230	219	216	211 ±	201	186	180	176 ±	168
		±		±		±	±				±	±	±	±	0.7*	±	±	±	0.5*	±
	0.7*	0.6*	0.5*	0.5	0.7*	0.5*	0.7	0.5*	0.5*	0.5*	0.9*	1.3*	0.3*	1.5*		0.5*	0.7*	1.1*		0.7*
	~~-								0.40		~~~					100	10-	101		100
G-VI	297	293	286	269	299	291	287	274	249	244	237	226	213	209	207 ±	199	185	181	176 ±	166
	±	±	±	±	±	±	±	±	±	±	±	±	±	±	0.7*	±	±	±	1.1*	±
	0.7*	0.6*	0.5*	0.5*	0.6	0.5	0.7*	0.5*	1.5*	1.0*	0.8*	0.9*	1.2*	1.1*		0.5*	1.9*	0.7*		0.7*

Table III: Effect of ethanolic extract of *Phyllanthus emblica* on blood glucose level at various time intervals

Two way ANOVA was applied and the values /data were presented in tabular form as a Mean  $\pm$  standard error of mean. The results were summarized by comparing the experimental groups to disease groups and the normal group. The level of significance was shown in term of P-value (P  $\leq$  0.05)

# Effect of Extract on Body Weight of Diabetic Rats

To determine the effect of ethanolic extract of *Phyllanthus emblica* on body weight of diabetic rats, it was compared with standard drug glimepiride. There was a significant reduction in body weight in dexamethasone treated rats when compared to normal control.

Treatment with the standard drug and ethanolic extract of *Phyllanthus emblica* in the dose 80 mg/Kg significantly prevented the loss in body weight. However *Phyllanthus emblica* in dose of 40 mg/Kg and 60 mg/Kg also prevented loss in body weight but the effect was dose dependent (Table IV).

Treatment Groups	Before Treatment Mean±SEM	After Treatment Mean ±SEM
Normal Control	215.66 ± 0.49	216.10 ± 0.71
Disease Control	248.76 ± 0.55	168.37 ± 0.83
Standard Control	274.50 ± 0.42	241.81 ± 0.94
Phyllanthus emblica 40mg/kg	264.93 ± 0.66	190.33 ± 0.64
Phyllanthus emblica 60mg/kg	271.69 ± 0.49	216.23 ± 0.91
Phyllanthus emblica 80mg/kg	310.71 ± 0.57	274.11 ± 0.97

# DISCUSSION

The current study showed the presence of medicinally active constituents in fruit extract of *Phyllanthus emblica*. The qualitative analysis revealed the presence of proteins and amino acids, carbohydrates, flavonoids, saponins, alkaloids, glycosides and steroids. Tannins and phenols were present but to a much lesser extent, whereas the quantitative analysis showed the percentage of the crude chemical constituents that exhibited the therapeutic and physiological activity.

Polyphenolic compounds found in a variety of medicinal plants are responsible for antidiabetic activity and cardio protective effects (Subramanian *et al.*, 1983). Moreover, it has been investigated that flavonoids are also responsible for hypoglycemic effects and they raise the levels of hemoglobin in rats (Sri *et al.*, 2013).

The fruit extract of *Phyllanthus emblica* possessed significant hypoglycemic activity and also it might have improved insulin resistance through enhanced insulin sensitivity in the peripheral tissues as was evident from decreased glucose levels. The tannins present in plant extractsare potential inhibitors of aldose reductase (AR) and they further oppose the pathways induced by oxidative stress as it was seen reversal of changes with reference to lipid

peroxidation and protein carbonyl content and activities of antioxidant enzymes. *Phyllanthus emblica* also averted the accumulation of lens proteins which are triggered by hyperglycemia (Suryanarayana *et al.*, 2007). Aldose reductase shows a role in the development and progress of secondary complication of diabetes that also includes cataract(Daisy *et al.*, 2005), thus *Phyllanthus emblica* is supposed to prevent the levels of aldose reductase and protect the body from harmful effects of aldose in diabetic rats.

Findings further suggested thatdue to potent antioxidant activity, the plant extract is highly efficacious in controlling diabetes. The ability of tannins to augment glucose uptake and prevent adipogenesis, makes it potent remedy in treating non-insulin dependent diabetes mellitus (Kim *et al.*, 2010). It has also been stated that insulin obtained with chromium is proficient enough to reverse blood sugar, serum cholesterol and phospholipids levels to those of normal rats (Arts *et al.*, 2000).

#### CONCLUSION

It is concluded that ethanolic fruit extract of *Phyllanthus emblica* not only improves diabetes mellitus by increasing insulin sensitivity towards peripheral tissues but also reduces hepatic glucose outputs.

- Arts, I. C., Van De Putte, B. & Hollman, P.C., (2000). Catechin contents of foods commonly consumed in The Netherlands. 1. Fruits, vegetables, staple foods, and processed foods. *J. Agr. Food Chem.*,**48(5)**: 1746-1751.
- Blois, M. S., (1958). Antioxidant determinations by the use of a stable free radical. *Nature*,**181**(4617): 1199-1200.
- Daisy, P., Averal, H. & Modilal, R.D., (2005). Curative properties of Phyllanthus extracts in alloxan diabetic rats. *J. Trop. Med. Plant.*, **5**: 21-27.
- Dang, G., Parekar, R., Kamat, S., Scindia, A. & Rege, N., (2011). Antiinflammatory of activity Phyllanthus emblica, Plumbago zevlanica and Cyperus rotundus in acute models of inflammation. Phytother. Res.,25(6): 904-908.
- Jung, M., Park, M., Lee, H.C., Kang, Y.-H., Kang, E. S. & Kim, S. K., (2006). Antidiabetic agents from medicinal plants. *Curr. Med. Chem.*,**13(10)**: 1203-1218.
- Kim, H. Y., Okubo, T., Juneja, L.R. & Yokozawa, T., (2010). The protective role of amla (*Emblica officinalis Gaertn.*) against fructose-induced metabolic syndrome in a rat model. *Brit. J. Nutr.*,**103(04)**: 502-512.
- Kumaran, A. & Karunakaran, R.J., (2006). Nitric oxide radical scavenging active components from *Phyllanthus emblica L. Plant Food. Hum. Nutr.*,**61(1)**: 1-10.
- Kumaran, A. & Karunakaran, R.J., (2007). In vitro antioxidant activities of methanol extracts of five Phyllanthus species from India. *LWT-Food Sci. Technol.*,**40(2)**: 344-352.
- Mushtaq, A., Mahmood, A. & Jabeen, Q., (2013). Hepatoprotective action of a polyherbal aqueous ethanolic extract against nimesulide intoxicated albino rats. *IJPRBS*.2(6): 332-347.
- Naik, G., Priyadarsini, K., Bhagirathi, R., Mishra, B., Mishra, K., Banavalikar, M. & Mohan, H., (2005). In vitro antioxidant studies and free radical reactions of

triphala, an ayurvedic formulation and its constituents. *Phytother. Res.*,**19(7)**: 582-586.

- Nisha, P., Singhal, R. S. & Pandit, A. B., (2004). A study on degradation kinetics of ascorbic acid in amla (*Phyllanthus emblica* L.) during cooking. *Int. J. Food Sci. Nutr.*,**55(5)**: 415-422.
- Shankar, U., Murali, K., Shaanker, R.U., Ganeshaiah, K. & Bawa, K., (1996). Extraction of non-timber forest products in the forests of Biligiri Rangan Hills, India. 3. Productivity, extraction and prospects of sustainable harvest of Amla *Phyllanthus emblica*,(Euphorbiaceae). *Econ Bot.*,**50(3)**: 270-279.
- Sri, K. S., Kumari, D. J. & Sivannarayana, G., (2013). Effect of Amla, an approach towards the control of diabetes mellitus. *Int. J. Curr. Microbiol. Appl. Sci.*2(9): 103-108.
- Subramanian, V., Butler, L. G., Jambunathan, R. & Rao, K. P., (1983). Some agronomic and biochemical characters of brown sorghums and their possible role in bird resistance. *J. Agr. Food Chem.*,**31(6)**: 1303-1307.
- Suryanarayana, P., Saraswat, M., Petrash, J. M. & Reddy, G. B., (2007). Emblica officinalis and its enriched tannoids delay streptozotocin-induced diabetic cataract in rats. *Mol. Vis.*, **13**: 1291-1297.
- Tsay, H.-S. & Agrawal, D. C., (2005). Tissue culture technology of Chinese medicinal plant resources in Taiwan and their sustainable utilization. *Int. J. App. Sci. Eng.***3**: 215-223.
- Unander, D. W., Webster, G. L. & Blumberg, B. S., (1990). Records of usage or assays in Phyllanthus (Euphorbiaceae) I. subgenera Isocladus, Kirganelia, Cicca and Emblica. *J. Ethnopharmacol.*,**30(3)**: 233-264.
- Zhang, Y.-J., Tanaka, T., Iwamoto, Y., Yang, C.-R. & Kouno, I., (2001). Novel Sesquiterpenoids from the Roots of *Phyllanthus emblica. J. Nat. Prod.*,**64(7)**: 870-873.